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APPLICATION FOR UNITED STATES LETTERS PATENT

INVENTORS:

Luca Rastelli
Mary Gerritsen

TITLE:

ANGIOGENESIS ASSOCIATED
PROTEINS AND NUCLEIC ACIDS
ENCODING THE SAME

ATTORNEY:

Paul E. Rauch, Ph.D.
BRINKS HOFER GILSON & LIONE
P.O. BOX 10395
CHICAGO, ILLINOIS 60610
(312) 321-4200

ANGIOGENESIS ASSOCIATES PROTEINS, AND NUCLEIC ACIDS
ENCODING THE SAME

RELATED APPLICATIONS

This application claims priority to U.S. provisional application Serial No. 60/191,134 filed 03/22/2000, which is incorporated herein by reference in its entirety.

BACKGROUND

Cities have roads and alleys, plants have xylem and phloem, and people have arteries, veins and lymphatics. Without these byways, the vertebrate animal cells would starve or drown in their metabolic refuse. Not only do blood vessels deliver food and oxygen and carry away metabolic wastes, but they also transport signaling substances that apprise cells of situations remote to them but to which they need to respond. Hormonal messages are a common signal.

All blood vessels are ensheathed by a basal lamina and a delicate monolayer of remarkably plastic endothelial cells lining the luminal walls. Depending on location and function, smooth muscle and connective tissue may also be present.

Not only do healthy cells depend on the blood resources transported by the circulatory system, but so, too, unwanted cells: tumorigenic and malignant cells. These cells colonize and proliferate if they are able to divert blood resources to themselves. Angiogenesis, the type of blood vessel formation where new vessels emerge from the proliferation of preexisting vessels (Risau, 1995; Risau and Flamme, 1995), is exploited not only by usual processes, such as in wound healing or myocardial infarction repair, but also by tumors themselves and in cancers, diabetic retinopathy, macular degeneration, psoriasis, and rheumatoid arthritis. Regardless of the process, whether pathological or usual physiological, endothelial cells mediate angiogenesis in a multi-step fashion: (1) endothelia receive an extracellular cue, (2) the signaled cells breach the basal lamina sheath, abetted by proteases they secrete, (3) the cells then migrate to the signal and proliferate, and finally, (4) the cells form a tube, a morphogenic event (Alberts *et al.*, 1994). The complexity of this process indicates complex changes in cellular physiology and morphology, gene expression, and signaling. Angiogenic accomplices that are cues

include basic fibroblast growth factors (bFGF), angiopoietins (such as ANG1) and various forms of vascular endothelial growth factor (VEGF).

The molecular events and the order in which they occur and the pathways that are required for this process are of fundamental importance to understand angiogenesis. *In vitro* models are useful for identifying alterations in gene expression that occur during angiogenesis. A particularly fruitful model systems involves the suspension in a three-dimensional type I collagen gel and various stimuli, such as phorbol myristate acetate (PMA), basic fibroblast growth factor (bFGF), and VEGF. The combination of the stimuli and the collagen gel results in the formation of a three-dimensional tubular network of endothelial cells with interconnecting luminal structures. In this model, endothelial differentiation into tubelike structures is completely blocked by inhibitors of new mRNA or protein synthesis. Furthermore, the cells progress through differentiation in a coordinated and synchronized manner, thus optimizing the profile of gene expression (Kahn *et al.*, 2000; Yang *et al.*, 1999).

Tumor cells exploit angiogenesis to facilitate tumor growth. Controlling angiogenesis, by controlling the activity or expression of genes and proteins associated with angiogenesis, provides a way to prevent tumor growth, or even destroy tumors.

SUMMARY

The invention is based in part upon the discovery of novel nucleic acid sequences encoding novel polypeptides. Nucleic acids encoding the polypeptides disclosed in the invention, and derivatives and fragments thereof, will hereinafter be collectively designated as "AAP" nucleic acid or polypeptide sequences. AAP, or angiogenesis associated polypeptides (AAP) comprises kelch-like polypeptide (KLP), human ortholog of mouse BAZF (hBAZF), hmt-elongation factor G (hEF-G), human ortholog of rat TRG (hTRG), human myosin X (hMX1) and its splice variant (hMX2), nuclear hormone receptor (NHR), and human mitochondrial protein (hMP).

The invention is based in part upon the discovery of novel nucleic acid sequences encoding novel polypeptides. Nucleic acids encoding the polypeptides disclosed in the invention, and derivatives and fragments thereof, will hereinafter be collectively designated as "AAP" nucleic acid or polypeptide sequences."

In a first aspect, the present invention is an isolated polypeptide having at least 80% sequence identity to the sequence SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16, polynucleotides encoding the same, and antibodies that specifically bind the same.

In a second aspect, the present invention is an isolated polynucleotide having at least 80% sequence identity to the sequence SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15, or a complement thereof.

In a third aspect, the present invention is a transgenic non-human animal, having a disrupted *AAP* gene or a transgenic non-human animal expressing an exogenous polynucleotide having at least 80% sequence identity to the sequence SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15, or a complement of said polynucleotide.

In a fourth aspect, the present invention is a method of screening a sample for a mutation in an *AAP* gene.

In a fifth aspect, the present invention is a method of modulating angiogenesis comprising modulating the activity of at least one *AAP* polypeptide.

In a sixth aspect, the present invention is a method of increasing, as well as decreasing angiogenesis, comprising modulating the activity of at least one *AAP* polypeptide. Activity modulation of *AAP* polypeptides may be over-expressing or eliminating expression of the gene, or impairing a *AAP* polypeptide's function by contact with specific antagonists or agonists, such as antibodies or aptamers.

In a seventh aspect, the present invention is a method of treating various pathologies, including tumors, cancers, myocardial infarctions and the like.

In an eighth aspect, the present invention is a method of measuring a *AAP* transcriptional and translational up-regulation or down-regulation activity of a compound.

In a ninth aspect, the invention is a method of screening a tissue sample for tumorigenic potential.

In a tenth aspect, the invention is a method of determining the clinical stage of tumor which compares the expression of at least one *AAP* in a sample with expression of said at least one gene in control samples.

Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. All publications, patent applications, patents, and other

references mentioned herein are incorporated by reference in their entirety. In the case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

DETAILED DESCRIPTION

A model of angiogenesis-the suspension of endothelial cells in type I collagen gels with various stimuli-was used to identify a molecular fingerprint or transcriptional profile of endothelial differentiation into tubelike structures, using amplification and an imaging approach called GeneCalling (Shimkets *et al.*, 1999). This method was previously shown to provide a comprehensive sampling of cDNA populations in conjunction with the sensitive detection of quantitative differences in mRNA abundance for both known and novel genes. Many differentially expressed cDNA fragments were found. The identification and differential expression of these genes was confirmed by a second independent method employing real-time quantitative polymerase chain reaction (PCR). Although some of the identified cDNA fragments were genes known to play some role in angiogenesis, many other differentially expressed genes were unexpected. The inventors have identified the unexpected genes and polypeptides that are expressed in response to this model of angiogenesis, collectively referred to as angiogenesis associated polypeptides (AAP). AAP are kelch-like polypeptide (KLP), human ortholog of mouse BAZF (hBAZF), hmt-elongation factor G (hEF-G), human ortholog of rat TRG (hTRG), human myosin X (hMX1) and its splice variant (hMX2), nuclear hormone receptor (NHR), and human mitochondrial protein (hMP).

Definitions

Unless defined otherwise, all technical and scientific terms have the same meaning as is commonly understood by one of skill in the art to which this invention belongs. The definitions below are presented for clarity. All patents and publications referred to herein are, unless noted otherwise, incorporated by reference in their entirety.

The recommendations of (Demerec *et al.*, 1966) where these are relevant to genetics are adapted herein. To distinguish between genes (and related nucleic acids) and the proteins that they encode, the abbreviations for genes are indicated by *italicized* (or

underlined) text while abbreviations for the proteins start with a capital letter and are not italicized. Thus, *AAP* or AAP refers to the nucleotide sequence that encodes AAP.

“Isolated,” when referred to a molecule, refers to a molecule that has been identified and separated and/or recovered from a component of its natural environment. Contaminant components of its natural environment are materials that interfere with diagnostic or therapeutic use.

“Container” is used broadly to mean any receptacle for holding material or reagent. Containers may be fabricated of glass, plastic, ceramic, metal, or any other material that can hold reagents. Acceptable materials will not react adversely with the contents.

1. *Nucleic acid-related definitions*

(a) *control sequences*

Control sequences are DNA sequences that enable the expression of an operably-linked coding sequence in a particular host organism. Prokaryotic control sequences include promoters, operator sequences, and ribosome binding sites. Eukaryotic cells utilize promoters, polyadenylation signals, and enhancers.

(b) *operably-linked*

Nucleic acid is operably-linked when it is placed into a functional relationship with another nucleic acid sequence. For example, a promoter or enhancer is operably-linked to a coding sequence if it affects the transcription of the sequence, or a ribosome-binding site is operably-linked to a coding sequence if positioned to facilitate translation. Generally, “operably-linked” means that the DNA sequences being linked are contiguous, and, in the case of a secretory leader, contiguous and in reading phase. However, enhancers do not have to be contiguous. Linking is accomplished by conventional recombinant DNA methods.

(c) *isolated nucleic acids*

An isolated nucleic acid molecule is purified from the setting in which it is found in nature and is separated from at least one contaminant nucleic acid molecule. Isolated *AAP* molecules are distinguished from the specific *AAP* molecule, as it exists in cells. However, an isolated *AAP* molecule includes *AAP* molecules contained in cells that

ordinarily express an AAP where, for example, the nucleic acid molecule is in a chromosomal location different from that of natural cells.

2. *Protein-related definitions*

(a) *purified polypeptide*

When the molecule is a purified polypeptide, the polypeptide will be purified (1) to obtain at least 15 residues of N-terminal or internal amino acid sequence using a sequenator, or (2) to homogeneity by SDS-PAGE under non-reducing or reducing conditions using Coomassie blue or silver stain. Isolated polypeptides include those expressed heterologously in genetically-engineered cells or expressed *in vitro*, since at least one component of an AAP natural environment will not be present. Ordinarily, isolated polypeptides are prepared by at least one purification step.

(b) *active polypeptide*

An active AAP or AAP fragment retains a biological and/or an immunological activity of the native or naturally-occurring AAP. Immunological activity refers to the ability to induce the production of an antibody against an antigenic epitope possessed by a native AAP; biological activity refers to a function, either inhibitory or stimulatory, caused by a native AAP that excludes immunological activity. A biological activity of AAP includes, for example, modulating angiogenesis.

(c) *Abs*

Antibody may be single anti-AAP monoclonal Abs (including agonist, antagonist, and neutralizing Abs), anti-AAP antibody compositions with polyepitopic specificity, single chain anti-AAP Abs, and fragments of anti-AAP Abs. A “monoclonal antibody” refers to an antibody obtained from a population of substantially homogeneous Abs, *i.e.*, the individual Abs comprising the population are identical except for naturally-occurring mutations that may be present in minor amounts

(d) *epitope tags*

An epitope tagged polypeptide refers to a chimeric polypeptide fused to a “tag polypeptide”. Such tags provide epitopes against which Abs can be made or are available, but do not interfere with polypeptide activity. To reduce anti-tag antibody reactivity with endogenous epitopes, the tag polypeptide is preferably unique. Suitable tag polypeptides generally have at least six amino acid residues and usually between about 8

and 50 amino acid residues, preferably between 8 and 20 amino acid residues). Examples of epitope tag sequences include HA from *Influenza A* virus and FLAG.

The novel AAP of the invention include the nucleic acids whose sequences are provided in Tables 1, 3, 5, 7, 9, 11, 13 and 15, or a fragment thereof. The invention also includes a mutant or variant AAP, any of whose bases may be changed from the corresponding base shown in Tables 1, 3, 5, 7, 9, 11, 13 and 15 while still encoding a protein that maintains the activities and physiological functions of the AAP fragment, or a fragment of such a nucleic acid. The invention further includes nucleic acids whose sequences are complementary to those just described, including complementary nucleic acid fragments. The invention additionally includes nucleic acids or nucleic acid fragments, or complements thereto, whose structures include chemical modifications. Such modifications include, by way of nonlimiting example, modified bases, and nucleic acids whose sugar phosphate backbones are modified or derivatized. These modifications are carried out at least in part to enhance the chemical stability of the modified nucleic acid, such that they may be used, for example, as anti-sense binding nucleic acids in therapeutic applications in a subject. In the mutant or variant nucleic acids, and their complements, up to 20% or more of the bases may be so changed.

The novel AAP of the invention include the protein fragments whose sequences are provided in Tables 2, 4, 6, 8, 10, 12, 14 and 16. The invention also includes an AAP mutant or variant protein, any of whose residues may be changed from the corresponding residue shown in Tables 2, 4, 6, 8, 10, 12, 14 and 16 while still encoding a protein that maintains its native activities and physiological functions, or a functional fragment thereof. In the mutant or variant AAP, up to 20% or more of the residues may be so changed. The invention further encompasses Abs and antibody fragments, such as F_{ab} or (F_{ab})'₂, that bind immunospecifically to any of the AAP of the invention.

The AAP nucleic acids are shown in Tables 1, 3, 5, 7, 9, 11, 13 and 15, and the corresponding polypeptides are shown in Tables 2, 4, 6, 8, 10, 12, 14 and 16, respectively. Start and stop codons in the polynucleotide sequences are indicated in **boldface** and with underlining. SEQ ID NO:3 lacks a stop codon. The sequences of *hMX1* and *hMX2* do not have start codons (see Table 17); consequently, hMX1 and hMX2 polypeptides do not start with a Met. For any lacking polynucleotide sequence, one of skill in the art may

retrieve the full length sequence by, for example, probing cDNA or genomic libraries with probes designed according to the sequences of the instant invention.

Table 1 KLP nucleotide sequence (SEQ ID NO:1)

ctggcctaga	tactacaact	gaactttttt	tcttttttagt	tactccacag	gatccgctga	60
acataggatg	ttgccacaaa	atctacctcg	tgtatttttc	tctttcactc	atgagctgca	120
caattgcaga	tttgagcaca	atgtctgcag	actgtgttga	aaaactctga	agaacctaat	180
taacacagga	tgacctagga	gtgattctaa	gtctgtgtaa	caagatatta	ctcatttagtg	240
aatgtgtcag	tcttgggtact	gaatgctgca	gataacagca	agtaggttct	cctttatttct	300
tgaagtattc	acttgacctt	ccatcagtaa	gacggacttt	tctaactctgt	tcttgagat	360
attaatggaa	tacagtc atg	tccactcaag	acgagaggca	gatcaatact	gaatatgctg	420
tgtcattggt	ggaacagtgt	aaactgtttt	atgaacagca	gttgtttact	gacatagtgt	480
taattgttga	gggcactgaa	ttcccttgct	ataagatggg	tcttgcaaca	tgtagctctt	540
atttcagggc	catgtttatg	agtggactaa	gtgaaagcaa	acaaacccat	gtacacctga	600
ggaatgtcga	tgctgccacc	ttacagataa	taataactta	tgcatacacg	ggtaacttgg	660
caatgaatga	cagcactgta	gaacagcttt	atgaaacagc	ttgcttccta	caggtagaag	720
atgtgttaca	acgttgtcga	gaatatTTAA	ttaaaaaat	aatgacagag	aattgtgtac	780
gattgttgag	ttttgctgat	ctcttcagtt	gtgaggaatt	aaaacagagt	gctaaaagaa	840
tgggtggagca	caagttcact	gctgtgtatc	atcaggacgc	gttcatgcag	ctgtcacatg	900
acctactgat	agatattctc	agtagtgaca	atttaaagt	agaaaaggaa	gaaaccgttc	960
gagaagctgc	tatgctgtgg	ctagagtata	acacagaatc	acgatcccag	tatttgtctt	1020
ctgttcttag	ccaaatcaga	attgatgcac	tttcagaagt	aacacagaga	gcttggtttc	1080
aaggctctgcc	acccaatgat	aagtcagtgg	tggttcaagg	tctgtataag	tccatgcca	1140
agtttttcaa	accaagactt	gggatgacta	aagaggaaat	gatgattttc	attgaagcat	1200
cttcagaaaa	tcctttagt	ctttactctt	ctgtctgtta	cagcccccaa	gcagaaaaag	1260
tttacaagtt	atgtagccca	ccagctgatt	tgcataaggt	tgggaccgtt	gtaactcctg	1320
ataatgatat	ctacatagca	gggggtcaag	ttcctctgaa	aaacacaaaa	acaaatcaca	1380
gtaaaacaag	caaacttcag	actgccttca	gaactgtgaa	ttgcttttat	tggtttgatg	1440
cacagcaaaa	tacctggttt	ccaagagacc	caatgctttt	tgtccgcata	aagccatctt	1500
tggtttgctg	tgaaggctat	atctatgcaa	ttggaggaga	tagcgtaggt	ggagaactta	1560
atcggaggac	cgtagaaaga	tacgacactg	agaaagatga	gtggacgatg	gtaagccctt	1620
taccttgtgc	ttggcaatgg	agtgcagcag	ttgtggttca	tgactgcatt	tatgtgatga	1680
cactgaacct	catgtactgt	tattttccaa	ggtctgactc	atgggtagaa	atggccatga	1740
gacagactag	taggtccttt	gcttcagctg	cagcttttgg	tgataaaatt	ttctatattg	1800
gaggggttga	tattgctacc	aattccggca	taagactccc	ctctggcact	gtagatgggt	1860
cttcagtaac	tgtggaaatt	tatgatgtga	ataaaaatga	gtggaaaatg	gcagccaaca	1920
tccctgctaa	gaggtactct	gacccctgtg	ttagagctgt	tgtgatctca	aattctctat	1980
gtgtgtttat	gcgagaaacc	cacttaaagt	agcgagctaa	atacgtcacc	taccaatatg	2040
acctggaact	tgaccggtgg	tctctgcggc	agcatatatc	tgaacgtgta	ctgtgggact	2100
tggggagaga	ttttcgaatc	actgtgggga	aactctatcc	atcctgcctt	gaagagtctc	2160
catggaaacc	accaacttat	cttttttcaa	cggatgggac	agaagagttt	gaactggatg	2220
gagaaatggg	tgcactacca	cctgtat ag	ggggaagtgc	agggagtgca	cgcttgagtt	2280

atgtgctttg tcattttctt tgctaaacaa aagaggctat gaaagaacta aatatgagta	2340
cataaaattc tatctttgat aaattttatt tttatgccct acttaatat tgcacagta	2400
taatatatat cagtgagtct tacagaaaga tatgcttcca taatatgaaa tagattattc	2460
aataattgag aaactttatg tgtaatcatg agagtataag aatctggatt atctaacatt	2520
gtagccctg tgtatgtaca gttcaaaaag ttcatttata aaagtagttt cctgttc	2577

Table 2 KLP polypeptide sequence (SEQ ID NO:2)

Met	Ser	Thr	Gln	Asp	Glu	Arg	Gln	Ile	Asn	Thr	Glu	Tyr	Ala	Val	Ser	
1				5					10					15		
Leu	Leu	Glu	Gln	Leu	Lys	Leu	Phe	Tyr	Glu	Gln	Gln	Leu	Phe	Thr	Asp	
			20					25					30			
Ile	Val	Leu	Ile	Val	Glu	Gly	Thr	Glu	Phe	Pro	Cys	His	Lys	Met	Val	
		35					40					45				
Leu	Ala	Thr	Cys	Ser	Ser	Tyr	Phe	Arg	Ala	Met	Phe	Met	Ser	Gly	Leu	
	50					55					60					
Ser	Glu	Ser	Lys	Gln	Thr	His	Val	His	Leu	Arg	Asn	Val	Asp	Ala	Ala	
65				70						75				80		
Thr	Leu	Gln	Ile	Ile	Ile	Thr	Tyr	Ala	Tyr	Thr	Gly	Asn	Leu	Ala	Met	
			85					90					95			
Asn	Asp	Ser	Thr	Val	Glu	Gln	Leu	Tyr	Glu	Thr	Ala	Cys	Phe	Leu	Gln	
			100					105					110			
Val	Glu	Asp	Val	Leu	Gln	Arg	Cys	Arg	Glu	Tyr	Leu	Ile	Lys	Lys	Ile	
		115					120					125				
Asn	Ala	Glu	Asn	Cys	Val	Arg	Leu	Leu	Ser	Phe	Ala	Asp	Leu	Phe	Ser	
	130				135					140						
Cys	Glu	Glu	Leu	Lys	Gln	Ser	Ala	Lys	Arg	Met	Val	Glu	His	Lys	Phe	
145				150					155						160	
Thr	Ala	Val	Tyr	His	Gln	Asp	Ala	Phe	Met	Gln	Leu	Ser	His	Asp	Leu	
			165					170						175		
Leu	Ile	Asp	Ile	Leu	Ser	Ser	Asp	Asn	Leu	Asn	Val	Glu	Lys	Glu	Glu	
		180					185					190				
Thr	Val	Arg	Glu	Ala	Ala	Met	Leu	Trp	Leu	Glu	Tyr	Asn	Thr	Glu	Ser	
	195					200						205				
Arg	Ser	Gln	Tyr	Leu	Ser	Ser	Val	Leu	Ser	Gln	Ile	Arg	Ile	Asp	Ala	
	210				215						220					
Leu	Ser	Glu	Val	Thr	Gln	Arg	Ala	Trp	Phe	Gln	Gly	Leu	Pro	Pro	Asn	
225				230					235						240	
Asp	Lys	Ser	Val	Val	Val	Gln	Gly	Leu	Tyr	Lys	Ser	Met	Pro	Lys	Phe	
			245					250					255			
Phe	Lys	Pro	Arg	Leu	Gly	Met	Thr	Lys	Glu	Glu	Met	Met	Ile	Phe	Ile	
		260					265						270			

Glu	Ala	Ser	Ser	Glu	Asn	Pro	Cys	Ser	Leu	Tyr	Ser	Ser	Val	Cys	Tyr	
		275					280					285				
Ser	Pro	Gln	Ala	Glu	Lys	Val	Tyr	Lys	Leu	Cys	Ser	Pro	Pro	Ala	Asp	
	290					295					300					
Leu	His	Lys	Val	Gly	Thr	Val	Val	Thr	Pro	Asp	Asn	Asp	Ile	Tyr	Ile	
305					310					315					320	
Ala	Gly	Gly	Gln	Val	Pro	Leu	Lys	Asn	Thr	Lys	Thr	Asn	His	Ser	Lys	
				325					330					335		
Thr	Ser	Lys	Leu	Gln	Thr	Ala	Phe	Arg	Thr	Val	Asn	Cys	Phe	Tyr	Trp	
			340					345					350			
Phe	Asp	Ala	Gln	Gln	Asn	Thr	Trp	Phe	Pro	Lys	Thr	Pro	Met	Leu	Phe	
		355					360					365				
Val	Arg	Ile	Lys	Pro	Ser	Leu	Val	Cys	Cys	Glu	Gly	Tyr	Ile	Tyr	Ala	
	370					375					380					
Ile	Gly	Gly	Asp	Ser	Val	Gly	Gly	Glu	Leu	Asn	Arg	Arg	Thr	Val	Glu	
385					390					395					400	
Arg	Tyr	Asp	Thr	Glu	Lys	Asp	Glu	Trp	Thr	Met	Val	Ser	Pro	Leu	Pro	
				405					410					415		
Cys	Ala	Trp	Gln	Trp	Ser	Ala	Ala	Val	Val	Val	His	Asp	Cys	Ile	Tyr	
			420					425					430			
Val	Met	Thr	Leu	Asn	Leu	Met	Tyr	Cys	Tyr	Phe	Pro	Arg	Ser	Asp	Ser	
		435					440					445				
Trp	Val	Glu	Met	Ala	Met	Arg	Gln	Thr	Ser	Arg	Ser	Phe	Ala	Ser	Ala	
	450					455					460					
Ala	Ala	Phe	Gly	Asp	Lys	Ile	Phe	Tyr	Ile	Gly	Gly	Leu	His	Ile	Ala	
465					470					475					480	
Thr	Asn	Ser	Gly	Ile	Arg	Leu	Pro	Ser	Gly	Thr	Val	Asp	Gly	Ser	Ser	
				485					490					495		
Val	Thr	Val	Glu	Ile	Tyr	Asp	Val	Asn	Lys	Asn	Glu	Trp	Lys	Met	Ala	
			500					505					510			
Ala	Asn	Ile	Pro	Ala	Lys	Arg	Tyr	Ser	Asp	Pro	Cys	Val	Arg	Ala	Val	
		515					520					525				
Val	Ile	Ser	Asn	Ser	Leu	Cys	Val	Phe	Met	Arg	Glu	Thr	His	Leu	Asn	
	530					535					540					
Glu	Arg	Ala	Lys	Tyr	Val	Thr	Tyr	Gln	Tyr	Asp	Leu	Glu	Leu	Asp	Arg	
545					550					555					560	
Trp	Ser	Leu	Arg	Gln	His	Ile	Ser	Glu	Arg	Val	Leu	Trp	Asp	Leu	Gly	
				565					570					575		
Arg	Asp	Phe	Arg	Cys	Thr	Val	Gly	Lys	Leu	Tyr	Pro	Ser	Cys	Leu	Glu	
			580					585					590			
Glu	Ser	Pro	Trp	Lys	Pro	Pro	Thr	Tyr	Leu	Phe	Ser	Thr	Asp	Gly	Thr	
		595					600					605				
Glu	Glu	Phe	Glu	Leu	Asp	Gly	Glu	Met	Val	Ala	Leu	Pro	Pro	Val		
	610					615					620					

Table 3 hBAZF nucleotide sequence (SEQ ID NO:3)

caagggagcg	aggggtgtcgt	agagggcaga	atgaacaaga	agaattagga	gggaggctgc	60
gtgtgccggg	gctaggggct	ggaagtcctg	gctctagttg	cacctcggaa	ggaaaaggca	120
aacagaggag	ggaaggcgtc	ttaggactgc	ctggatccag	agcactttcc	tcggcctcta	180
caggcctgtg	tcgctatggg	ttcccccgcc	gccccggagg	gagcgtggg	ctacgtccgc	240
gagttcactc	gccactcctc	cgacgtgctg	ggcaacctca	acgagctgcg	cctgcgcggg	300
atcctcactg	acgtcacgct	gctggttggc	gggcaacccc	tcagagcaca	caaggcagtt	360
ctcatcgctt	gcagtggcct	cttctattca	attttccggg	gccgtgcggg	agtcgggggtg	420
gacgtgctct	ctctgcccgg	gggtcccgaa	gcgagaggct	tcgcccctct	attggacttc	480
atgtacactt	cgcgccctgc	cctctctcca	gccactgcac	cagcagtcct	agcggccgcc	540
acctatttgc	agatggagca	cgtggtccag	gcatgccacc	gcttcatcca	ggccagctat	600
gaacctctgg	gcatctccct	gcgccccctg	gaagcagaac	ccccaacacc	cccaacggcc	660
cctccaccag	gtagtccag	gcgctccgaa	ggacaccag	accacctac	tgaatctcga	720
agctgcagtc	aaggccccc	cagtccagcc	agccctgacc	ccaaggcctg	caactggaaa	780
aagtacaagt	acatcgtgct	aaactctcag	gcctcccaag	cagggagcct	ggtcgggggag	840
agaagttctg	gtcaaccttg	cccccaagcc	aggctcccca	gtggagacga	ggcctccagc	900
agcagcagca	gcagcagcag	cagcagcagt	gaagaaggac	ccattccttg	tccccagagc	960
aggtctctct	caactgctgc	cactgtgcag	ttcaaagtgtg	gggctccagc	cagtaccccc	1020
tacctcctca	catcccaggc	tcaagacacc	tctggatcac	cctctgaacg	ggctcgtcca	1080
ctaccgggga	gtgaattttt	cagctgccag	aactgtgagg	ctgtggcagg	gtgctcatcg	1140
gggctggact	ccttggttcc	tggggacgaa	gacaaaccct	ataagtgtca	gctgtgccgg	1200
tcttcgttcc	gctacaaggg	caaccttgcc	agtcaccgta	cagtgcacac	aggggaaaag	1260
ccttaccact	gctcaatctg	cggagcccg	tttaaccggc	cagcaaacc	gaaaacgcac	1320
agccgcaccc	attcggggaga	gaagccgtat	aagtgtgaga	cgtgcggctc	gcgctttgta	1380
caggtaacga	gccagcctcc	aagtggcttc	caaggcaaac	ctgcaagagg	tgggggtgggc	1440
caaaagggag	ggttctgttc	ctcccagagg	caggacttga	agtctcctcc	ctcccagggtg	1500
gcacatctgc	gggcgacgt	gctgatccac	accggggaga	agccctaccc	ttgccctacc	1560
tgcggaaccc	gcttccgcca	cctgcagacc	ctcaagagcc	acgttcgcat	ccacaccgga	1620
gagaagcctt	accactgcga	cccctgtggc	ctgcatttcc	ggcacaagag	tcaactgcgg	1680
ctgcacatgc	gccagaaaca	cggagctgct	accaacacca	aagtgcacta	ccacattctc	1740
ggggggcccc						1749

Table 4 hBAZF polypeptide sequence (SEQ ID NO:4)

Met	Gly	Ser	Pro	Ala	Ala	Pro	Glu	Gly	Ala	Leu	Gly	Tyr	Val	Arg	Glu
1				5					10					15	
Phe	Thr	Arg	His	Ser	Ser	Asp	Val	Leu	Gly	Asn	Leu	Asn	Glu	Leu	Arg
			20					25					30		
Leu	Arg	Gly	Ile	Leu	Thr	Asp	Val	Thr	Leu	Leu	Val	Gly	Gly	Gln	Pro
		35					40					45			

Leu	Arg	Ala	His	Lys	Ala	Val	Leu	Ile	Ala	Cys	Ser	Gly	Phe	Phe	Tyr	
50						55					60					
Ser	Ile	Phe	Arg	Gly	Arg	Ala	Gly	Val	Gly	Val	Asp	Val	Leu	Ser	Leu	
65					70					75					80	
Pro	Gly	Gly	Pro	Glu	Ala	Arg	Gly	Phe	Ala	Pro	Leu	Leu	Asp	Phe	Met	
				85					90					95		
Tyr	Thr	Ser	Arg	Leu	Arg	Leu	Ser	Pro	Ala	Thr	Ala	Pro	Ala	Val	Leu	
			100				105						110			
Ala	Ala	Ala	Thr	Tyr	Leu	Gln	Met	Glu	His	Val	Val	Gln	Ala	Cys	His	
		115					120					125				
Arg	Phe	Ile	Gln	Ala	Ser	Tyr	Glu	Pro	Leu	Gly	Ile	Ser	Leu	Arg	Pro	
	130					135					140					
Leu	Glu	Ala	Glu	Pro	Pro	Thr	Pro	Pro	Thr	Ala	Pro	Pro	Pro	Gly	Ser	
145					150					155					160	
Pro	Arg	Arg	Ser	Glu	Gly	His	Pro	Asp	Pro	Pro	Thr	Glu	Ser	Arg	Ser	
				165					170					175		
Cys	Ser	Gln	Gly	Pro	Pro	Ser	Pro	Ala	Ser	Pro	Asp	Pro	Lys	Ala	Cys	
			180					185					190			
Asn	Trp	Lys	Lys	Tyr	Lys	Tyr	Ile	Val	Leu	Asn	Ser	Gln	Ala	Ser	Gln	
		195					200					205				
Ala	Gly	Ser	Leu	Val	Gly	Glu	Arg	Ser	Ser	Gly	Gln	Pro	Cys	Pro	Gln	
	210					215					220					
Ala	Arg	Leu	Pro	Ser	Gly	Asp	Glu	Ala	Ser	Ser	Ser	Ser	Ser	Ser	Ser	
225					230					235					240	
Ser	Ser	Ser	Ser	Ser	Glu	Glu	Gly	Pro	Ile	Pro	Gly	Pro	Gln	Ser	Arg	
				245					250					255		
Leu	Ser	Pro	Thr	Ala	Ala	Thr	Val	Gln	Phe	Lys	Cys	Gly	Ala	Pro	Ala	
			260					265					270			
Ser	Thr	Pro	Tyr	Leu	Leu	Thr	Ser	Gln	Ala	Gln	Asp	Thr	Ser	Gly	Ser	
		275					280					285				
Pro	Ser	Glu	Arg	Ala	Arg	Pro	Leu	Pro	Gly	Ser	Glu	Phe	Phe	Ser	Cys	
	290					295					300					
Gln	Asn	Cys	Glu	Ala	Val	Ala	Gly	Cys	Ser	Ser	Gly	Leu	Asp	Ser	Leu	
305					310					315					320	
Val	Pro	Gly	Asp	Glu	Asp	Lys	Pro	Tyr	Lys	Cys	Gln	Leu	Cys	Arg	Ser	
				325					330					335		
Ser	Phe	Arg	Tyr	Lys	Gly	Asn	Leu	Ala	Ser	His	Arg	Thr	Val	His	Thr	
			340					345					350			
Gly	Glu	Lys	Pro	Tyr	His	Cys	Ser	Ile	Cys	Gly	Ala	Arg	Phe	Asn	Arg	
		355					360				365					
Pro	Ala	Asn	Leu	Lys	Thr	His	Ser	Arg	Ile	His	Ser	Gly	Glu	Lys	Pro	
	370					375					380					
Tyr	Lys	Cys	Glu	Thr	Cys	Gly	Ser	Arg	Phe	Val	Gln	Val	Arg	Ser	Gln	
385					390					395					400	

Pro	Pro	Ser	Gly	Phe	Gln	Gly	Lys	Pro	Ala	Arg	Gly	Gly	Val	Gly	Gln	
				405					410					415		
Lys	Gly	Gly	Phe	Cys	Ser	Ser	Gln	Arg	Gln	Asp	Leu	Lys	Ser	Pro	Pro	
			420					425					430			
Ser	Gln	Val	Ala	His	Leu	Arg	Ala	His	Val	Leu	Ile	His	Thr	Gly	Glu	
		435					440					445				
Lys	Pro	Tyr	Pro	Cys	Pro	Thr	Cys	Gly	Thr	Arg	Phe	Arg	His	Leu	Gln	
	450					455					460					
Thr	Leu	Lys	Ser	His	Val	Arg	Ile	His	Thr	Gly	Glu	Lys	Pro	Tyr	His	
465					470					475					480	
Cys	Asp	Pro	Cys	Gly	Leu	His	Phe	Arg	His	Lys	Ser	Gln	Leu	Arg	Leu	
			485					490						495		
His	Leu	Arg	Gln	Lys	His	Gly	Ala	Ala	Thr	Asn	Thr	Lys	Val	His	Tyr	
		500					505						510			
His	Ile	Leu	Gly	Gly	Pro											
		515														

Table 5 hEF-G nucleotide sequence (SEQ ID NO:5)

tctttttcct	cgcgctcctt	gccccggaag	tgctcttaca	acattggctg	cgggcgtgac	60
tttgaccgct	tcccgtgctg	ttaccggcag	ctgaaccac	ccggcgccac	gggactttga	120
cgcgctgctct	gcgcttgcca	tgagactcct	gggagctgca	gccgtcgctg	ctctggggcg	180
cgggaagggcc	cccgcctccc	taggctggca	gaggaagcag	gttaattgga	aggcctgccg	240
atggtcttca	tcaggggtga	ttcctaata	aaaaatacga	aatattggaa	tctcagctca	300
cattgattct	gggaaaacta	cattaacaga	acgagtcctt	tactacactg	gcagaattgc	360
aaagatgcat	gaggtgaaag	gtaaagatgg	agttggtgct	gtcatggatt	ccatggaact	420
agagagacaa	agaggaatca	ctattcagtc	agcagccact	ttcaccatgt	ggaaagatgt	480
caatattaac	attatagata	ctcctgggca	tgtggacttc	acaatagaag	tggaaagggc	540
cctgagagtg	ttggatggtg	cagtccttgt	tctctgtgct	gttgaggagg	tacagtgccca	600
gaccatgact	gtcaatcgtc	agatgaagcg	ctacaacgtt	ccgtttctaa	cttttattaa	660
caaattggac	cgaatgggct	ccaaccagc	cagggccttg	cagcaaatga	ggtctaaact	720
aaatcataat	acagcgttta	tgcagatacc	catgggtttg	gagggttaatt	ttaaaggtat	780
tgtagatctt	attgaggaac	gagccatcta	ttttgatgga	gacttttagtc	agattgttctg	840
atatggtgag	attccagctg	aattaagggc	ggcgccact	gaccaccggc	aggagctaata	900
tgaatgtgtt	gccaatcag	atgaacagct	tggtgagatg	tttctggaag	aaaaaatccc	960
ctcgatttct	gatttaaagc	tagcaattcg	aagagctact	ctgaaaagat	catttactcc	1020
tgtatttttg	ggaagcgctt	tgaagaacaa	aggagttcag	cctcttttag	atgctgtttt	1080
agaatacctc	ccaaatccat	ctgaagtcca	gaactatgct	attctcaata	aaaaggatga	1140
ctcaaaagag	aaaacaaaa	tcctaataga	ctccagtaga	cacaattccc	accatttgt	1200
aggcctggct	tttcccctgg	aggtaggtcg	atttgacaaa	ttaacttatg	ttcgcagtta	1260
tcagggagag	ctaaagaagg	gtgacaccat	ctataacaca	aggacaagaa	agaaagtacg	1320
gttgcaacgg	ctggctcgca	tgcattgccga	catgatggag	gcaagtacag	aggaagtata	1380
tgccggagac	atctgtgcat	tgtttggcat	tgactgtgct	agtggagaca	cattcacaga	1440

caaagccaac	agcggccttt	ctatggagtc	aattcatggt	cctgatcctg	tcatttcaat	1500
agcaatgaag	ccttctaaca	agaacgatct	ggaaaaat	tcaaaaggta	ttggcagggt	1560
tacaagagaa	gatcccat	ttaaagtata	ctttgacact	gagaacaaag	agacagttat	1620
atctggaatg	ggagaattac	acctggaaat	ctatgctcag	aggctggaaa	gagagtatgg	1680
ctgtccttgt	atcacaggaa	agccaaaagt	tgcctttcga	gagaccatta	ctgcccctgt	1740
cccgtttgac	tttacacata	aaaaacaatc	agggtggtgca	ggccagtatg	gaaaagtaat	1800
agggtgtcctg	gagcctctgg	acccagagga	ctacactaaa	ttggaatttt	cagatgaaac	1860
attcggatca	aatattccaa	agcagtttgt	gcctgctgta	gaaaaggggt	ttttagatgc	1920
ctgcgagaag	ggcctctttt	ctggtcacaa	gctctctggg	ctccggtttg	tcctgcaaga	1980
tggagcacac	cacatgggtg	atttctaata	aatctctttc	atccgagcag	gagaagggtgc	2040
tcttaaacia	gccttggcaa	atgcaacatt	atgtattctt	gaacctatta	tggctgtgga	2100
agttgtagct	ccaaatgaat	ttcagggaca	agtaattgca	ggaattaacc	gacgccatgg	2160
ggtaatcact	gggcaagatg	gagttgagga	ctattttaca	ctgtatgcag	atgtccctct	2220
aaatgatatg	tttgggtatt	ccactgaact	taggtcatgc	acagagggaa	agggagaata	2280
cacaatggag	tatagcaggt	atcagccatg	tttaccatcc	acacaagaag	acgtcattaa	2340
taagtatttg	gaagctacag	gtcaacttcc	tgttaaaaaa	ggaaaagcca	agaactaaact	2400
ttgcttactg	tgagttgact	gactctaatt	gaatctgctg	ggttttgata	ctttgatgga	2460
ttccagtggg	ataaattcag	gctgctgaaa	caagaaattc	tgagcccagg	aagcgggctc	2520
ttctttcttc	aaaagaagcc	cttcttggtc	atattcagga	gcttctgtta	tattcaaagg	2580
taattctatg	tctatctcaa	ctctattgat	tggttttata	gttcattgaa	aatcctcaaa	2640
taaaatataa	ttattactga	aatatgttta	atatttaagg	ggaaaagaga	ctaatttcag	2700
ttatactttt	aagcttagaa	tgtatgttca	tttccaaatt	ttgtatcata	agagttttca	2760
acatagagaa	aagctgaaaa	aatgcaaaga	ataaccacat	actttccatc	taccttcctt	2820
tgttaacggg	ttgtttatca	tataataatt	tgttttgtca	tatttgcttt	cactgtctat	2880
tatctgttta	agtctcataa	ctctattttt	agtttgctga	agacttgaaa	gtgaatcgca	2940
tatatcatga	cacttcttgg	agtgtcatta	atgggcaggc	ttttctgttg	aagagtggat	3000
tcogtatgtt	cttcatagag	agtgtttttc	agattcttca	ttgggatatt	aaaatattag	3060
ccaaatttcn	ctctgtttta	tatatgncag	tttatttcag	tttgtggttt	ctgcaaattt	3120
gtaactgcct	ctgttttagg	agtataagta	ttacttcctt	gtggtctatt	gtgaagtaaa	3180
aagtagaccc	ttgcatatac	tattcttgtt	tgtgttcac	ttaatgtttt	tgtacagcta	3240
aatcaaatgt	aatttataga	gttagtttca	tcaacctaat	gaatgctagt	taaatttgaa	3300
ttccttgga	tttatcgat	attgtattca	ctgagattat	gaagggacaa	atgttaatct	3360
tttgtttcca	gaaaaagt	ggctttccca	agcagttcta	ttacccggtt	cagaattgct	3420
tcacccaaaa	atcatctgat	ggtatagatg	gacccatg	cttttcatta	cctgatggta	3480
gaaataaaat	aattgatttt	a				3501

Table 6 hEF-G polypeptide (SEQ ID NO:6)

Met	Arg	Leu	Leu	Gly	Ala	Ala	Ala	Val	Ala	Ala	Leu	Gly	Arg	Gly	Arg
1				5				10						15	
Ala	Pro	Ala	Ser	Leu	Gly	Trp	Gln	Arg	Lys	Gln	Val	Asn	Trp	Lys	Ala
			20					25					30		
Cys	Arg	Trp	Ser	Ser	Ser	Gly	Val	Ile	Pro	Asn	Glu	Lys	Ile	Arg	Asn
		35					40					45			

Ile	Gly	Ile	Ser	Ala	His	Ile	Asp	Ser	Gly	Lys	Thr	Thr	Leu	Thr	Glu	
50						55					60					
Arg	Val	Leu	Tyr	Tyr	Thr	Gly	Arg	Ile	Ala	Lys	Met	His	Glu	Val	Lys	
65					70					75					80	
Gly	Lys	Asp	Gly	Val	Gly	Ala	Val	Met	Asp	Ser	Met	Glu	Leu	Glu	Arg	
				85					90					95		
Gln	Arg	Gly	Ile	Thr	Ile	Gln	Ser	Ala	Ala	Thr	Phe	Thr	Met	Trp	Lys	
			100					105					110			
Asp	Val	Asn	Ile	Asn	Ile	Ile	Asp	Thr	Pro	Gly	His	Val	Asp	Phe	Thr	
		115					120					125				
Ile	Glu	Val	Glu	Arg	Ala	Leu	Arg	Val	Leu	Asp	Gly	Ala	Val	Leu	Val	
	130					135					140					
Leu	Cys	Ala	Val	Gly	Gly	Val	Gln	Cys	Gln	Thr	Met	Thr	Val	Asn	Arg	
145					150					155				160		
Gln	Met	Lys	Arg	Tyr	Asn	Val	Pro	Phe	Leu	Thr	Phe	Ile	Asn	Lys	Leu	
				165					170					175		
Asp	Arg	Met	Gly	Ser	Asn	Pro	Ala	Arg	Ala	Leu	Gln	Gln	Met	Arg	Ser	
			180					185					190			
Lys	Leu	Asn	His	Asn	Thr	Ala	Phe	Met	Gln	Ile	Pro	Met	Gly	Leu	Glu	
		195					200					205				
Gly	Asn	Phe	Lys	Gly	Ile	Val	Asp	Leu	Ile	Glu	Glu	Arg	Ala	Ile	Tyr	
	210					215					220					
Phe	Asp	Gly	Asp	Phe	Ser	Gln	Ile	Val	Arg	Tyr	Gly	Glu	Ile	Pro	Ala	
225					230					235					240	
Glu	Leu	Arg	Ala	Ala	Ala	Thr	Asp	His	Arg	Gln	Glu	Leu	Ile	Glu	Cys	
			245						250					255		
Val	Ala	Asn	Ser	Asp	Glu	Gln	Leu	Gly	Glu	Met	Phe	Leu	Glu	Glu	Lys	
		260						265					270			
Ile	Pro	Ser	Ile	Ser	Asp	Leu	Lys	Leu	Ala	Ile	Arg	Arg	Ala	Thr	Leu	
		275					280					285				
Lys	Arg	Ser	Phe	Thr	Pro	Val	Phe	Leu	Gly	Ser	Ala	Leu	Lys	Asn	Lys	
	290					295					300					
Gly	Val	Gln	Pro	Leu	Leu	Asp	Ala	Val	Leu	Glu	Tyr	Leu	Pro	Asn	Pro	
305					310					315				320		
Ser	Glu	Val	Gln	Asn	Tyr	Ala	Ile	Leu	Asn	Lys	Lys	Asp	Asp	Ser	Lys	
				325					330					335		
Glu	Lys	Thr	Lys	Ile	Leu	Met	Asn	Ser	Ser	Arg	His	Asn	Ser	His	Pro	
			340					345					350			
Phe	Val	Gly	Leu	Ala	Phe	Pro	Leu	Glu	Val	Gly	Arg	Phe	Gly	Gln	Leu	
		355					360					365				
Thr	Tyr	Val	Arg	Ser	Tyr	Gln	Gly	Glu	Leu	Lys	Lys	Gly	Asp	Thr	Ile	
	370					375					380					
Tyr	Asn	Thr	Arg	Thr	Arg	Lys	Lys	Val	Arg	Leu	Gln	Arg	Leu	Ala	Arg	
385					390					395					400	

Met	His	Ala	Asp	Met	Met	Glu	Ala	Ser	Thr	Glu	Glu	Val	Tyr	Ala	Gly	
				405					410						415	
Asp	Ile	Cys	Ala	Leu	Phe	Gly	Ile	Asp	Cys	Ala	Ser	Gly	Asp	Thr	Phe	
			420					425					430			
Thr	Asp	Lys	Ala	Asn	Ser	Gly	Leu	Ser	Met	Glu	Ser	Ile	His	Val	Pro	
		435					440					445				
Asp	Pro	Val	Ile	Ser	Ile	Ala	Met	Lys	Pro	Ser	Asn	Lys	Asn	Asp	Leu	
		450				455					460					
Glu	Lys	Phe	Ser	Lys	Gly	Ile	Gly	Arg	Phe	Thr	Arg	Glu	Asp	Pro	Thr	
465					470					475					480	
Phe	Lys	Val	Tyr	Phe	Asp	Thr	Glu	Asn	Lys	Glu	Thr	Val	Ile	Ser	Gly	
				485					490						495	
Met	Gly	Glu	Leu	His	Leu	Glu	Ile	Tyr	Ala	Gln	Arg	Leu	Glu	Arg	Glu	
			500					505					510			
Tyr	Gly	Cys	Pro	Cys	Ile	Thr	Gly	Lys	Pro	Lys	Val	Ala	Phe	Arg	Glu	
		515					520					525				
Thr	Ile	Thr	Ala	Pro	Val	Pro	Phe	Asp	Phe	Thr	His	Lys	Lys	Gln	Ser	
	530					535					540					
Gly	Gly	Ala	Gly	Gln	Tyr	Gly	Lys	Val	Ile	Gly	Val	Leu	Glu	Pro	Leu	
545					550					555					560	
Asp	Pro	Glu	Asp	Tyr	Thr	Lys	Leu	Glu	Phe	Ser	Asp	Glu	Thr	Phe	Gly	
				565					570					575		
Ser	Asn	Ile	Pro	Lys	Gln	Phe	Val	Pro	Ala	Val	Glu	Lys	Gly	Phe	Leu	
			580					585					590			
Asp	Ala	Cys	Glu	Lys	Gly	Pro	Leu	Ser	Gly	His	Lys	Leu	Ser	Gly	Leu	
		595					600					605				
Arg	Phe	Val	Leu	Gln	Asp	Gly	Ala	His	His	Met	Val	Asp	Ser	Asn	Glu	
	610					615					620					
Ile	Ser	Phe	Ile	Arg	Ala	Gly	Glu	Gly	Ala	Leu	Lys	Gln	Ala	Leu	Ala	
625					630					635					640	
Asn	Ala	Thr	Leu	Cys	Ile	Leu	Glu	Pro	Ile	Met	Ala	Val	Glu	Val	Val	
				645					650					655		
Ala	Pro	Asn	Glu	Phe	Gln	Gly	Gln	Val	Ile	Ala	Gly	Ile	Asn	Arg	Arg	
			660					665					670			
His	Gly	Val	Ile	Thr	Gly	Gln	Asp	Gly	Val	Glu	Asp	Tyr	Phe	Thr	Leu	
		675					680					685				
Tyr	Ala	Asp	Val	Pro	Leu	Asn	Asp	Met	Phe	Gly	Tyr	Ser	Thr	Glu	Leu	
	690					695					700					
Arg	Ser	Cys	Thr	Glu	Gly	Lys	Gly	Glu	Tyr	Thr	Met	Glu	Tyr	Ser	Arg	
705					710					715					720	
Tyr	Gln	Pro	Cys	Leu	Pro	Ser	Thr	Gln	Glu	Asp	Val	Ile	Asn	Lys	Tyr	
			725					730					735			
Leu	Glu	Ala	Thr	Gly	Gln	Leu	Pro	Val	Lys	Lys	Gly	Lys	Ala	Lys	Asn	
			740					745					750			

Table 7 hTRG nucleotide sequence (SEQ ID NO:7)

gccgcgggag	caggcggagg	cggaggcggc	gggggcagga	ggatg	tcgca	gccgccgctg	60
ctccccgcct	cggcggagac	tcggaagttc	acccggggcg	tgagtaagcc	gggcacggcg		120
gccgagctgc	ggcagagcgt	gtctgaggtg	gtgcgcggct	ccgtgctcct	ggcaaagcca		180
aagctaattg	agccactcga	ctatgaaaat	gtcatcgctc	agaagaagac	tcagatcctg		240
aacgactgtt	tacgggagat	gctgctcttc	ccttacgatg	actttcagac	ggccatcctg		300
agacgacagg	gtcgatacat	atgctcaaca	gtgcctgcga	aggcgggaaga	ggaagcacag		360
agcttgtttg	ttacagagtg	catcaaaacc	tataactctg	actggcatct	tgtgaactat		420
aaatatgaag	attactcagg	agagtttcga	cagcttcgga	acaaagtggg	caagttggat		480
aaacttccag	ttcatgtcta	tgaagttgac	gaggaggctg	acaaagatga	ggatgctgcc		540
tcccttggtt	cccagaaagg	tgggatcacc	aagcatggct	ggctgtacaa	aggcaacatg		600
aacagtgcc	tcagcgtgac	catgaggtca	tttaagagac	gatttttcca	cctgattcaa		660
cttggcgatg	gacccataaa	atttgaattt	ttaaaagatc	tccaaaagga	acccaaaagga		720
tcaatatattc	tgggattcct	gtatggggtg	tcgttcagga	acaacaaagt	caggcgtttt		780
gcttttgagc	tcaagatgca	ggacaaaagt	agttatctct	tggcagcaga	cagtgaagtg		840
gaaatggaag	aatggatcac	aattctaaat	aagatcctcc	agctcaactt	tgaagctgca		900
atgcaagaaa	agcgaaatgg	cgactctcac	gaagatgatg	aacaaagcaa	attggaagggt		960
tctggttccg	gtttagatag	ctacctgccg	gaacttgcca	agagtgcagg	agaagcagaa		1020
atcaaaactga	aaagtgaag	cagagtcaaa	cttttttatt	tggaccaga	tgcccagaag		1080
cttgacttct	catcagctga	gccagaagtg	aagtcatttg	aagagaagtt	tggaaaaagg		1140
atccttgtca	agtgcfaatga	tttatctttc	aatttgcaat	gctgtgttgc	cgaaaatgaa		1200
gaaggacca	ctacaaatgt	tgaacctttc	tttgttactc	tatccctggt	tgacataaaa		1260
tacaaccgga	agattttctgc	cgatttccac	gtagacctga	accatttctc	agtgaggcaa		1320
atgatcgcca	ccacgtcccc	ggcgtgatg	aatggcagtg	ggccggaaac	ccaatctgcc		1380
ctcaggggca	tccttcatga	agccgccatg	cagtatccga	agcagggaat	attttcagtc		1440
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aaaactagtc	aagcagthta	gaaccaaagg	cctatatthaa	taaccgcaac	tatgctgaaa	7020
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cagthtttaag	aattthcatc	ttttgccaaa	atgggtggagt	atgtaattgg	taaatcataa	7320
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ccatta						7506

Table 8 hTRG polypeptide sequence (SEQ ID NO:8)

Met	Ser	Gln	Pro	Pro	Leu	Leu	Pro	Ala	Ser	Ala	Glu	Thr	Arg	Lys	Phe
1				5					10					15	
Thr	Arg	Ala	Leu	Ser	Lys	Pro	Gly	Thr	Ala	Ala	Glu	Leu	Arg	Gln	Ser
			20					25					30		
Val	Ser	Glu	Val	Val	Arg	Gly	Ser	Val	Leu	Leu	Ala	Lys	Pro	Lys	Leu
		35				40						45			
Ile	Glu	Pro	Leu	Asp	Tyr	Glu	Asn	Val	Ile	Val	Gln	Lys	Lys	Thr	Gln
	50					55					60				
Ile	Leu	Asn	Asp	Cys	Leu	Arg	Glu	Met	Leu	Leu	Phe	Pro	Tyr	Asp	Asp
65				70					75					80	
Phe	Gln	Thr	Ala	Ile	Leu	Arg	Arg	Gln	Gly	Arg	Tyr	Ile	Cys	Ser	Thr
				85					90					95	
Val	Pro	Ala	Lys	Ala	Glu	Glu	Glu	Ala	Gln	Ser	Leu	Phe	Val	Thr	Glu
			100					105					110		
Cys	Ile	Lys	Thr	Tyr	Asn	Ser	Asp	Trp	His	Leu	Val	Asn	Tyr	Lys	Tyr
		115					120					125			
Glu	Asp	Tyr	Ser	Gly	Glu	Phe	Arg	Gln	Leu	Pro	Asn	Lys	Val	Val	Lys
	130					135					140				
Leu	Asp	Lys	Leu	Pro	Val	His	Val	Tyr	Glu	Val	Asp	Glu	Glu	Val	Asp
145					150					155					160
Lys	Asp	Glu	Asp	Ala	Ala	Ser	Leu	Gly	Ser	Gln	Lys	Gly	Gly	Ile	Thr
				165					170					175	
Lys	His	Gly	Trp	Leu	Tyr	Lys	Gly	Asn	Met	Asn	Ser	Ala	Ile	Ser	Val
		180						185					190		
Thr	Met	Arg	Ser	Phe	Lys	Arg	Arg	Phe	Phe	His	Leu	Ile	Gln	Leu	Gly
		195					200					205			
Asp	Gly	Ser	Tyr	Lys	Phe	Glu	Phe	Leu	Lys	Asp	Leu	Gln	Lys	Glu	Pro
	210					215					220				
Lys	Gly	Ser	Ile	Phe	Leu	Gly	Phe	Leu	Tyr	Gly	Val	Ser	Phe	Arg	Asn
225					230					235					240
Asn	Lys	Val	Arg	Arg	Phe	Ala	Phe	Glu	Leu	Lys	Met	Gln	Asp	Lys	Ser
				245					250					255	
Ser	Tyr	Leu	Leu	Ala	Ala	Asp	Ser	Glu	Val	Glu	Met	Glu	Glu	Trp	Ile
		260						265					270		
Thr	Ile	Leu	Asn	Lys	Ile	Leu	Gln	Leu	Asn	Phe	Glu	Ala	Ala	Met	Gln
		275					280					285			
Glu	Lys	Arg	Asn	Gly	Asp	Ser	His	Glu	Asp	Asp	Glu	Gln	Ser	Lys	Leu
	290					295					300				
Glu	Gly	Ser	Gly	Ser	Gly	Leu	Asp	Ser	Tyr	Leu	Pro	Glu	Leu	Ala	Lys
305					310					315					320
Ser	Ala	Arg	Glu	Ala	Glu	Ile	Lys	Leu	Lys	Ser	Glu	Ser	Arg	Val	Lys
				325					330					335	
Leu	Phe	Tyr	Leu	Asp	Pro	Asp	Ala	Gln	Lys	Leu	Asp	Phe	Ser	Ser	Ala

340						345						350					
Glu	Pro	Glu	Val	Lys	Ser	Phe	Glu	Glu	Lys	Phe	Gly	Lys	Arg	Ile	Leu		
		355					360					365					
Val	Lys	Cys	Asn	Asp	Leu	Ser	Phe	Asn	Leu	Gln	Cys	Cys	Val	Ala	Glu		
	370					375				380							
Asn	Glu	Glu	Gly	Pro	Thr	Thr	Asn	Val	Glu	Pro	Phe	Phe	Val	Thr	Leu		
385					390				395						400		
Ser	Leu	Phe	Asp	Ile	Lys	Tyr	Asn	Arg	Lys	Ile	Ser	Ala	Asp	Phe	His		
				405					410					415			
Val	Asp	Leu	Asn	His	Phe	Ser	Val	Arg	Gln	Met	Ile	Ala	Thr	Thr	Ser		
			420					425					430				
Pro	Ala	Leu	Met	Asn	Gly	Ser	Gly	Pro	Glu	Thr	Gln	Ser	Ala	Leu	Arg		
		435					440					445					
Gly	Ile	Leu	His	Glu	Ala	Ala	Met	Gln	Tyr	Pro	Lys	Gln	Gly	Ile	Phe		
	450					455					460						
Ser	Val	Thr	Cys	Pro	His	Pro	Asp	Ile	Phe	Leu	Val	Ala	Arg	Ile	Glu		
465					470					475					480		
Lys	Val	Leu	Gln	Gly	Ser	Ile	Thr	His	Cys	Ala	Glu	Pro	Tyr	Met	Lys		
				485					490					495			
Ser	Ser	Asp	Ser	Ser	Lys	Val	Ala	Gln	Lys	Val	Leu	Lys	Asn	Ala	Lys		
			500					505					510				
Gln	Ala	Cys	Gln	Arg	Leu	Gly	Gln	Tyr	Arg	Met	Pro	Phe	Ala	Trp	Ala		
		515					520					525					
Ala	Arg	Thr	Leu	Phe	Lys	Asp	Ala	Ser	Gly	Asn	Leu	Asp	Lys	Asn	Ala		
	530					535					540						
Arg	Phe	Ser	Ala	Ile	Tyr	Arg	Gln	Asp	Ser	Asn	Lys	Leu	Ser	Asn	Asp		
545					550					555					560		
Asp	Met	Leu	Lys	Leu	Leu	Ala	Asp	Phe	Arg	Lys	Pro	Glu	Lys	Met	Ala		
				565					570					575			
Lys	Leu	Pro	Val	Ile	Leu	Gly	Asn	Leu	Asp	Ile	Thr	Ile	Asp	Asn	Val		
			580					585					590				
Ser	Ser	Asp	Phe	Pro	Asn	Tyr	Val	Asn	Ser	Ser	Tyr	Ile	Pro	Thr	Lys		
		595					600					605					
Gln	Phe	Glu	Thr	Cys	Ser	Lys	Thr	Pro	Ile	Thr	Phe	Glu	Val	Glu	Glu		
	610					615					620						
Phe	Val	Pro	Cys	Ile	Pro	Lys	His	Thr	Gln	Pro	Tyr	Thr	Ile	Tyr	Thr		
625					630					635					640		
Asn	His	Leu	Tyr	Val	Tyr	Pro	Lys	Tyr	Leu	Lys	Tyr	Asp	Ser	Gln	Lys		
				645					650					655			
Ser	Phe	Ala	Lys	Ala	Arg	Asn	Ile	Ala	Ile	Cys	Ile	Glu	Phe	Lys	Asp		
			660					665					670				
Ser	Asp	Glu	Glu	Asp	Ser	Gln	Pro	Leu	Lys	Cys	Ile	Tyr	Gly	Arg	Pro		
		675					680					685					
Gly	Gly	Pro	Val	Phe	Thr	Arg	Ser	Ala	Phe	Ala	Ala	Val	Leu	His	His		

690	695	700
His Gln Asn Pro Glu Phe Tyr Asp Glu Ile Lys Ile Glu Leu Pro Thr 705 710 715 720		
Gln Leu His Glu Lys His His Leu Leu Leu Thr Phe Phe His Val Ser 725 730 735		
Cys Asp Asn Ser Ser Lys Gly Ser Thr Lys Lys Arg Asp Val Val Glu 740 745 750		
Thr Gln Val Gly Tyr Ser Trp Leu Pro Leu Leu Lys Asp Gly Arg Val 755 760 765		
Val Thr Ser Glu Gln His Ile Pro Val Ser Ala Asn Leu Pro Ser Gly 770 775 780		
Tyr Leu Gly Tyr Gln Glu Leu Gly Met Gly Arg His Tyr Gly Pro Glu 785 790 795 800		
Ile Lys Trp Val Asp Gly Gly Lys Pro Leu Leu Lys Ile Ser Thr His 805 810 815		
Leu Val Ser Thr Val Tyr Thr Gln Asp Gln His Leu His Asn Phe Phe 820 825 830		
Gln Tyr Cys Gln Lys Thr Glu Ser Gly Ala Gln Ala Leu Gly Asn Glu 835 840 845		
Leu Val Lys Tyr Leu Lys Ser Leu His Ala Met Glu Gly His Val Met 850 855 860		
Ile Ala Phe Leu Pro Thr Ile Leu Asn Gln Leu Phe Arg Val Leu Thr 865 870 875 880		
Arg Ala Thr Gln Glu Glu Val Ala Val Asn Val Thr Arg Val Ile Ile 885 890 895		
His Val Val Ala Gln Cys His Glu Glu Gly Leu Glu Ser His Leu Arg 900 905 910		
Ser Tyr Val Lys Tyr Ala Tyr Lys Ala Glu Pro Tyr Val Ala Ser Glu 915 920 925		
Tyr Lys Thr Val His Glu Glu Leu Thr Lys Ser Met Thr Thr Ile Leu 930 935 940		
Lys Pro Ser Ala Asp Phe Leu Thr Ser Asn Lys Leu Leu Lys Tyr Ser 945 950 955 960		
Trp Phe Phe Phe Asp Val Leu Ile Lys Ser Met Ala Gln His Leu Ile 965 970 975		
Glu Asn Ser Lys Val Lys Leu Leu Arg Asn Gln Arg Phe Pro Ala Ser 980 985 990		
Tyr His His Ala Val Glu Thr Val Val Asn Met Leu Met Pro His Ile 995 1000 1005		
Thr Gln Lys Phe Arg Asp Asn Pro Glu Ala Ser Lys Asn Ala Asn 1010 1015 1020		
His Ser Leu Ala Val Phe Ile Lys Arg Cys Phe Thr Phe Met Asp 1025 1030 1035		

Arg	Gly	Phe	Val	Phe	Lys	Gln	Ile	Asn	Asn	Tyr	Ile	Ser	Cys	Phe
1040						1045					1050			
Ala	Pro	Gly	Asp	Pro	Lys	Thr	Leu	Phe	Glu	Tyr	Lys	Phe	Glu	Phe
1055						1060					1065			
Leu	Arg	Val	Val	Cys	Asn	His	Glu	His	Tyr	Ile	Pro	Leu	Asn	Leu
1070						1075					1080			
Pro	Met	Pro	Phe	Gly	Lys	Gly	Arg	Ile	Gln	Arg	Tyr	Gln	Asp	Leu
1085						1090					1095			
Gln	Leu	Asp	Tyr	Ser	Leu	Thr	Asp	Glu	Phe	Cys	Arg	Asn	His	Phe
1100						1105					1110			
Leu	Val	Gly	Leu	Leu	Leu	Arg	Glu	Val	Gly	Thr	Ala	Leu	Gln	Glu
1115						1120					1125			
Phe	Arg	Glu	Val	Arg	Leu	Ile	Ala	Ile	Ser	Val	Leu	Lys	Asn	Leu
1130						1135					1140			
Leu	Ile	Lys	His	Ser	Phe	Asp	Asp	Arg	Tyr	Ala	Ser	Arg	Ser	His
1145						1150					1155			
Gln	Ala	Arg	Ile	Ala	Thr	Leu	Tyr	Leu	Pro	Leu	Phe	Gly	Leu	Leu
1160						1165					1170			
Ile	Glu	Asn	Val	Gln	Arg	Ile	Asn	Val	Arg	Asp	Val	Ser	Pro	Phe
1175						1180					1185			
Pro	Val	Asn	Ala	Gly	Met	Thr	Val	Lys	Asp	Glu	Ser	Leu	Ala	Leu
1190						1195					1200			
Pro	Ala	Val	Asn	Pro	Leu	Val	Thr	Pro	Gln	Lys	Gly	Ser	Thr	Leu
1205						1210					1215			
Asp	Asn	Ser	Leu	His	Lys	Asp	Leu	Leu	Gly	Ala	Ile	Ser	Gly	Ile
1220						1225					1230			
Ala	Ser	Pro	Tyr	Thr	Thr	Ser	Thr	Pro	Asn	Ile	Asn	Ser	Val	Arg
1235						1240					1245			
Asn	Ala	Asp	Ser	Arg	Gly	Ser	Leu	Ile	Ser	Thr	Asp	Ser	Gly	Asn
1250						1255					1260			
Ser	Leu	Pro	Glu	Arg	Asn	Ser	Glu	Lys	Ser	Asn	Ser	Leu	Asp	Lys
1265						1270					1275			
His	Gln	Gln	Ser	Ser	Thr	Leu	Gly	Asn	Ser	Val	Val	Arg	Cys	Asp
1280						1285					1290			
Lys	Leu	Asp	Gln	Ser	Glu	Ile	Lys	Ser	Leu	Leu	Met	Cys	Phe	Leu
1295						1300					1305			
Tyr	Ile	Leu	Lys	Ser	Met	Ser	Asp	Asp	Ala	Leu	Phe	Thr	Tyr	Trp
1310						1315					1320			
Asn	Lys	Ala	Ser	Thr	Ser	Glu	Leu	Met	Asp	Phe	Phe	Thr	Ile	Ser
1325						1330					1335			
Glu	Val	Cys	Leu	His	Gln	Phe	Gln	Tyr	Met	Gly	Lys	Arg	Tyr	Ile
1340						1345					1350			
Ala	Arg	Thr	Gly	Met	Met	His	Ala	Arg	Leu	Gln	Gln	Leu	Gly	Ser
1355						1360					1365			

Leu 1370	Asp	Asn	Ser	Leu	Thr	Phe 1375	Asn	His	Ser	Tyr	Gly 1380	His	Ser	Asp
Ala 1385	Asp	Val	Leu	His	Gln	Ser 1390	Leu	Leu	Glu	Ala	Asn 1395	Ile	Ala	Thr
Glu 1400	Val	Cys	Leu	Thr	Ala	Leu 1405	Asp	Thr	Leu	Ser	Leu 1410	Phe	Thr	Leu
Ala 1415	Phe	Lys	Asn	Gln	Leu	Leu 1420	Ala	Asp	His	Gly	His 1425	Asn	Pro	Leu
Met 1430	Lys	Lys	Val	Phe	Asp	Val 1435	Tyr	Leu	Cys	Phe	Leu 1440	Gln	Lys	His
Gln 1445	Ser	Glu	Thr	Ala	Leu	Lys 1450	Asn	Val	Phe	Thr	Ala 1455	Leu	Arg	Ser
Leu 1460	Ile	Tyr	Lys	Phe	Pro	Ser 1465	Thr	Phe	Tyr	Glu	Gly 1470	Arg	Ala	Asp
Met 1475	Cys	Ala	Ala	Leu	Cys	Tyr 1480	Glu	Ile	Leu	Lys	Cys 1485	Cys	Asn	Ser
Lys 1490	Leu	Ser	Ser	Ile	Arg	Thr 1495	Glu	Ala	Ser	Gln	Leu 1500	Leu	Tyr	Phe
Leu 1505	Met	Arg	Asn	Asn	Phe	Asp 1510	Tyr	Thr	Gly	Lys	Lys 1515	Ser	Phe	Val
Arg 1520	Thr	His	Leu	Gln	Val	Ile 1525	Ile	Ser	Val	Ser	Gln 1530	Leu	Ile	Ala
Asp 1535	Val	Val	Gly	Ile	Gly	Gly 1540	Thr	Arg	Phe	Gln	Gln 1545	Ser	Leu	Ser
Ile 1550	Ile	Asn	Asn	Cys	Ala	Asn 1555	Ser	Asp	Arg	Leu	Ile 1560	Lys	His	Thr
Ser 1565	Phe	Ser	Ser	Asp	Val	Lys 1570	Asp	Leu	Thr	Lys	Arg 1575	Ile	Arg	Thr
Val 1580	Leu	Met	Ala	Thr	Ala	Gln 1585	Met	Lys	Glu	His	Glu 1590	Asn	Asp	Pro
Glu 1595	Met	Leu	Val	Asp	Leu	Gln 1600	Tyr	Ser	Leu	Ala	Lys 1605	Ser	Tyr	Ala
Ser 1610	Thr	Pro	Glu	Leu	Arg	Lys 1615	Thr	Trp	Leu	Asp	Ser 1620	Met	Ala	Arg
Ile 1625	His	Val	Lys	Asn	Gly	Asp 1630	Leu	Ser	Glu	Ala	Ala 1635	Met	Cys	Tyr
Val 1640	His	Val	Thr	Ala	Leu	Val 1645	Ala	Glu	Tyr	Leu	Thr 1650	Arg	Lys	Glu
Ala 1655	Val	Gln	Trp	Glu	Pro	Pro 1660	Leu	Leu	Pro	His	Ser 1665	His	Ser	Ala
Cys 1670	Leu	Arg	Arg	Ser	Arg	Gly 1675	Gly	Val	Phe	Arg	Gln 1680	Gly	Cys	Thr
Ala 1685	Phe	Arg	Val	Ile	Thr	Pro 1690	Asn	Ile	Asp	Glu	Glu 1695	Ala	Ser	Met

Met	Glu	Asp	Val	Gly	Met	Gln	Asp	Val	His	Phe	Asn	Glu	Asp	Val
1700						1705					1710			
Leu	Met	Glu	Leu	Leu	Glu	Gln	Cys	Ala	Asp	Gly	Leu	Trp	Lys	Ala
1715						1720					1725			
Glu	Arg	Tyr	Glu	Leu	Ile	Ala	Asp	Ile	Tyr	Lys	Leu	Ile	Ile	Pro
1730						1735					1740			
Ile	Tyr	Glu	Lys	Arg	Arg	Asp	Phe	Glu	Arg	Leu	Ala	His	Leu	Tyr
1745						1750					1755			
Asp	Thr	Leu	His	Arg	Ala	Tyr	Ser	Lys	Val	Thr	Glu	Val	Met	His
1760						1765					1770			
Ser	Gly	Arg	Arg	Leu	Leu	Gly	Thr	Tyr	Phe	Arg	Val	Ala	Phe	Phe
1775						1780					1785			
Gly	Gln	Ala	Ala	Gln	Tyr	Gln	Phe	Thr	Asp	Ser	Glu	Thr	Asp	Val
1790						1795					1800			
Glu	Gly	Phe	Phe	Glu	Asp	Glu	Asp	Gly	Lys	Glu	Tyr	Ile	Tyr	Lys
1805						1810					1815			
Glu	Pro	Lys	Leu	Thr	Pro	Leu	Ser	Glu	Ile	Ser	Gln	Arg	Leu	Leu
1820						1825					1830			
Lys	Leu	Tyr	Ser	Asp	Lys	Phe	Gly	Ser	Glu	Asn	Val	Lys	Met	Ile
1835						1840					1845			
Gln	Asp	Ser	Gly	Lys	Val	Asn	Pro	Lys	Asp	Leu	Asp	Ser	Lys	Tyr
1850						1855					1860			
Ala	Tyr	Ile	Gln	Val	Thr	His	Val	Ile	Pro	Phe	Phe	Asp	Glu	Lys
1865						1870					1875			
Glu	Leu	Gln	Glu	Arg	Lys	Thr	Glu	Phe	Glu	Arg	Ser	His	Asn	Ile
1880						1885					1890			
Arg	Arg	Phe	Met	Phe	Glu	Met	Pro	Phe	Thr	Gln	Thr	Gly	Lys	Arg
1895						1900					1905			
Gln	Gly	Gly	Val	Glu	Glu	Gln	Cys	Lys	Arg	Arg	Thr	Ile	Leu	Thr
1910						1915					1920			
Ala	Ile	His	Cys	Phe	Pro	Tyr	Val	Lys	Lys	Arg	Ile	Pro	Val	Met
1925						1930					1935			
Tyr	Gln	His	His	Thr	Asp	Leu	Asn	Pro	Ile	Glu	Val	Ala	Ile	Asp
1940						1945					1950			
Glu	Met	Ser	Lys	Lys	Val	Ala	Glu	Leu	Arg	Gln	Leu	Cys	Ser	Ser
1955						1960					1965			
Ala	Glu	Val	Asp	Met	Ile	Lys	Leu	Gln	Leu	Lys	Leu	Gln	Gly	Ser
1970						1975					1980			
Val	Ser	Val	Gln	Val	Asn	Ala	Gly	Pro	Leu	Ala	Tyr	Ala	Arg	Ala
1985						1990					1995			
Phe	Leu	Asp	Asp	Thr	Asn	Thr	Lys	Arg	Tyr	Pro	Asp	Asn	Lys	Val
2000						2005					2010			
Lys	Leu	Leu	Lys	Glu	Val	Phe	Arg	Gln	Phe	Val	Glu	Ala	Cys	Gly
2015						2020					2025			

Gln	Ala	Leu	Ala	Val	Asn	Glu	Arg	Leu	Ile	Lys	Glu	Asp	Gln	Leu
2030						2035					2040			
Glu	Tyr	Gln	Glu	Glu	Met	Lys	Ala	Asn	Tyr	Arg	Glu	Met	Ala	Lys
2045						2050					2055			
Glu	Leu	Ser	Glu	Ile	Met	His	Glu	Gln	Ile	Cys	Pro	Leu	Glu	Glu
2060						2065					2070			
Lys	Thr	Ser	Val	Leu	Pro	Asn	Ser	Leu	His	Ile	Phe	Asn	Ala	Ile
2075						2080					2085			
Ser	Gly	Thr	Pro	Thr	Ser	Thr	Met	Val	His	Gly	Met	Thr	Ser	Ser
2090						2095					2100			
Ser	Ser	Val	Val											
2105														

Table 9 hMX1 nucleotide sequence (SEQ ID NO:9)

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ttccagcggg	ataagagaaa	tcaaataatg	acctacatcg	gtcccatcct	ggcctctgtg	300
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Table 10 hMX1 polypeptide sequence (SEQ ID NO:10)

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			20					25					30		
Thr	Asp	Tyr	Gly	Gln	Val	Phe	Thr	Tyr	Lys	Gln	Ser	Thr	Ile	Thr	His
		35					40					45			
Gln	Lys	Val	Thr	Ala	Met	His	Pro	Thr	Asn	Glu	Glu	Gly	Val	Asp	Asp

50					55					60					
Met 65	Ala	Ser	Leu	Thr	Glu 70	Leu	His	Gly	Gly	Ser 75	Ile	Met	Tyr	Asn	Leu 80
Phe	Gln	Arg	Tyr	Lys 85	Arg	Asn	Gln	Ile	Trp 90	Thr	Tyr	Ile	Gly	Ser 95	Ile
Leu	Ala	Ser	Val 100	Asn	Pro	Tyr	Gln	Pro 105	Ile	Ala	Gly	Leu	Tyr 110	Glu	Pro
Ala	Thr	Met 115	Glu	Gln	Tyr	Ser	Arg 120	Arg	His	Leu	Gly	Glu 125	Leu	Pro	Pro
His	Ile 130	Phe	Ala	Ile	Ala	Asn 135	Glu	Cys	Tyr	Arg	Cys 140	Leu	Trp	Lys	Arg
His 145	Asp	Asn	Gln	Cys	Ile 150	Leu	Ile	Lys	Gly	Glu 155	Ser	Gly	Ala	Gly	Lys 160
Thr	Glu	Ser	Thr	Lys 165	Leu	Ile	Leu	Lys	Phe 170	Leu	Ser	Val	Ile	Ser 175	Gln
Gln	Ser	Leu	Glu 180	Leu	Ser	Leu	Lys	Glu 185	Lys	Thr	Ser	Cys	Val 190	Glu	Arg
Ala	Ile	Leu 195	Glu	Ser	Ser	Pro	Ile 200	Met	Glu	Ala	Phe	Gly 205	Asn	Ala	Lys
Thr	Val 210	Tyr	Asn	Asn	Asn	Ser 215	Ser	Arg	Phe	Gly	Lys 220	Phe	Val	Gln	Leu
Asn 225	Ile	Cys	Gln	Lys	Gly 230	Asn	Ile	Gln	Gly	Gly 235	Arg	Ile	Val	Asp	Cys 240
Ile	Leu	Ser	Ser	Gln 245	Asn	Arg	Val	Val	Arg 250	Gln	Asn	Pro	Gly	Glu 255	Arg
Asn	Tyr	His	Ile 260	Phe	Tyr	Ala	Leu 265	Leu	Ala	Gly	Leu	Glu 270	His	Glu	Glu
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Gln	Ser	Gly 290	Cys	Val	Glu	Asp 295	Lys	Thr	Ile	Ser	Asp 300	Gln	Glu	Ser	Phe
Arg 305	Glu	Val	Ile	Thr	Ala 310	Met	Asp	Val	Met	Gln 315	Phe	Ser	Lys	Glu	Glu 320
Val	Arg	Glu	Val	Ser 325	Arg	Leu	Leu	Ala	Gly 330	Ile	Leu	His	Leu	Gly 335	Asn
Ile	Glu	Phe	Ile 340	Thr	Ala	Gly	Gly	Ala 345	Gln	Val	Ser	Phe	Lys 350	Thr	Ala
Leu	Gly	Arg 355	Ser	Ala	Glu	Leu	Leu 360	Gly	Leu	Asp	Pro	Thr 365	Gln	Leu	Thr
Asp 370	Ala	Leu	Thr	Gln	Arg	Ser 375	Met	Phe	Leu	Arg	Gly 380	Glu	Glu	Ile	Leu
Thr 385	Pro	Leu	Asn	Val	Gln 390	Gln	Ala	Val	Asp	Ser 395	Arg	Asp	Ser	Leu	Ala 400
Met	Ala	Leu	Tyr	Ala	Cys	Cys	Phe	Glu	Trp	Val	Ile	Lys	Lys	Ile	Asn

				405				410				415			
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Lys	His	Tyr	Ala	Gly	Glu 550	Val	Gln	Tyr	Asp	Val 555	Arg	Gly	Ile	Leu	Glu 560
Lys	Asn	Arg	Asp	Thr 565	Phe	Arg	Asp	Asp	Leu 570	Leu	Asn	Leu	Leu	Arg 575	Glu
Ser	Arg	Phe	Asp 580	Phe	Ile	Tyr	Asp	Leu 585	Phe	Glu	His	Val	Ser 590	Ser	Arg
Asn	Asn	Gln 595	Asp	Thr	Leu	Lys	Cys 600	Gly	Ser	Lys	His	Arg 605	Arg	Pro	Thr
Val	Ser	Ser	Gln	Phe	Lys	Val 615	Asp	Ser	Leu	His	Ser 620	Leu	Met	Ala	Thr
Leu	Ser	Ser	Ser	Asn	Pro 630	Phe	Phe	Val	Arg	Cys 635	Ile	Lys	Pro	Asn	Met 640
Gln	Lys	Met	Pro	Asp 645	Gln	Phe	Asp	Gln	Ala 650	Val	Val	Leu	Asn	Gln 655	Leu
Arg	Tyr	Ser	Gly 660	Met	Leu	Glu	Thr	Val 665	Arg	Ile	Arg	Lys	Ala 670	Gly	Tyr
Ala	Val	Arg 675	Arg	Pro	Phe	Gln	Asp 680	Phe	Tyr	Lys	Arg	Tyr 685	Lys	Val	Leu
Met	Arg	Asn	Leu	Ala	Leu	Pro 695	Glu	Asp	Val	Arg	Gly 700	Lys	Cys	Thr	Ser
Leu	Leu	Gln	Leu	Tyr	Asp 710	Ala	Ser	Asn	Ser	Glu 715	Trp	Gln	Leu	Gly	Lys 720
Thr	Lys	Val	Phe	Leu 725	Arg	Glu	Ser	Leu	Glu 730	Gln	Lys	Leu	Glu	Lys 735	Arg
Arg	Glu	Glu	Glu 740	Val	Ser	His	Ala	Ala 745	Met	Val	Ile	Arg	Ala 750	His	Val
Leu	Gly	Phe	Leu	Ala	Arg	Lys	Gln	Tyr	Arg	Lys	Val	Leu	Tyr	Cys	Val

755					760					765					
Val	Ile	Ile	Gln	Lys	Asn	Tyr	Arg	Ala	Phe	Leu	Leu	Arg	Arg	Arg	Phe
770						775					780				
Leu	His	Leu	Lys	Lys	Ala	Ala	Ile	Val	Phe	Gln	Lys	Gln	Leu	Arg	Gly
785					790					795					800
Gln	Ile	Ala	Arg	Arg	Val	Tyr	Arg	Gln	Leu	Leu	Ala	Glu	Lys	Arg	Glu
				805					810					815	
Gln	Glu	Glu	Lys	Lys	Lys	Gln	Glu	Glu	Glu	Glu	Lys	Lys	Lys	Arg	Glu
			820					825						830	
Glu	Glu	Glu	Arg	Glu	Arg	Glu	Arg	Glu	Arg	Arg	Glu	Ala	Glu	Leu	Arg
			835				840					845			
Ala	Gln	Gln	Glu	Glu	Glu	Thr	Arg	Lys	Gln	Gln	Glu	Leu	Glu	Ala	Leu
850						855					860				
Gln	Lys	Ser	Gln	Lys	Glu	Ala	Glu	Leu	Thr	Arg	Glu	Leu	Glu	Lys	Gln
865					870					875					880
Lys	Glu	Asn	Lys	Gln	Val	Glu	Glu	Ile	Leu	Arg	Leu	Glu	Lys	Glu	Ile
				885					890					895	
Glu	Asp	Leu	Gln	Arg	Met	Lys	Glu	Gln	Gln	Glu	Leu	Ser	Leu	Thr	Glu
			900					905					910		
Ala	Ser	Leu	Gln	Lys	Leu	Gln	Glu	Arg	Arg	Asp	Gln	Glu	Leu	Arg	Arg
		915					920					925			
Leu	Glu	Glu	Glu	Ala	Cys	Arg	Ala	Ala	Gln	Glu	Phe	Leu	Glu	Ser	Leu
930						935					940				
Asn	Phe	Asp	Glu	Ile	Asp	Glu	Cys	Val	Arg	Asn	Ile	Glu	Arg	Ser	Leu
945					950					955					960
Ser	Gly	Gly	Ser	Glu	Phe	Ser	Ser	Glu	Leu	Ala	Glu	Ser	Ala	Cys	Glu
				965					970					975	
Glu	Lys	Pro	Asn	Phe	Asn	Phe	Ser	Gln	Pro	Tyr	Pro	Glu	Glu	Glu	Val
			980					985					990		
Asp	Glu	Gly	Phe	Glu	Ala	Asp	Asp	Asp	Ala	Phe	Lys	Asp	Ser	Pro	Asn
		995					1000					1005			
Pro	Ser	Glu	His	Gly	His	Ser	Asp	Gln	Arg	Thr	Ser	Gly	Ile	Arg	
1010						1015						1020			
Thr	Ser	Asp	Asp	Ser	Ser	Glu	Glu	Asp	Pro	Tyr	Met	Asn	Asp	Thr	
1025						1030					1035				
Val	Val	Pro	Thr	Ser	Pro	Ser	Ala	Asp	Ser	Thr	Val	Leu	Leu	Ala	
1040						1045					1050				
Pro	Ser	Val	Gln	Asp	Ser	Gly	Ser	Leu	His	Asn	Ser	Ser	Ser	Gly	
1055						1060					1065				
Glu	Ser	Thr	Tyr	Cys	Met	Pro	Gln	Asn	Ala	Gly	Asp	Leu	Pro	Ser	
1070						1075					1080				
Pro	Asp	Gly	Asp	Tyr	Asp	Tyr	Asp	Gln	Asp	Asp	Tyr	Glu	Asp	Gly	
1085						1090					1095				
Ala	Ile	Thr	Ser	Gly	Ser	Ser	Val	Thr	Phe	Ser	Asn	Ser	Tyr	Gly	

1100	1105	1110
Ser Gln Trp Ser Pro Asp Tyr Arg Cys Ser Val Gly Thr Tyr Asn 1115 1120 1125		
Ser Ser Gly Ala Tyr Arg Phe Ser Ser Glu Gly Ala Gln Ser Ser 1130 1135 1140		
Phe Glu Asp Ser Glu Glu Asp Phe Asp Ser Arg Phe Asp Thr Asp 1145 1150 1155		
Asp Glu Leu Ser Tyr Arg Arg Asp Ser Val Tyr Ser Cys Val Thr 1160 1165 1170		
Leu Pro Tyr Phe His Ser Phe Leu Tyr Met Lys Gly Gly Leu Met 1175 1180 1185		
Asn Ser Trp Lys Arg Arg Trp Cys Val Leu Lys Asp Glu Thr Phe 1190 1195 1200		
Leu Trp Phe Arg Ser Lys Gln Glu Ala Leu Lys Gln Gly Trp Leu 1205 1210 1215		
His Lys Lys Gly Gly Gly Ser Ser Thr Leu Ser Arg Arg Asn Trp 1220 1225 1230		
Lys Lys Arg Trp Phe Val Leu Arg Gln Ser Lys Leu Met Tyr Phe 1235 1240 1245		
Glu Asn Asp Ser Glu Glu Lys Leu Lys Gly Thr Val Glu Val Arg 1250 1255 1260		
Thr Ala Lys Glu Ile Ile Asp Asn Thr Thr Lys Glu Asn Gly Ile 1265 1270 1275		
Asp Ile Ile Met Ala Asp Arg Thr Phe His Leu Ile Ala Glu Ser 1280 1285 1290		
Pro Glu Asp Ala Ser Gln Trp Phe Ser Val Leu Ser Gln Val His 1295 1300 1305		
Ala Ser Thr Asp Gln Glu Ile Gln Glu Met His Asp Glu Gln Ala 1310 1315 1320		
Asn Pro Gln Asn Ala Val Gly Thr Leu Asp Val Gly Leu Ile Asp 1325 1330 1335		
Ser Val Cys Ala Ser Asp Ser Pro Asp Arg Pro Asn Ser Phe Val 1340 1345 1350		
Ile Ile Thr Ala Asn Arg Val Leu His Cys Asn Ala Asp Thr Pro 1355 1360 1365		
Glu Glu Met His His Trp Ile Thr Leu Leu Gln Arg Ser Lys Gly 1370 1375 1380		
Asp Thr Arg Val Glu Gly Gln Glu Phe Ile Val Arg Gly Trp Leu 1385 1390 1395		
His Lys Glu Val Lys Asn Ser Pro Lys Met Ser Ser Leu Lys Leu 1400 1405 1410		
Lys Lys Arg Trp Phe Val Leu Thr His Asn Ser Leu Asp Tyr Tyr 1415 1420 1425		
Lys Ser Ser Glu Lys Asn Ala Leu Lys Leu Gly Thr Leu Val Leu		

1430	1435	1440
Asn Ser Leu Cys Ser Val Val Pro Pro Asp Glu Lys Ile Phe Lys 1445 1450 1455		
Glu Thr Gly Tyr Trp Asn Val Thr Val Tyr Gly Arg Lys His Cys 1460 1465 1470		
Tyr Arg Leu Tyr Thr Lys Leu Leu Asn Glu Ala Thr Arg Trp Ser 1475 1480 1485		
Ser Val Ile Gln Asn Val Thr Asp Thr Lys Ala Pro Ile Asp Thr 1490 1495 1500		
Pro Thr Gln Gln Leu Ile Gln Asp Ile Lys Glu Asn Cys Leu Asn 1505 1510 1515		
Ser Asp Val Val Glu Gln Ile Tyr Lys Arg Asn Pro Ile Leu Arg 1520 1525 1530		
Tyr Thr His His Pro Leu His Ser Pro Leu Leu Pro Leu Pro Tyr 1535 1540 1545		
Gly Asp Ile Asn Leu Asn Leu Leu Lys Asp Lys Gly Tyr Thr Thr 1550 1555 1560		
Leu Gln Asp Glu Ala Ile Lys Ile Phe Asn Ser Leu Gln Gln Leu 1565 1570 1575		
Glu Ser Met Ser Asp Pro Ile Pro Ile Ile Gln Gly Ile Leu Gln 1580 1585 1590		
Thr Gly His Asp Leu Arg Pro Leu Arg Asp Glu Leu Tyr Cys Gln 1595 1600 1605		
Leu Ile Lys Gln Thr Asn Lys Val Pro His Pro Gly Ser Val Gly 1610 1615 1620		
Asn Leu Tyr Ser Trp Gln Ile Leu Thr Cys Leu Ser Cys Thr Phe 1625 1630 1635		
Leu Pro Ser Arg Gly Ile Leu Lys Tyr Leu Lys Phe His Leu Lys 1640 1645 1650		
Arg Ile Arg Glu Gln Phe Pro Gly Thr Glu Met Glu Lys Tyr Ala 1655 1660 1665		
Leu Phe Thr Tyr Glu Ser Leu Lys Lys Thr Lys Cys Arg Glu Phe 1670 1675 1680		
Val Pro Ser Arg Asp Glu Ile Glu Ala Leu Ile His Arg Gln Glu 1685 1690 1695		
Met Thr Ser Thr Val Tyr Cys His Gly Gly Gly Ser Cys Lys Ile 1700 1705 1710		
Thr Ile Asn Ser His Thr Thr Ala Gly Glu Val Val Glu Lys Leu 1715 1720 1725		
Ile Arg Gly Leu Ala Met Glu Asp Ser Arg Asn Met Phe Ala Leu 1730 1735 1740		
Phe Glu Tyr Asn Gly His Val Asp Lys Ala Ile Glu Ser Arg Thr 1745 1750 1755		
Val Val Ala Asp Val Leu Ala Lys Phe Glu Lys Leu Ala Ala Thr		

1760	1765	1770
Ser Glu Val Gly Asp Leu Pro Trp Lys Phe Tyr Phe Lys Leu Tyr	1775	1780
Cys Phe Leu Asp Thr Asp Asn Val Pro Lys Asp Ser Val Glu Phe	1790	1800
Ala Phe Met Phe Glu Gln Ala His Glu Ala Val Ile His Gly His	1805	1810
His Pro Ala Pro Glu Glu Asn Leu Gln Val Leu Ala Ala Leu Arg	1820	1825
Leu Gln Tyr Leu Gln Gly Asp Tyr Thr Leu His Ala Ala Ile Pro	1835	1840
Pro Leu Glu Glu Val Tyr Ser Leu Gln Arg Leu Lys Ala Arg Ile	1850	1855
Ser Gln Ser Thr Lys Thr Phe Thr Pro Cys Glu Arg Leu Glu Lys	1865	1870
Arg Arg Thr Ser Phe Leu Glu Gly Thr Leu Arg Arg Ser Phe Arg	1880	1885
Thr Gly Ser Val Val Arg Gln Lys Val Glu Glu Glu Gln Met Leu	1895	1900
Asp Met Trp Ile Lys Glu Glu Val Ser Ser Ala Arg Ala Ser Ile	1910	1915
Ile Asp Lys Trp Arg Lys Phe Gln Gly Met Asn Gln Glu Gln Ala	1925	1930
Met Ala Lys Tyr Met Ala Leu Ile Lys Glu Trp Pro Gly Tyr Gly	1940	1945
Ser Thr Leu Phe Asp Val Glu Cys Lys Glu Gly Gly Phe Pro Gln	1955	1960
Glu Leu Trp Leu Gly Val Ser Ala Asp Ala Val Ser Val Tyr Lys	1970	1975
Arg Gly Glu Gly Arg Pro Leu Glu Val Phe Gln Tyr Glu His Ile	1985	1990
Leu Ser Phe Gly Ala Pro Leu Ala Asn Thr Tyr Lys Ile Val Val	2000	2005
Asp Glu Arg Glu Leu Leu Phe Glu Thr Ser Glu Val Val Asp Val	2015	2020
Ala Lys Leu Met Lys Ala Tyr Ile Ser Met Ile Val Lys Lys Arg	2030	2035
Tyr Ser Thr Thr Arg Ser Ala Ser Ser Gln Gly Ser Ser Arg	2045	2050

Table 11 hMX2 nucleotide sequence (SEQ ID NO:11)

agctagtatc ttttattgtc agaacttctg tgagccaaca aacagttttg catggttgta

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gcgagctcc	tgggtccgcc	gtcctctgcag	cttctgcagg	gaagcctcgg	tcagcgacag	6900
ctcctgctgc	tccttcacgc	gctgcaggte	ctcgatttct	ttctccagac	ggaggatctc	6960
ttccacctgc	ttattttcct	tctgtttctc	cagttcacgg	gtcagttcag	cttctctctg	7020
gctcttctgc	aaggcttcga	gttcttctg	cttctctgtt	tcttcttctc	gctgggcggg	7080
gagctcggct	tctctctgct	ctctctctct	ttctcttctc	tcttctctcc	gttctctctt	7140
ttcttctctc	tcctgtttct	tcttttcttc	ttgtctcctt	ttctctgcca	gcaattgtct	7200
gtaaactctc	cgagcaatct	gacctctgag	ttgcttctgg	aaaactatgg	ctgccttttt	7260
cagggtgcaa	aatctcctcc	tcagaaggaa	tgctctgtaa	ttcttctgta	ttatcaccac	7320
acaataaagg	acctttctgt	attgtttccg	tgctaagaag	ccaagacat	ggggccgaat	7380
caccatggcc	gcgtgggtca	cttctctctc	cctccgcttc	tcagtttct	gttccaagga	7440
ttctcgaaga	aataccttgg	tcttccccag	ctgccactcg	ctgttgagg	catcatagag	7500
ctgcagcagg	ctcgtgact	tcctctggac	gtcctcaggc	agagccagat	tcctcatcag	7560
cactttatac	cttttgtaaa	agtccctgaa	gggtcttcgg	accgcatacc	cagcttttgcg	7620
gattctcaca	gtctccagca	tcctctagta	ccgcagctgg	ttcagcacia	ccgcctgggtc	7680
aaactgggtc	ggcatcttct	gcatgtttgg	cttgatacag	cgaacaaaga	aaggattaga	7740
ggagcttagc	gttgccatta	aggaatgcag	tgagtcaacc	ttgaactgtg	agctgactgt	7800
aggccgccga	tgtttgcttc	cacatttcaa	ggatctctgg	ttgttgccgc	ttgaaacatg	7860
ttcaaaaaga	tcgtagataa	agtcaaaccg	gctttctctt	agcaaattga	gaaggctcatc	7920
tcgaaatgta	tctctgttct	tctccaagat	acctcgga	tcataattgca	cctctccage	7980

atagtgttcc	actccaaaat	tgttaactgc	aactctgggc	ttcacataaa	agtgggttatt	8040
cgcattgtga	ctgtgttagct	tctccaataa	gggtgtgtct	gtggcttgag	gaaaatggct	8100
ttcttcattg	ataagggcta	ggaggccaag	tttcttctca	atcaagtcca	ggcattctcc	8160
attgtctatc	cagtcaatat	cttcccacac	taatccttcc	ctgctatatt	ctagttgttc	8220
taaagaaaaa	atatgcttgt	tgaagtactc	ctgaagtttc	tcgtttgcat	agtttatatt	8280
gaactgttca	aagtgattaa	cctcaaagtt	ttcaaattcca	aagatgtcga	ggatgccaat	8340
agacttgaag	tcctcattgc	ctttgatcct	gctgttgatc	ttcttgatta	cccactcaaa	8400
gcagcacgca	tacagagcca	tggccaggga	gtccctgctg	tctactgcct	gttgaacatt	8460
gagaggcgtg	aggatctctt	ctcccctgag	gaacattgat	ctctgggtca	aagcatctgt	8520
gagctgtgtt	gggtccagcc	caagtaactc	cgcagatctg	cccaaagctg	ttttgaagga	8580
aacctgtgcc	ccaccagcag	tgataaatcc	tatgttccca	agatgcagta	taccagcaag	8640
cagcctcgac	acttcccga	cttctcctt	gctgaactgc	atcacgtcca	ttgccgta	8700
aacttcccta	aaggattcct	ggtcactgat	tgtcttgtct	tctacacatc	cagactgatt	8760
caagtagtgg	tagttttctg	gcgtagataa	ataaaattct	tctctttctt	catgtttccag	8820
ccctgccagc	agtgcataaa	atatgtgata	attcctttcc	ccgggatttt	gccttactac	8880
tcggttcttg	gaagagagga	tacaatctac	aattctcccg	ccctgaatat	ttcctttctg	8940
acagatgttc	agctgaacaa	acttcccaaa	gcgactagag	ttgttgttgt	acacgggtctt	9000
cgcattgccg	aaagcttcca	tgatggggct	gctttcaaga	atagctcggt	caacacagga	9060
tgtcttctcc	tttaaggaca	attccaaaga	ctgttgactg	atgactgaca	gaaacttgag	9120
gatcaattta	gtgctttcgg	ttttacctgc	cccactttca	cccttgatga	ggatgcactg	9180
gttgctgtgg	cgcttccaca	ggcagcggta	gcactcgttg	gcgatggcga	agatgtgcgg	9240
gggcagctcg	cccaggtggc	gccggctgta	ctgctccatg	gtggcaggct	cgtacagccc	9300
ggcgatgggc	tggtaggggt	tcacagaggg	caggatggag	ccgatgtagg	tccatatttg	9360
atttctctta	taccgctgga	ataagttata	catgatggag	ccgccatgga	gctctgtcaa	9420
ggacgccatg	tcattccacg	cctcctcggt	cgtgggggtg	atagcagtca	ccttctgggtg	9480
ggtaattgtg	ctctgcttgt	aagtgaatac	ctgaccatag	tctgtccgga	agacgacgat	9540
gccttctgca	caggaattta	cagtacttgg	aaaatgctgg	ccattttctc	tcagccagac	9600
ccgtgttccc	tgtaaacaaa	a				9621

Table 12 hMX2 polypeptide sequence (SEQ ID NO:12)

Phe	Cys	Leu	Gln	Gly	Thr	Arg	Val	Trp	Leu	Arg	Glu	Asn	Gly	Gln	His
1			5					10						15	
Phe	Pro	Ser	Thr	Val	Asn	Ser	Cys	Ala	Glu	Gly	Ile	Val	Val	Phe	Arg
		20					25					30			
Thr	Asp	Tyr	Gly	Gln	Val	Phe	Thr	Tyr	Lys	Gln	Ser	Thr	Ile	Thr	His
	35						40					45			
Gln	Lys	Val	Thr	Ala	Met	His	Pro	Thr	Asn	Glu	Glu	Gly	Val	Asp	Asp
	50					55				60					
Met	Ala	Ser	Leu	Thr	Glu	Leu	His	Gly	Gly	Ser	Ile	Met	Tyr	Asn	Leu
65				70					75					80	
Phe	Gln	Arg	Tyr	Lys	Arg	Asn	Gln	Ile	Trp	Thr	Tyr	Ile	Gly	Ser	Ile
			85					90					95		
Leu	Ala	Ser	Val	Asn	Pro	Tyr	Gln	Pro	Ile	Ala	Gly	Leu	Tyr	Glu	Pro

100							105					110				
Ala	Thr	Met	Glu	Gln	Tyr	Ser	Arg	Arg	His	Leu	Gly	Glu	Leu	Pro	Pro	
		115					120					125				
His	Ile	Phe	Ala	Ile	Ala	Asn	Glu	Cys	Tyr	Arg	Cys	Leu	Trp	Lys	Arg	
	130					135					140					
His	Asp	Asn	Gln	Cys	Ile	Leu	Ile	Lys	Gly	Glu	Ser	Gly	Ala	Gly	Lys	
145					150					155					160	
Thr	Glu	Ser	Thr	Lys	Leu	Ile	Leu	Lys	Phe	Leu	Ser	Val	Ile	Ser	Gln	
				165					170					175		
Gln	Ser	Leu	Glu	Leu	Ser	Leu	Lys	Glu	Lys	Thr	Ser	Cys	Val	Glu	Arg	
			180					185					190			
Ala	Ile	Leu	Glu	Ser	Ser	Pro	Ile	Met	Glu	Ala	Phe	Gly	Asn	Ala	Lys	
		195					200					205				
Thr	Val	Tyr	Asn	Asn	Asn	Ser	Ser	Arg	Phe	Gly	Lys	Phe	Val	Gln	Leu	
	210					215					220					
Asn	Ile	Cys	Gln	Lys	Gly	Asn	Ile	Gln	Gly	Gly	Arg	Ile	Val	Asp	Cys	
225					230					235					240	
Ile	Leu	Ser	Ser	Gln	Asn	Arg	Val	Val	Arg	Gln	Asn	Pro	Gly	Glu	Arg	
				245					250					255		
Asn	Tyr	His	Ile	Phe	Tyr	Ala	Leu	Leu	Ala	Gly	Leu	Glu	His	Glu	Glu	
			260					265					270			
Arg	Glu	Glu	Phe	Tyr	Leu	Ser	Thr	Pro	Glu	Asn	Tyr	His	Tyr	Leu	Asn	
		275					280					285				
Gln	Ser	Gly	Cys	Val	Glu	Asp	Lys	Thr	Ile	Ser	Asp	Gln	Glu	Ser	Phe	
	290					295					300					
Arg	Glu	Val	Ile	Thr	Ala	Met	Asp	Val	Met	Gln	Phe	Ser	Lys	Glu	Glu	
305					310					315					320	
Val	Arg	Glu	Val	Ser	Arg	Leu	Leu	Ala	Gly	Ile	Leu	His	Leu	Gly	Asn	
				325					330					335		
Ile	Glu	Phe	Ile	Thr	Ala	Gly	Gly	Ala	Gln	Val	Ser	Phe	Lys	Thr	Ala	
			340					345					350			
Leu	Gly	Arg	Ser	Ala	Glu	Leu	Leu	Gly	Leu	Asp	Pro	Thr	Gln	Leu	Thr	
		355					360					365				
Asp	Ala	Leu	Thr	Gln	Arg	Ser	Met	Phe	Leu	Arg	Gly	Glu	Glu	Ile	Leu	
	370					375					380					
Thr	Pro	Leu	Asn	Val	Gln	Gln	Ala	Val	Asp	Ser	Arg	Asp	Ser	Leu	Ala	
385					390					395					400	
Met	Ala	Leu	Tyr	Ala	Cys	Cys	Phe	Glu	Trp	Val	Ile	Lys	Lys	Ile	Asn	
				405					410					415		
Ser	Arg	Ile	Lys	Gly	Asn	Glu	Asp	Phe	Lys	Ser	Ile	Gly	Ile	Leu	Asp	
			420					425					430			
Ile	Phe	Gly	Phe	Glu	Asn	Phe	Glu	Val	Asn	His	Phe	Glu	Gln	Phe	Asn	
		435					440					445				
Ile	Asn	Tyr	Ala	Asn	Glu	Lys	Leu	Gln	Glu	Tyr	Phe	Asn	Lys	His	Ile	

450	455	460
Phe Ser Leu Glu Gln Leu Glu Tyr Ser Arg Glu Gly Leu Val Trp Glu 465 470 475 480		
Asp Ile Asp Trp Ile Asp Asn Gly Glu Cys Leu Asp Leu Ile Glu Lys 485 490 495		
Lys Leu Gly Leu Leu Ala Leu Ile Asn Glu Glu Ser His Phe Pro Gln 500 505 510		
Ala Thr Asp Ser Thr Leu Leu Glu Lys Leu His Ser Gln His Ala Asn 515 520 525		
Asn His Phe Tyr Val Lys Pro Arg Val Ala Val Asn Asn Phe Gly Val 530 535 540		
Lys His Tyr Ala Gly Glu Val Gln Tyr Asp Val Arg Gly Ile Leu Glu 545 550 555 560		
Lys Asn Arg Asp Thr Phe Arg Asp Asp Leu Leu Asn Leu Leu Arg Glu 565 570 575		
Ser Arg Phe Asp Phe Ile Tyr Asp Leu Phe Glu His Val Ser Ser Arg 580 585 590		
Asn Asn Gln Asp Thr Leu Lys Cys Gly Ser Lys His Arg Arg Pro Thr 595 600 605		
Val Ser Ser Gln Phe Lys Val Asp Ser Leu His Ser Leu Met Ala Thr 610 615 620		
Leu Ser Ser Ser Asn Pro Phe Phe Val Arg Cys Ile Lys Pro Asn Met 625 630 635 640		
Gln Lys Met Pro Asp Gln Phe Asp Gln Ala Val Val Leu Asn Gln Leu 645 650 655		
Arg Tyr Ser Gly Met Leu Glu Thr Val Arg Ile Arg Lys Ala Gly Tyr 660 665 670		
Ala Val Arg Arg Pro Phe Gln Asp Phe Tyr Lys Arg Tyr Lys Val Leu 675 680 685		
Met Arg Asn Leu Ala Leu Pro Glu Asp Val Arg Gly Lys Cys Thr Ser 690 695 700		
Leu Leu Gln Leu Tyr Asp Ala Ser Asn Ser Glu Trp Gln Leu Gly Lys 705 710 715 720		
Thr Lys Val Phe Leu Arg Glu Ser Leu Glu Gln Lys Leu Glu Lys Arg 725 730 735		
Arg Glu Glu Glu Val Ser His Ala Ala Met Val Ile Arg Ala His Val 740 745 750		
Leu Gly Phe Leu Ala Arg Lys Gln Tyr Arg Lys Val Leu Tyr Cys Val 755 760 765		
Val Ile Ile Gln Lys Asn Tyr Arg Ala Phe Leu Leu Arg Arg Arg Phe 770 775 780		
Leu His Leu Lys Lys Ala Ala Ile Val Phe Gln Lys Gln Leu Arg Gly 785 790 795 800		
Gln Ile Ala Arg Arg Val Tyr Arg Gln Leu Leu Ala Glu Lys Arg Glu		

805						810						815				
Gln	Glu	Glu	Lys	Lys	Lys	Gln	Glu	Glu	Glu	Glu	Lys	Lys	Lys	Arg	Glu	
			820						825					830		
Glu	Glu	Glu	Arg	Glu	Arg	Glu	Arg	Glu	Arg	Arg	Glu	Ala	Glu	Leu	Arg	
			835					840				845				
Ala	Gln	Gln	Glu	Glu	Glu	Thr	Arg	Lys	Gln	Gln	Glu	Leu	Glu	Ala	Leu	
			850			855						860				
Gln	Lys	Ser	Gln	Lys	Glu	Ala	Glu	Leu	Thr	Arg	Glu	Leu	Glu	Lys	Gln	
865					870					875					880	
Lys	Glu	Asn	Lys	Gln	Val	Glu	Glu	Ile	Leu	Arg	Leu	Glu	Lys	Glu	Ile	
				885					890					895		
Glu	Asp	Leu	Gln	Arg	Met	Lys	Glu	Gln	Gln	Glu	Leu	Ser	Leu	Thr	Glu	
			900					905						910		
Ala	Ser	Leu	Gln	Lys	Leu	Gln	Glu	Arg	Arg	Asp	Gln	Glu	Leu	Arg	Arg	
			915				920					925				
Leu	Glu	Glu	Glu	Ala	Cys	Arg	Ala	Ala	Gln	Glu	Phe	Leu	Glu	Ser	Leu	
			930			935						940				
Asn	Phe	Asp	Glu	Ile	Asp	Glu	Cys	Val	Arg	Asn	Ile	Glu	Arg	Ser	Leu	
945					950					955					960	
Ser	Gly	Gly	Ser	Glu	Phe	Ser	Ser	Glu	Leu	Ala	Glu	Ser	Ala	Cys	Glu	
				965					970					975		
Glu	Lys	Pro	Asn	Phe	Asn	Phe	Ser	Gln	Pro	Tyr	Pro	Glu	Glu	Glu	Val	
			980					985						990		
Asp	Glu	Gly	Phe	Glu	Ala	Asp	Asp	Asp	Ala	Phe	Lys	Asp	Ser	Pro	Asn	
			995				1000					1005				
Pro	Ser	Glu	His	Gly	His	Ser	Asp	Gln	Arg	Thr	Ser	Gly	Ile	Arg		
			1010			1015						1020				
Thr	Ser	Asp	Asp	Ser	Ser	Glu	Glu	Asp	Pro	Tyr	Met	Asn	Asp	Thr		
			1025			1030						1035				
Val	Val	Pro	Thr	Ser	Pro	Ser	Ala	Asp	Ser	Thr	Val	Leu	Leu	Ala		
			1040			1045						1050				
Pro	Ser	Val	Gln	Asp	Ser	Gly	Ser	Leu	His	Asn	Ser	Ser	Ser	Gly		
			1055			1060						1065				
Glu	Ser	Thr	Tyr	Cys	Met	Pro	Gln	Asn	Ala	Gly	Asp	Leu	Pro	Ser		
			1070			1075						1080				
Pro	Asp	Gly	Asp	Tyr	Asp	Tyr	Asp	Gln	Asp	Asp	Tyr	Glu	Asp	Gly		
			1085			1090						1095				
Ala	Ile	Thr	Ser	Gly	Ser	Ser	Val	Thr	Phe	Ser	Asn	Ser	Tyr	Gly		
			1100			1105						1110				
Ser	Gln	Trp	Ser	Pro	Asp	Tyr	Arg	Cys	Ser	Val	Gly	Thr	Tyr	Asn		
			1115			1120						1125				
Ser	Ser	Gly	Ala	Tyr	Arg	Phe	Ser	Ser	Glu	Gly	Ala	Gln	Ser	Ser		
			1130			1135						1140				
Phe	Glu	Asp	Ser	Glu	Glu	Asp	Phe	Asp	Ser	Arg	Phe	Asp	Thr	Asp		

1145	1150	1155
Asp Glu Leu Ser Tyr Arg Arg	Asp Ser Val Tyr Ser	Cys Val Thr
1160	1165	1170
Leu Pro Tyr Phe His Ser Phe	Leu Tyr Met Lys Gly	Gly Leu Met
1175	1180	1185
Asn Ser Trp Lys Arg Arg Trp	Cys Val Leu Lys Asp	Glu Thr Phe
1190	1195	1200
Leu Trp Phe Arg Ser Lys Gln	Glu Ala Leu Lys Gln	Gly Trp Leu
1205	1210	1215
His Lys Lys Gly Gly Gly Ser	Ser Thr Leu Ser Arg	Arg Asn Trp
1220	1225	1230
Lys Lys Arg Trp Phe Val Leu	Arg Gln Ser Lys Leu	Met Tyr Phe
1235	1240	1245
Glu Asn Asp Ser Glu Glu Lys	Leu Lys Gly Thr Val	Glu Val Arg
1250	1255	1260
Thr Ala Lys Glu Ile Ile Asp	Asn Thr Thr Lys Glu	Asn Gly Ile
1265	1270	1275
Asp Ile Ile Met Ala Asp Arg	Thr Phe His Leu Ile	Ala Glu Ser
1280	1285	1290
Pro Glu Asp Ala Ser Gln Trp	Phe Ser Val Leu Ser	Gln Val His
1295	1300	1305
Ala Ser Thr Asp Gln Glu Ile	Gln Glu Met His Asp	Glu Gln Ala
1310	1315	1320
Asn Pro Gln Asn Ala Val Gly	Thr Leu Asp Val Gly	Leu Ile Asp
1325	1330	1335
Ser Val Cys Ala Ser Asp Ser	Pro Asp Arg Pro Asn	Ser Phe Val
1340	1345	1350
Ile Ile Thr Ala Asn Arg Val	Leu His Cys Asn Ala	Asp Thr Pro
1355	1360	1365
Glu Glu Met His His Trp Ile	Thr Leu Leu Gln Arg	Ser Lys Gly
1370	1375	1380
Asp Thr Arg Val Glu Gly Gln	Glu Phe Ile Val Arg	Gly Trp Leu
1385	1390	1395
His Lys Glu Val Lys Asn Ser	Pro Lys Met Ser Ser	Leu Lys Leu
1400	1405	1410
Lys Lys Arg Trp Phe Val Leu	Thr His Asn Ser Leu	Asp Tyr Tyr
1415	1420	1425
Lys Ser Ser Glu Lys Asn Ala	Leu Lys Leu Gly Thr	Leu Val Leu
1430	1435	1440
Asn Ser Leu Cys Ser Val Val	Pro Pro Asp Glu Lys	Ile Phe Lys
1445	1450	1455
Glu Thr Gly Tyr Trp Asn Val	Thr Val Tyr Gly Arg	Lys His Cys
1460	1465	1470
Tyr Arg Leu Tyr Thr Lys Leu	Leu Asn Glu Ala Thr	Arg Trp Ser

1475					1480					1485				
Ser	Val	Ile	Gln	Asn	Val	Thr	Asp	Thr	Lys	Ala	Pro	Ile	Asp	Thr
1490						1495					1500			
Pro	Thr	Gln	Gln	Leu	Ile	Gln	Asp	Ile	Lys	Glu	Asn	Cys	Leu	Asn
1505						1510					1515			
Ser	Asp	Val	Val	Glu	Gln	Ile	Tyr	Lys	Arg	Asn	Pro	Ile	Leu	Arg
1520						1525					1530			
Tyr	Thr	His	His	Pro	Leu	His	Ser	Pro	Leu	Leu	Pro	Leu	Pro	Tyr
1535						1540					1545			
Gly	Asp	Ile	Asn	Leu	Asn	Leu	Leu	Lys	Asp	Lys	Gly	Tyr	Thr	Thr
1550						1555					1560			
Leu	Gln	Asp	Glu	Ala	Ile	Lys	Ile	Phe	Asn	Ser	Leu	Gln	Gln	Leu
1565						1570					1575			
Glu	Ser	Met	Ser	Asp	Pro	Ile	Pro	Ile	Ile	Gln	Gly	Ile	Leu	Gln
1580						1585					1590			
Thr	Gly	His	Asp	Leu	Arg	Pro	Leu	Arg	Asp	Glu	Leu	Tyr	Cys	Gln
1595						1600					1605			
Leu	Ile	Lys	Gln	Thr	Asn	Lys	Val	Pro	His	Pro	Gly	Ser	Val	Gly
1610						1615					1620			
Asn	Leu	Tyr	Ser	Trp	Gln	Ile	Leu	Thr	Cys	Leu	Ser	Cys	Thr	Phe
1625						1630					1635			
Leu	Pro	Ser	Arg	Gly	Ile	Leu	Lys	Tyr	Leu	Lys	Phe	His	Leu	Lys
1640						1645					1650			
Arg	Ile	Arg	Glu	Gln	Phe	Pro	Gly	Thr	Glu	Met	Glu	Lys	Tyr	Ala
1655						1660					1665			
Leu	Phe	Thr	Tyr	Glu	Ser	Leu	Lys	Lys	Thr	Lys	Cys	Arg	Glu	Phe
1670						1675					1680			
Val	Pro	Ser	Arg	Asp	Glu	Ile	Glu	Ala	Leu	Ile	His	Arg	Gln	Glu
1685						1690					1695			
Met	Thr	Ser	Thr	Val	Tyr	Cys	His	Gly	Gly	Gly	Ser	Cys	Lys	Ile
1700						1705					1710			
Thr	Ile	Asn	Ser	His	Thr	Thr	Ala	Gly	Glu	Val	Val	Glu	Lys	Leu
1715						1720					1725			
Ile	Arg	Gly	Leu	Ala	Met	Glu	Asp	Ser	Arg	Asn	Met	Phe	Ala	Leu
1730						1735					1740			
Phe	Glu	Tyr	Asn	Gly	His	Val	Asp	Lys	Ala	Ile	Glu	Ser	Arg	Thr
1745						1750					1755			
Val	Val	Ala	Asp	Val	Leu	Ala	Lys	Phe	Glu	Lys	Leu	Ala	Ala	Thr
1760						1765					1770			
Ser	Glu	Val	Gly	Asp	Leu	Pro	Trp	Lys	Phe	Tyr	Phe	Lys	Leu	Tyr
1775						1780					1785			
Cys	Phe	Leu	Asp	Thr	Asp	Asn	Val	Pro	Lys	Asp	Ser	Val	Glu	Phe
1790						1795					1800			
Ala	Phe	Met	Phe	Glu	Gln	Ala	His	Glu	Ala	Val	Ile	His	Gly	His

1805	1810	1815
His Pro Ala Pro Glu Glu Asn Leu Gln Val Leu Ala Ala Leu Arg		
1820	1825	1830
Leu Gln Tyr Leu Gln Gly Asp Tyr Thr Leu His Ala Ala Ile Pro		
1835	1840	1845
Pro Leu Glu Glu Val Tyr Ser Leu Gln Arg Leu Lys Ala Arg Ile		
1850	1855	1860
Ser Gln Ser Thr Lys Thr Phe Thr Pro Cys Glu Arg Leu Glu Lys		
1865	1870	1875
Arg Arg Thr Ser Phe Leu Glu Gly Thr Leu Arg Arg Ser Phe Arg		
1880	1885	1890
Thr Gly Ser Val Val Arg Gln Lys Val Glu Glu Glu Gln Met Leu		
1895	1900	1905
Asp Met Trp Ile Lys Glu Glu Val Ser Ser Ala Arg Ala Ser Ile		
1910	1915	1920
Ile Asp Lys Trp Arg Lys Phe Gln Gly Met Asn Gln Glu Gln Ala		
1925	1930	1935
Met Ala Lys Tyr Met Ala Leu Ile Lys Glu Trp Pro Gly Tyr Gly		
1940	1945	1950
Ser Thr Leu Phe Asp Val Glu Val Arg Thr Gly Cys His Val Leu		
1955	1960	1965
Gly Trp Ala Gly Cys Trp His Leu Arg Thr Trp Ile Thr Ala Lys		
1970	1975	1980
Phe Met Trp Arg Glu Asp Lys Met Glu His Phe Ala Leu Ser Thr		
1985	1990	1995
Ser Phe Phe Arg Ala Pro Lys Ile Val Pro Leu Thr Pro Pro Phe		
2000	2005	2010
Ser Ser Gln Phe Leu Phe Ser Cys Val Val Asn Ala Ser Val Ile		
2015	2020	2025
Leu Gly Met Asn Ala Lys Leu Arg Cys His Leu Phe Phe Tyr Pro		
2030	2035	2040
Ser Leu Gly Lys Leu		
2045		

Table 13 hMP nucleotide sequence (SEQ ID NO:13)

ccaacttttg cagctccacc caggatgtgg cctcgctcca cccagctgt gcgcctctct	60
ccacccttag gcgaaggcac tagaatttcc caaattaaga acgaagagga agtttggacc	120
ttttcggcca ccgctcgctt caatatggct gccccaggg agagacgagg ctaccatgaa	180
ggagccgagc gcagaccctg agtccgtcac cc <u>atgg</u> atcg cagcgcgag ttcaggaaat	240
ggaaggcgca atgtttgagc aaagcggacc tcagccggaa gggcagtgtt gacgaggatg	300
tggtagagct tgtgcagttt ctgaacatgc gagatcagtt tttcaccacc agtccttctg	360
ctggccgcat cctactcctt gaccggggta taaatggttt tgaggttcag aaacaaaact	420
gttgctggct actggttaca cacaaacttt gtgtaaaaga tgatgtgatt gtagctctga	480

agaaagcaaa	tggtgatgcc	actttgaaat	ttgaaccatt	tgttcttcat	gtgcagtgtc	540
gacaattgca	ggatgcacag	attctgcatt	ccatggcaat	agattctggt	ttcaggaact	600
ctggcataac	ggtgggaaag	agaggaaaaa	ctatgttggc	tgtccggagt	acacatggct	660
tagaagttcc	attaagccat	aagggaatac	tgatggtgac	agaggaatat	attgacttcc	720
tgttaaatgt	ggcaaataca	aaaatggagg	aaaacaagaa	aagaattgag	aggttttaca	780
actgcctaca	gcatgctttg	gaaagggaaa	cgatgactaa	cttacatccc	aagatcaaa	840
agaaaaataa	ctcatcatat	attcataaga	aaaaaagaaa	cccagaaaaa	acacgtgcc	900
agtgtattac	taaagaaagt	gatgaagaac	ttgaaaatga	tgatgatgat	gatctaggaa	960
tcaatgttac	catcttccct	gaagattact	aagctttggt	tctgatgtgt	cttggccgta	1020
atgtttctag	taggttttat	aaagctgctc	ttcataagag	tatttttagtt	tgttgagtgt	1080
atcagccatt	cataagccag	taatgacaag	tcagagactt	caaactataa	ctttgttgcc	1140
cagaggatgt	gcagttgtca	tctaagctct	cagcagtacc	cggttatcc	tacgacttca	1200
cctgaaatgc	tatagttatc	cctacttttt	taccagtttc	tcccagaagc	acctgcttaa	1260
taaatcaaag	atgtttgaaa	aaaaaaaa				1288

Table 14 hMP polypeptide sequence (SEQ ID NO:14)

Met	Asp	Arg	Ser	Ala	Glu	Phe	Arg	Lys	Trp	Lys	Ala	Gln	Cys	Leu	Ser	1	5	10	15
Lys	Ala	Asp	Leu	Ser	Arg	Lys	Gly	Ser	Val	Asp	Glu	Asp	Val	Val	Glu	20	25	30	
Leu	Val	Gln	Phe	Leu	Asn	Met	Arg	Asp	Gln	Phe	Phe	Thr	Thr	Ser	Ser	35	40	45	
Phe	Ala	Gly	Arg	Ile	Leu	Leu	Leu	Asp	Arg	Gly	Ile	Asn	Gly	Phe	Glu	50	55	60	
Val	Gln	Lys	Gln	Asn	Cys	Cys	Trp	Leu	Leu	Val	Thr	His	Lys	Leu	Cys	65	70	75	80
Val	Lys	Asp	Asp	Val	Ile	Val	Ala	Leu	Lys	Lys	Ala	Asn	Gly	Asp	Ala	85	90	95	
Thr	Leu	Lys	Phe	Glu	Pro	Phe	Val	Leu	His	Val	Gln	Cys	Arg	Gln	Leu	100	105	110	
Gln	Asp	Ala	Gln	Ile	Leu	His	Ser	Met	Ala	Ile	Asp	Ser	Gly	Phe	Arg	115	120	125	
Asn	Ser	Gly	Ile	Thr	Val	Gly	Lys	Arg	Gly	Lys	Thr	Met	Leu	Ala	Val	130	135	140	
Arg	Ser	Thr	His	Gly	Leu	Glu	Val	Pro	Leu	Ser	His	Lys	Gly	Lys	Leu	145	150	155	160
Met	Val	Thr	Glu	Glu	Tyr	Ile	Asp	Phe	Leu	Leu	Asn	Val	Ala	Asn	Gln	165	170	175	
Lys	Met	Glu	Glu	Asn	Lys	Lys	Arg	Ile	Glu	Arg	Phe	Tyr	Asn	Cys	Leu	180	185	190	
Gln	His	Ala	Leu	Glu	Arg	Glu	Thr	Met	Thr	Asn	Leu	His	Pro	Lys	Ile	195	200	205	
Lys	Glu	Lys	Asn	Asn	Ser	Ser	Tyr	Ile	His	Lys	Lys	Lys	Arg	Asn	Pro				

210	215	220
Glu Lys Thr Arg Ala Gln Cys Ile Thr Lys Glu Ser Asp Glu Glu Leu		
225	230	235 240
Glu Asn Asp Asp Asp Asp Asp Leu Gly Ile Asn Val Thr Ile Phe Pro		
	245	250 255
Glu Asp Tyr		

Table 15 NHR nucleotide sequence (SEQ ID NO:15)

acgcgtgcag	gtggcgtggc	gccagggatt	tgaaccgcgc	tgacgaagtt	tggtgatcca	60
tcttccgagt	atcgccggga	tttcgaatcg	cgatgatcat	cccctctcta	gaggagctgg	120
actccctcaa	gtacagtgac	ctgcagaact	tagccaagag	tctgggtctc	cgggcccaacc	180
tgagggcaac	caagttgtta	aaagccttga	aaggctacat	taaacatgag	gcaagaaaag	240
gaaatgagaa	tcaggatgaa	agtcaaactt	ctgcatcctc	ttgtgatgag	actgagatac	300
agatcagcaa	ccaggaagag	ctgagagaca	gccacttggc	catgtcacca	aaacaaggag	360
aagggtgcaag	actgtccgtg	tggaccctga	ctcacagaga	atcattcaga	gataaaaata	420
agtaatccca	ctgaattcca	gaatcatgaa	aagcaggaaa	gccaggatct	cagagcactg	480
caaaagttcc	ttctccacca	gacgagcacc	aagaagctga	gaatgctgtt	tcctcaggta	540
acagagattc	aaaggtacct	tcagaaggaa	agaaatctct	ctacacagat	gagtcattcca	600
aacctggaaa	aaataaaaaga	actgcaatca	ctactccaaa	ctttaagaag	cttcatgaag	660
ctcattttta	ggaaatggag	tccattgatc	caatatatng	aggagaaaaa	aagaaacatt	720
ttgaagaaca	caattccatg	aatgaactga	agcagccgcc	catcaataag	ggaggggtca	780
ggactccagt	acctccaaga	ggaagactct	ctgtggcttc	tactcccata	agccaacgac	840
gctcgcaagg	cgggtcttgt	ggccctgcaa	gtcagagtac	cttgggtctg	aaggggtcac	900
tcaagcgctc	tgtatctctc	gcagctaaaa	cgggtgtcag	gttttcagct	gctactaaag	960
ataatgagca	taagcgttca	ctgaccaaga	ctccagccag	aaagtctgca	catgtgaccg	1020
tgtctggggg	cacccaaaaa	ggcgaggctg	tgcttgggac	acacaaatta	aagaccatca	1080
cggggaattc	tgctgctgtt	attaccccat	tcaagttgac	aactgaggca	acgcgactc	1140
cagtctccaa	taagaaacca	gtgtttgatc	ttaaagcaag	tttgtctcgt	cccctcaact	1200
atgaaccaca	caaaggaaag	ctaaaaccat	gggggcaatc	taaagaaaat	aattatctaa	1260
atcaacatgt	caacaaatta	acttctacaa	gaaaacttac	aaacaacccc	atctccagac	1320
aaaggaagag	caacggaaga	aacgcgagca	agaagaaagg	agaagaaagc	aaagggtttg	1380
ggaatgcgaa	ggggcctcat	tttggttgaa	gattaataat	tttttaacat	cttgtaaata	1440
ttcctgtatt	ctcaactttt	ttccttttgt	aaattttttt	tttttgctgt	catccccact	1500
ttagtcacga	gatctttttc	tgctaactgt	tcatagtctg	tgtagtgtcc	atgggttctt	1560
catgtgctat	gatctctgaa	aagacgttat	caccttaaag	ctcaaattct	ttgggatggt	1620
ttttacttaa	gtccattaac	aattcagggt	tctaacgaga	cccatcctaa	aattctcttt	1680
ctagtttttt	aatgtcacca	tcccaaactc	cgttttctgg	atttttaatc	cccagctccc	1740
cagttccctc	ttatcgtact	aatattaaca	gaactgcagt	cttctgctag	ccaatagcat	1800
ttacctgatg	gcagctagtt	atgcaagctt	caggagaatt	tgaacaataa	caagaatagg	1860
gtaagctggg	atagaaaggc	cacctcttca	ctctctatag	aatatagtaa	cctttatgaa	1920
acggggccat	atagtttggg	tatgacatca	atattttacc	taggtgaaat	tgtttaggct	1980
tatgtacctt	cgttcaaata	tcctcatgta	attgccatct	gtcactcact	atattcacia	2040

aaataaaaact ctacaactca ttctaacatt gcttacttaa aagctacata gccctatcga	2100
aatgcgagga ttaatgctttt aatgctttta gagacagggt ctactgtgtg tgcccaggct	2160
ggctctcaaac tccaccaa at gtactttctta ttcatTTTTat ggaaaagact aggctttgct	2220
tagtatcatg tccatgtttc cttcacctca gtggagcttc tgagttttat actgctcaag	2280
atcgtcataa ataaaatttt ttctcattgt caaaaaaaaa aaaaaaaaaa aaaaaaaaaa	2340
aaaaaaaaaa aa	2352

Table 16 NHR polypeptide sequence (SEQ ID NO:16)

Met	Ile	Ile	Pro	Ser	Leu	Glu	Glu	Leu	Asp	Ser	Leu	Lys	Tyr	Ser	Asp	
1				5					10					15		
Leu	Gln	Asn	Leu	Ala	Lys	Ser	Leu	Gly	Leu	Arg	Ala	Asn	Leu	Arg	Ala	
			20					25					30			
Thr	Lys	Leu	Leu	Lys	Ala	Leu	Lys	Gly	Tyr	Ile	Lys	His	Glu	Ala	Arg	
			35				40					45				
Lys	Gly	Asn	Glu	Asn	Gln	Asp	Glu	Ser	Gln	Thr	Ser	Ala	Ser	Ser	Cys	
	50					55					60					
Asp	Glu	Thr	Glu	Ile	Gln	Ile	Ser	Asn	Gln	Glu	Glu	Ala	Glu	Arg	Gln	
65					70					75					80	
Pro	Leu	Gly	His	Val	Thr	Lys	Thr	Arg	Arg	Arg	Cys	Lys	Thr	Val	Arg	
				85					90					95		
Val	Asp	Pro	Asp	Ser	Gln	Gln	Asn	His	Ser	Glu	Ile	Lys	Ile	Ser	Asn	
			100					105					110			
Pro	Thr	Glu	Phe	Gln	Asn	His	Glu	Lys	Gln	Glu	Ser	Gln	Asp	Leu	Arg	
		115					120					125				
Ala	Thr	Ala	Lys	Val	Pro	Ser	Pro	Pro	Asp	Glu	His	Gln	Glu	Ala	Glu	
		130				135					140					
Asn	Ala	Val	Ser	Ser	Gly	Asn	Arg	Asp	Ser	Lys	Val	Pro	Ser	Glu	Gly	
145					150					155					160	
Lys	Lys	Ser	Leu	Tyr	Thr	Asp	Glu	Ser	Ser	Lys	Pro	Gly	Lys	Asn	Lys	
			165						170					175		
Arg	Thr	Ala	Ile	Thr	Thr	Pro	Asn	Phe	Lys	Lys	Leu	His	Glu	Ala	His	
			180					185					190			
Phe	Lys	Glu	Met	Glu	Ser	Ile	Asp	Pro	Ile	Tyr	Xaa	Gly	Glu	Lys	Lys	
		195					200					205				
Lys	His	Phe	Glu	Glu	His	Asn	Ser	Met	Asn	Glu	Leu	Lys	Gln	Pro	Pro	
	210					215					220					
Ile	Asn	Lys	Gly	Gly	Val	Arg	Thr	Pro	Val	Pro	Pro	Arg	Gly	Arg	Leu	
225					230					235					240	
Ser	Val	Ala	Ser	Thr	Pro	Ile	Ser	Gln	Arg	Arg	Ser	Gln	Gly	Arg	Ser	
			245						250				255			
Cys	Gly	Pro	Ala	Ser	Gln	Ser	Thr	Leu	Gly	Leu	Lys	Gly	Ser	Leu	Lys	
		260						265				270				
Arg	Ser	Ala	Ile	Ser	Ala	Ala	Lys	Thr	Gly	Val	Arg	Phe	Ser	Ala	Ala	

275					280					285					
Thr	Lys	Asp	Asn	Glu	His	Lys	Arg	Ser	Leu	Thr	Lys	Thr	Pro	Ala	Arg
	290					295					300				
Lys	Ser	Ala	His	Val	Thr	Val	Ser	Gly	Gly	Thr	Gln	Lys	Gly	Glu	Ala
305					310					315					320
Val	Leu	Gly	Thr	His	Lys	Leu	Lys	Thr	Ile	Thr	Gly	Asn	Ser	Ala	Ala
				325					330					335	
Val	Ile	Thr	Pro	Phe	Lys	Leu	Thr	Thr	Glu	Ala	Thr	Gln	Thr	Pro	Val
			340					345					350		
Ser	Asn	Lys	Lys	Pro	Val	Phe	Asp	Leu	Lys	Ala	Ser	Leu	Ser	Arg	Pro
		355					360					365			
Leu	Asn	Tyr	Glu	Pro	His	Lys	Gly	Lys	Leu	Lys	Pro	Trp	Gly	Gln	Ser
	370					375					380				
Lys	Glu	Asn	Asn	Tyr	Leu	Asn	Gln	His	Val	Asn	Arg	Ile	Asn	Phe	Tyr
385					390					395					400
Lys	Lys	Thr	Tyr	Lys	Gln	Pro	His	Leu	Gln	Thr	Lys	Glu	Glu	Gln	Arg
				405					410					415	
Lys	Lys	Arg	Glu	Gln	Glu	Arg	Lys	Glu	Lys	Lys	Ala	Lys	Val	Leu	Gly
			420					425					430		
Met	Arg	Arg	Gly	Leu	Ile	Leu	Ala	Glu	Asp						
			435				440								

Table 17 displays alignment of hMX1, hMX2 with human myosin (SEQ ID NO:31; GenBank AF247457) (Berg *et al.*, 2000). As seen from the alignment, hMX1 and hMX2 have a likely N-terminus of M N D residues. One of skill in the art can easily verify this observation by probing cDNA or genomic human libraries, or PCR techniques, to acquire the full length polynucleotide sequence.

Table 17 Alignment of hMX1, hMX2 and human myosin X

10	1	---FCLQGTRVWLRENGQHFPSTVNSCAEGIVVFR	TDYGQVFTYKQSTIT
12	1	---FCLQGTRVWLRENGQHFPSTVNSCAEGIVVFR	TDYGQVFTYKQSTIT
humX	1	MDNFFTEGTRVWLRENGQHFPSTVNSCAEGIVVFR	TDYGQVFTYKQSTIT
10	48	HQKVTAMHPTNEEGVDDMASLTELHGGSIMYNLFQRYKRNQIW	TYIGSIL
12	48	HQKVTAMHPTNEEGVDDMASLTELHGGSIMYNLFQRYKRNQIW	TYIGSIL
humX	51	HQKVTAMHPTNEEGVDDMASLTELHGGSIMYNLFQRYKRNQI	YTYIGSIL
10	98	ASVNPYQPIAGLYEPATMEQYSRRHLGELPPHIFAIANECYRCLWKR	HDN
12	98	ASVNPYQPIAGLYEPATMEQYSRRHLGELPPHIFAIANECYRCLWKR	HDN
humX	101	ASVNPYQPIAGLYEPATMEQYSRRHLGELPPHIFAIANECYRCLWK	RYDN
10	148	QCILIKGESGAGKTESTKLILKFLSVISQQSLELSLKEKTSCVERA	ILES
12	148	QCILIKGESGAGKTESTKLILKFLSVISQQSLELSLKEKTSCVERA	ILES
humX	151	QCILISGESGAGKTESTKLILKFLSVISQQSLELSLKEKTSCVERA	ILES
10	198	SPIMEAFGNAKTVYNNSSRFGKFVQLNICQKGNIQGGRIVDCILSSQ	NR

12	198	SPIMEAFGNAKTVYNNSSRFQKQVQLNICQKGNIQGGRIVDCILSSQNR
humX	201	SPIMEAFGNAKTVYNNSSRFQKQVQLNICQKGNIQGGRIVDYLLK-KNR
10	248	VVRQNPGERNYHIFYALLAGLEHEEREFFYLSTPENYHYLNQSGCVEDKT
12	248	VVRQNPGERNYHIFYALLAGLEHEEREFFYLSTPENYHYLNQSGCVEDKT
humX	250	VVRQNPGERNYHIFYALLAGLEHEEREFFYLSTPENYHYLNQSGCVEDKT
10	298	ISDQESFREVITAMDVMQFSKEEVREVSRLLAGILHLGNIEFITAGGAQV
12	298	ISDQESFREVITAMDVMQFSKEEVREVSRLLAGILHLGNIEFITAGGAQV
humX	300	ISDQESFREVITAMDVMQFSKEEVREVSRLLAGILHLGNIEFITAGGAQV
10	348	SFKTALGRSAELLGLDPTQLTDALTQSRMFLRGEEILTPLNVQQAVDSRD
12	348	SFKTALGRSAELLGLDPTQLTDALTQSRMFLRGEEILTPLNVQQAVDSRD
humX	350	SFKTALGRSAELLGLDPTQLTDALTQSRMFLRGEEILTPLNVQQAVDSRD
10	398	SLAMALYACCFEWIKKINSRIKGNEDFKSIGILDIFGFENFEVNHFEQF
12	398	SLAMALYACCFEWIKKINSRIKGNEDFKSIGILDIFGFENFEVNHFEQF
humX	400	SLAMALYACCFEWIKKINSRIKGNEDFKSIGILDIFGFENFEVNHFEQF
10	448	NINYANEKLQEYFNKHIFSLEQLEYSREGLVWEDIDWIDNGECLDLIEKK
12	448	NINYANEKLQEYFNKHIFSLEQLEYSREGLVWEDIDWIDNGECLDLIEKK
humX	450	NINYANEKLQEYFNKHIFSLEQLEYSREGLVWEDIDWIDNGECLDLIEKK
10	498	LGLLALINEESHFPQATDSTLLEKLHSQHANNHFYVKPRVAVNNFGVKHY
12	498	LGLLALINEESHFPQATDSTLLEKLHSQHANNHFYVKPRVAVNNFGVKHY
humX	500	LGLLALINEESHFPQATDSTLLEKLHSQHANNHFYVKPRVAVNNFGVKHY
10	548	AGEVQYDVRGILEKNRDTFRDILLNLLRESRFDIFYDLFEHVSSRNQDT
12	548	AGEVQYDVRGILEKNRDTFRDILLNLLRESRFDIFYDLFEHVSSRNQDT
humX	550	AGEVQYDVRGILEKNRDTFRDILLNLLRESRFDIFYDLFEHVSSRNQDT
10	598	LKCGSKHRRPTVSSQFQVDSLHSLMATLSSSNPFFVRCIKPNMQKMPDQF
12	598	LKCGSKHRRPTVSSQFQVDSLHSLMATLSSSNPFFVRCIKPNMQKMPDQF
humX	600	LKCGSKHRRPTVSSQFQVDSLHSLMATLSSSNPFFVRCIKPNMQKMPDQF
10	648	DQAVVLNQLRYSGLMETVRIRKAGYAVRRPFQDFYKRYKVLMRNLALPED
12	648	DQAVVLNQLRYSGLMETVRIRKAGYAVRRPFQDFYKRYKVLMRNLALPED
humX	649	DQAVVLNQLRYSGLMETVRIRKAGYAVRRPFQDFYKRYKVLMRNLALPED
10	698	VRGKCTSLQLYDASNSEWQLGKTKVFLRESLEQKLEKRREEEVSHAAMV
12	698	VRGKCTSLQLYDASNSEWQLGKTKVFLRESLEQKLEKRREEEVSHAAMV
humX	699	VRGKCTSLQLYDASNSEWQLGKTKVFLRESLEQKLEKRREEEVSHAAMV
10	748	IRAHVLGFLARKQYRKVLYCVVIIQKNYRAFLRRRFLHLKKAIVFQKQ
12	748	IRAHVLGFLARKQYRKVLYCVVIIQKNYRAFLRRRFLHLKKAIVFQKQ
humX	749	IRAHVLGFLARKQYRKVLYCVVIIQKNYRAFLRRRFLHLKKAIVFQKQ
10	798	LRGQIARRVYRQLLAEKREQEKKKQEEEEKKKREEREREREREREAEL
12	798	LRGQIARRVYRQLLAEKREQEKKKQEEEEKKKREEREREREREREAEL
humX	799	LRGQIARRVYRQLLAEKREQEKKKQEEEEKKKREEREREREREREAEL
10	848	RAQQEEETRQKQLEALQKSQKEAELTRELEKQKENKQVEEILRLEKEIE
12	848	RAQQEEETRQKQLEALQKSQKEAELTRELEKQKENKQVEEILRLEKEIE
humX	849	RAQQEEETRQKQLEALQKSQKEAELTRELEKQKENKQVEEILRLEKEIE
10	898	DLQRMKEQQELSLTEASLQKLQERRDQELRRLEEEACRAAQEFLESNFD
12	898	DLQRMKEQQELSLTEASLQKLQERRDQELRRLEEEACRAAQEFLESNFD
humX	899	DLQRMKEQQELSLTEASLQKLQERRDQELRRLEEEACRAAQEFLESNFD
10	948	EIDECVRNIERSLSGGSEFSSELAESACEEKPNFNFSQPYPEEEVDEGFE
12	948	EIDECVRNIERSLSGGSEFSSELAESACEEKPNFNFSQPYPEEEVDEGFE
humX	949	EIDECVRNIERSLSVGSEFSSELAESACEEKPNFNFSQPYPEEEVDEGFE
10	998	ADDDAFKDSPNPSEHGSDQRTSGIRTSDDSEEDPYMNDTVVPTSPSAD
12	998	ADDDAFKDSPNPSEHGSDQRTSGIRTSDDSEEDPYMNDTVVPTSPSAD
humX	999	ADDDAFKDSPNPSEHGSDQRTSGIRTSDDSEEDPYMNDTVVPTSPSAD

10	1048	STVLLAPSVQDSGSLHNSSSGESTYCMPQNAGDLPSPDGDYDYDQDDYED
12	1048	STVLLAPSVQDSGSLHNSSSGESTYCMPQNAGDLPSPDGDYDYDQDDYED
humX	1049	STVLLAPSVQDSGSLHNSSSGESTYCMPQNAGDLPSPDGDYDYDQDDYED
10	1098	GAITSGSSVTFNSYGSQWSPDYRCVGTYNSSGAYRFSSEGAQSSFEDS
12	1098	GAITSGSSVTFNSYGSQWSPDYRCVGTYNSSGAYRFSSEGAQSSFEDS
humX	1099	GAITSGSSVTFNSYGSQWSPDYRCVGTYNSSGAYRFSSEGAQSSFEDS
10	1148	EEDFDSRFDTDDELSYRRDSVYSCVTLPYFHSFLYMKGGLMNSWKRRWCV
12	1148	EEDFDSRFDTDDELSYRRDSVYSCVTLPYFHSFLYMKGGLMNSWKRRWCV
humX	1149	EEDFDSRFDTDDELSYRRDSVYSCVTLPYFHSFLYMKGGLMNSWKRRWCV
10	1198	LKDETFLLWFRSKQEALKQGWLHKKGGGSSTLSRRNWKRWVFLRQSKLMY
12	1198	LKDETFLLWFRSKQEALKQGWLHKKGGGSSTLSRRNWKRWVFLRQSKLMY
humX	1199	LKDETFLLWFRSKQEALKQGWLHKKGGGSSTLSRRNWKRWVFLRQSKLMY
10	1248	FENDSEEKLGKTVEVRTAKEIIDNTTKENGIDIIMADRTFHLLIAESPEDA
12	1248	FENDSEEKLGKTVEVRTAKEIIDNTTKENGIDIIMADRTFHLLIAESPEDA
humX	1249	FENDSEEKLGKTVEVRTAKEIIDNTTKENGIDIIMADRTFHLLIAESPEDA
10	1298	SQWFSVLSQVHASTDQEIQEMHDEQANPQNAVGTLDVGLIDSVCASDSPD
12	1298	SQWFSVLSQVHASTDQEIQEMHDEQANPQNAVGTLDVGLIDSVCASDSPD
humX	1299	SQWFSVLSQVHASTDQEIQEMHDEQANPQNAVGTLDVGLIDSVCASDSPD
10	1348	RPNSFVIITANRVLHCNADTPEEMHHWITLLQRSKGDTRVEGQEFIVRGW
12	1348	RPNSFVIITANRVLHCNADTPEEMHHWITLLQRSKGDTRVEGQEFIVRGW
humX	1349	RPNSFVIITANRVLHCNADTPEEMHHWITLLQRSKGDTRVEGQEFIVRGW
10	1398	LHKEVKNSPKMSSLKLKKRWVFLTHNSLDYYKSSEKNALKGLTLVLNSLC
12	1398	LHKEVKNSPKMSSLKLKKRWVFLTHNSLDYYKSSEKNALKGLTLVLNSLC
humX	1399	LHKEVKNSPKMSSLKLKKRWVFLTHNSLDYYKSSEKNALKGLTLVLNSLC
10	1448	SVVPPDEKIFKETGYWNVTVYGRKHCRYLYTKLLNEATRWSSVIQNVTD
12	1448	SVVPPDEKIFKETGYWNVTVYGRKHCRYLYTKLLNEATRWSSVIQNVTD
humX	1449	SVVPPDEKIFKETGYWNVTVYGRKHCRYLYTKLLNEATRWSSAIQNVTD
10	1498	KAPIDTPTQQLIQDIKENCLNSDVVEQIYKRNPILRYTHHPLHSPLLPLP
12	1498	KAPIDTPTQQLIQDIKENCLNSDVVEQIYKRNPILRYTHHPLHSPLLPLP
humX	1499	KAPIDTPTQQLIQDIKENCLNSDVVEQIYKRNPILRYTHHPLHSPLLPLP
10	1548	YGDINLNLKDKGYTTLQDEAIKIFNSLQQLESMSDPIPIIQGILQTGHD
12	1548	YGDINLNLKDKGYTTLQDEAIKIFNSLQQLESMSDPIPIIQGILQTGHD
humX	1549	YGDINLNLKDKGYTTLQDEAIKIFNSLQQLESMSDPIPIIQGILQTGHD
10	1598	LRPLRDELYCQLIKQTNKVPHPGSVGNLYSWQILTCLCTFLPSRGILKY
12	1598	LRPLRDELYCQLIKQTNKVPHPGSVGNLYSWQILTCLCTFLPSRGILKY
humX	1599	LRPLRDELYCQLIKQTNKVPHPGSVGNLYSWQILTCLCTFLPSRGILKY
10	1648	LKFHLKRIREQFPGTEMEKYALFTYESLKKTKREFVPSRDEIEALIHRO
12	1648	LKFHLKRIREQFPGTEMEKYALFTYESLKKTKREFVPSRDEIEALIHRO
humX	1649	LKFHLKRIREQFPGTEMEKYALFTYESLKKTKREFVPSRDEIEALIHRO
10	1698	EMTSTVYCHGGGSKITINSHTTAGEVVEKLIRGLAMEDSRNMFALFEYN
12	1698	EMTSTVYCHGGGSKITINSHTTAGEVVEKLIRGLAMEDSRNMFALFEYN
humX	1699	EMTSTVYCHGGGSKITINSHTTAGEVVEKLIRGLAMEDSRNMFALFEYN
10	1748	GHVDKAIESRTVVADVLAKFEKLAATSEVGDLPWKFYFKLYCFLDTDNVP
12	1748	GHVDKAIESRTVVADVLAKFEKLAATSEVGDLPWKFYFKLYCFLDTDNVP
humX	1749	GHVDKAIESRTVVADVLAKFEKLAATSEVGDLPWKFYFKLYCFLDTDNVP
10	1798	KDSVEFAFMFEQAHEAVIHGHHPAPEENLQVLAALRLQYLQGDYTLHAAI
12	1798	KDSVEFAFMFEQAHEAVIHGHHPAPEENLQVLAALRLQYLQGDYTLHAAI
humX	1799	KDSVEFAFMFEQAHEAVIHGHHPAPEENLQVLAALRLQYLQGDYTLHAAI
10	1848	PPLLEVYSLQRLKARISQSTKTFTPCERLEKRRTSFLEGTLLRRSFRTGSV

12	1848	PPLEEVYSLQRLKARISQSTKTFTPCERLEKRRTSFLEGLTLLRRSFRTGSV
humX	1849	PPLEEVYSLQRLKARISQSTKTFTPCERLEKRRTSFLEGLTLLRRSFRTGSV
10	1898	VRQKVEEEQMLDMWIKKEEVSSARASIIDKWRKFQGMNQEQAMAKYMALIK
12	1898	VRQKVEEEQMLDMWIKKEEVSSARASIIDKWRKFQGMNQEQAMAKYMALIK
humX	1899	VRQKVEEEQMLDMWIKKEEVSSARASIIDKWRKFQGMNQEQAMAKYMALIK
10	1948	EWPGYGSTLFDVECKEGGFPQELWLGVSADAVSVYKRGEGRPLEVFQYEH
12	1948	EWPGYGSTLFDVEVRTG-CHVLGWAGCWHLRTWITAKFMWREDKMEHFAL
humX	1949	EWPGYGSTLFDVECKEGGFPQELWLGVSADAVSVYKRGEGRPLEVFQYEH
10	1998	ILSFGAPLANTYKIVVDERELLFETSEVVDVAKLMKAYISMIVKKRYSTT
12	1997	STSFFRAPKIVPLTPPSSQFLFSCVVNASVILGMNAKLCHLFFYPPLG
humX	1999	ILSFGAPLANTYKIVVDERELLFETSEVVDVAKLMKAYISMIVKKRYSTT
10	2048	RSASSQGSSR
12	2047	KL-----
humX	2049	RSASSQGSSR

The invention also includes polypeptides and nucleotides having 80-100%, including 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98 and 99%, sequence identity to SEQ ID NOS:1-16, as well as nucleotides encoding any of these polypeptides, and compliments of any of these nucleotides. In an alternative embodiment, polypeptides and/or nucleotides (and compliments thereof) identical to any one of, or more than one of, SEQ ID NOS:1-16 are excluded. In yet another embodiment, polypeptides and/or nucleotides (and compliments thereof) having 81-100% identical, including 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98 and 99%, sequence identity to SEQ ID NOS:1-16 are excluded.

The nucleic acids and proteins of the invention are potentially useful in promoting wound healing, for example after organ transplantation, or in the treatment of myocardial infarction, but also in treating tumors, and in cancers, diabetic retinopathy, macular degeneration, psoriasis, and rheumatoid arthritis. For example, a cDNA encoding AAP may be useful in gene therapy, and AAP proteins may be useful when administered to a subject in need thereof. The novel nucleic acid encoding AAP, and the AAP proteins of the invention, or fragments thereof, may further be useful in diagnostic applications, wherein the presence or amount of the nucleic acid or the protein are to be assessed. These materials are further useful in the generation of Abs that bind immunospecifically to the novel substances of the invention for use in therapeutic or diagnostic methods.

Kelch-like protein (KLP)

The putative protein encoded by *KLP* contains 1 putative BTB domain and 4 putative Kelch motifs. The BTB (broad complex, tramtrack, bric-a-brac)/POZ (poxvirus, zinc finger) domain is involved in protein protein interactions. The kelch motif is sixfold tandem element in the sequence of the *Drosophila* kelch ORF1 protein that also contains BTB. Kelch ORF1 localizes to the ring canals in the egg chamber and helps to organize the F-actin cytoskeleton (Adams *et al.*, 2000). The repeated kelch motifs predict a conserved tertiary structure, a β -propeller. This module appears in many different polypeptide contexts and contains multiple potential protein-protein interaction sites. Members of this growing superfamily are present throughout the cell and extracellularly and have diverse activities (Adams *et al.*, 2000). Such activities include cytoskeleton organization, as well as other morphological processes, gene expression, interactions with viruses, and various extracellular events, such as cell spreading.

Alignment with *Drosophila* kelch and other kelch-like proteins, human kelch-like protein (GenBank AAF20938 (SEQ ID NO:17)), hypothetical *C. elegans* (GenBank O61795 (SEQ ID NO:18) and the skeletal muscle-specific sarcosin (GenBank O60662 (SEQ ID NO:19); (Taylor *et al.*, 1998)) reveals that the disclosed protein (SEQ ID NO:2) is a member of a new subfamily

KLP is associated with tube formation and angiogenesis because it is upregulated in the *in vitro* model of angiogenesis of Example 1. Kelch mediates cytoskeletal associations, it is involved in morphogenetic processes, such as tube formation, that depend on cytoskeletal arrangements and signaling. *KLP* represents an attractive target for small molecule drug therapy.

Human ortholog of mouse BAZF (hBAZF)

hBAZF (SEQ ID NO:4) is the human ortholog of mouse BAZF (GenBank AB011665; SEQ ID NO:20), BAZF is a Bcl-6 (LAZ3) homolog, a transcription repressor that controls germinal center formation and the T cell-dependent immune response. Expression of Bcl-6 negatively correlates with cellular proliferation: Bcl-6 suppresses growth associated with impaired mitotic S phase progression and apoptosis (Albagli *et al.*, 1999).

BAZF contains a BTB/POZ domain and five repeats of the Kruppel-like zinc finger motifs, instead of 6 in Bcl-6 (Okabe *et al.*, 1998). Expression of *BAZF* mRNA is

relegated to heart and lung, unlike *Bcl-6* mRNA, but is induced in activated lymphocytes as an immediate-early gene, like *Bcl-6* (Okabe *et al.*, 1998).

The hBAZF sequence was derived by using tblastn (protein query –translated database) (Altschul *et al.*, 1997), with the mouse protein sequence (GenBank O88282; SEQ ID NO:21) that has homology to GenBank AC015918 (SEQ ID NO:22), a clone of *Homo sapiens* chromosome 17. Human BAZF contains five Kruppel-like zinc finger motif repeats and a BTB/POZ domain.

The peptide sequence, “RSQ...PQV” that is present in the human sequence, might represent an alternative spliced form of the gene. Alignment with mouse BAZF, and alignment with mouse and human Bcl-6 demonstrates that the four proteins are almost identical in this region, but only human BAZF has this inserted sequence.

hBAZF is upregulated in HUVE cells grown embedded in collagen gels but not as a monolayer grown on collagen. When HUVE cells are suspended in collagen, they do not proliferate. Analogous to the role of mBAZF plays a role in regulating cell proliferation (Okabe *et al.*, 1998), hBAZF plays a role in cell proliferation in HUVE suspended in collagen. Because of its high expression during vessel morphogenesis, hBAZF represents an excellent molecular marker, as well as an attractive target for various therapies to inhibit angiogenesis.

hmt-Elongation Factor G (hEF-G)

The original isolation of hEF-G (SEQ ID NO:6) is 84% identical and colinear with *Rattus norvegicus* nuclear encoded mitochondrial elongation factor G (GenBank L14684 (SEQ ID NO:23); (Barker *et al.*, 1993). No human gene is described in GenBank. However, searching EST databases, the human gene is contained inside GenBank AC010936 (SEQ ID NO:24), a chromosome 3 clone. Alignment of hEF-G with rat mtEF-G and yeast EF-G1 demonstrates that the novel sequence is the ortholog of rat nuclear-encoded mitochondrial elongation factor G.

Bacterial elongation factor G (EF-G) physically associates with translocation-competent ribosomes and facilitates transition to the subsequent codon through the coordinate binding and hydrolysis of GTP. The deduced amino acid sequence of hmt-EF-G reveals characteristic motifs shared by all GTP binding proteins. Therefore, similarly

to other elongation factors, the enzymatic function of hmt-EF-G is predicted to depend on GTP binding and hydrolysis.

Hmt-EF-G is strongly induced (30-fold) in an *in vitro* model of angiogenesis (Example 1), and as such, hmt-EF-G represents an excellent molecular marker for vessel formation. Because of its putative localization to the mitochondrion, hmt-EF-G is also an attractive therapeutic target to treat disease states associated with mitochondrial dysfunction.

Human thyroid regulated transcript (hTRG)

hTRG (SEQ ID NO:8) is the human ortholog of rat TRG, a novel thyroid transcript negatively regulated by TSH (GenBank KIAA1058 (SEQ ID NO:25); (Bonapace *et al.*, 1990).

SEQ ID NO:25 appears to be a partial peptide since there are *C. elegans* homologous proteins of 2000 residues. Using tblastn (Altschul *et al.*, 1997) against genomic sequences, the hTRG sequence (SEQ ID NO:8) was assembled.

In *C. elegans*, homologous proteins localize either to the plasma membrane or to the mitochondrial inner membrane. A partial sequence, KIAA0694 (SEQ ID NO:26) also localizes to the mitochondrial matrix. hTRG has a PH domain, and has weak homology to an extracellular fibronectin-binding protein precursor. SEQ ID NO:26 has homology to *Drosophila* DOS and mouse Gab-2 proteins; both of which are involved in signal transduction, acting as adapter proteins between receptors and kinases like Ras1 (Hibi and Hirano, 2000).

Because of hTRG is upregulated during the *in vitro* model of angiogenesis (Example 1), and because of its homologies with adapter proteins, hTRG is likely to be involved in signal transduction between receptors and kinases. As such, hTRG represent an excellent candidate for small molecule drug therapy to modulate angiogenesis and treat angiogenesis-related diseases. In addition, because of its putative ability to respond to thyroid stimulating hormone (TSH), modulation of hTRG is useful to treat diseases related to TSH imbalance.

Human myosin X (hMX1(SEQ ID NO:10) and hMX2 (SEQ ID NO:12)

The hMX proteins represent the human ortholog of bovine myosin X, (GenBank AAB39486; SEQ ID NO:27). Using tblastn (Altschul *et al.*, 1997) and the bovine sequence, a series of genomic clones from human chromosome 5 were identified; GenBank AC010310 (SEQ ID NO:28) appears to contain the entire sequence.

Interestingly, a partial cDNA sequence from mouse (GenBank AF184153; SEQ ID NO:29) localizes to a 0.8 cM interval on the short arm of chromosome 5, between the polymorphic microsatellite markers D5S416 and D5S2114. In this region lies the gene for familial chondrocalcinosis (*CCAL2*) (Rojas *et al.*, 1999).

Another GenBank entry, AB018342 (SEQ ID NO:30) that represents the 3' region of *hMX*, appears to encode an alternative splice form. Noteworthy, this variant (*hMX2*) has a very hydrophobic carboxy terminus, while the more prevalent form (*hMX1*) is hydrophilic and potentially interacts with DNA/RNA since it has homology to high mobility group box (HMG) and ribosomal proteins. Additionally, a myosin head domain was found in the NH terminus, as well as a myosin talin domain, two calmodulin binding domains, four pleckstrin domains and a band 4.1 domain.

The band 4.1 domain represents a crossroads between cytoskeletal organization and signal transduction. The domain was first described in the red blood cell protein band 4.1. The ERM proteins ezrin, radixin, and moesin and the unconventional myosins VIIa and X all possess the band 4.1 domain (Louvet-Vallee, 2000). The band 4.1 domain binds single transmembrane protein at the membrane-proximal region in the C-terminal cytoplasmic tail.

HMX is upregulated during angiogenesis in an *in vitro* model (Example 1). Because *hMX* contains the protein-protein interaction domains PH and band 4.1 domain, *hMX1* and *hMX2* are involved in angiogenesis, likely transducing signals from angiogenic factors, perhaps modulating the cytoskeleton.

Human mitochondrial protein (hMP)

Analysis of *hMP* (SEQ ID NO:14) reveals several subdomain that are homologous to proteins involved in transport across membranes, K^+ ATPase α and γ chains. Further analysis indicates that *hMP* may bind DNA and or RNA, since *hMP* is homologous to histones and transcription factors, especially those possessing basic region plus leucine zipper domains.

Although PSORT analysis (Nakai and Horton, 1999) predicts nuclear localization (P=.6), hMP may in fact be a nuclear-encoded mitochondrial protein. Homologies with mostly bacterial proteins and a PSORT prediction of mitochondrial matrix space localization (P=0.4478) strongly support this contention.

Because hMP is upregulated in an *in vitro* model of angiogenesis (Example 1), and because of its homologies with mitochondrial and nuclear-localized polypeptides, hMP is important in vascular morphogenesis, most likely through either powering the cellular differentiation-redifferentiation process, and/or affecting changes in the nuclear matrix that change global gene expression. Alternatively, hMP may be a transcription factor for either the nuclear or mitochondrial genomes.

Nuclear hormone receptor (NHR)

NHR (SEQ ID NO:16) has two domains: (1) the NH region is similar to Swi3 (yeast SWI/SNF complexes regulate transcription by chromatin remodeling), indicating a role in transcriptional regulation, and (2) the COOH region is similar to parathyroid hormone-related proteins that bind parathyroid hormones. PSORT (Nakai and Horton, 1999) predicts the protein to localize in the nucleus P=0.9600.

The identification of this new putative hormone receptor-transcriptional regulator and hBAZF suggest a novel human transcriptional pathway that resembles, to some extent, that of Bcl-6.

Bcl-6 suppresses transcription via the BTB domain, which recruits a complex containing SMRT, retinoid thyroid hormone receptor, nuclear receptor corepressor (N-CoR), mammalian Sin3A, and histone deacetylase (HDAC). hBAZF, which also possesses a BTB domain, might recruit a similar complex containing deacetylase.

Expression data indicate that *hBAZF* is up-regulated while *NHR* is down-regulated. These data agree with other evidence related to tube formation. Testosterone (a steroid) and dexamethasone (a steroid-like molecule) strongly inhibit vessel formation, and all-trans retinoic acid (at-RA) and 9-cis retinoic acid (9-cis RA) stimulate capillary-like tubular structures (Lansink *et al.*, 1998).

Upon angiogenic stimulation, endothelial cells may become incompetent to respond to anti-angiogenic responses mediated by hormones using a dual mechanism, sequestering hormones and suppressing transcription. Because nHR is down-regulated

during *in vitro* angiogenesis (Example 1), this polypeptide is likely to be involved in non-angiogenesis-specific gene transcription. nHR is an attractive therapeutic target, especially in therapies that are directed at preventing vascularization.

AAP polynucleotides

One aspect of the invention pertains to isolated nucleic acid molecules that encode AAP or biologically-active portions thereof. Also included in the invention are nucleic acid fragments sufficient for use as hybridization probes to identify AAP-encoding nucleic acids (*e.g.*, AAP mRNAs) and fragments for use as polymerase chain reaction (PCR) primers for the amplification and/or mutation of AAP molecules. A “nucleic acid molecule” includes DNA molecules (*e.g.*, cDNA or genomic DNA), RNA molecules (*e.g.*, mRNA), analogs of the DNA or RNA generated using nucleotide analogs, and derivatives, fragments and homologs. The nucleic acid molecule may be single-stranded or double-stranded, but preferably comprises double-stranded DNA.

1. probes

Probes are nucleic acid sequences of variable length, preferably between at least about 10 nucleotides (nt), 100 nt, or many (*e.g.*, 6,000 nt) depending on the specific use. Probes are used to detect identical, similar, or complementary nucleic acid sequences. Longer length probes can be obtained from a natural or recombinant source, are highly specific, and much slower to hybridize than shorter-length oligomer probes. Probes may be single- or double-stranded and designed to have specificity in PCR, membrane-based hybridization technologies, or ELISA-like technologies. Probes are substantially purified oligonucleotides that will hybridize under stringent conditions to at least optimally 12, 25, 50, 100, 150, 200, 250, 300, 350 or 400 consecutive sense strand nucleotide sequence of SEQ ID NOS:1, 3, 5, 7, 9, 11, 13, or 15; or an anti-sense strand nucleotide sequence of these sequences; or of a naturally occurring mutant of these sequences.

The full- or partial length native sequence AAP may be used to “pull out” similar (homologous) sequences (Ausubel *et al.*, 1987; Sambrook, 1989), such as: (1) full-length or fragments of AAP cDNA from a cDNA library from any species (*e.g.* human, murine, feline, canine, bacterial, viral, retroviral, yeast), (2) from cells or tissues, (3) variants within a species, and (4) homologues and variants from other species. To find related sequences that may encode related genes, the probe may be designed to encode unique

sequences or degenerate sequences. Sequences may also be genomic sequences including promoters, enhancer elements and introns of native sequence *AAP*.

For example, an *AAP* coding region in another species may be isolated using such probes. A probe of about 40 bases is designed, based on an *AAP*, and made. To detect hybridizations, probes are labeled using, for example, radionuclides such as ^{32}P or ^{35}S , or enzymatic labels such as alkaline phosphatase coupled to the probe via avidin-biotin systems. Labeled probes are used to detect nucleic acids having a complementary sequence to that of an *AAP* in libraries of cDNA, genomic DNA or mRNA of a desired species.

Such probes can be used as a part of a diagnostic test kit for identifying cells or tissues which mis-express an *AAP*, such as by measuring a level of an *AAP* in a sample of cells from a subject *e.g.*, detecting *AAP* mRNA levels or determining whether a genomic *AAP* has been mutated or deleted.

2. *isolated nucleic acid*

An isolated nucleic acid molecule is separated from other nucleic acid molecules that are present in the natural source of the nucleic acid. Preferably, an isolated nucleic acid is free of sequences that naturally flank the nucleic acid (*i.e.*, sequences located at the 5'- and 3'-termini of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, isolated *AAP* molecules can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell/tissue from which the nucleic acid is derived (*e.g.*, brain, heart, liver, spleen, *etc.*). Moreover, an isolated nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material or culture medium when produced by recombinant techniques, or of chemical precursors or other chemicals when chemically synthesized.

A nucleic acid molecule of the invention, *e.g.*, a nucleic acid molecule having the nucleotide sequence of SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15, or a complement of this aforementioned nucleotide sequence, can be isolated using standard molecular biology techniques and the provided sequence information. Using all or a portion of the nucleic acid sequence of SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15 as a hybridization probe, *AAP*

molecules can be isolated using standard hybridization and cloning techniques (Ausubel *et al.*, 1987; Sambrook, 1989).

PCR amplification techniques can be used to amplify *AAP* using cDNA, mRNA or alternatively, genomic DNA, as a template and appropriate oligonucleotide primers. Such nucleic acids can be cloned into an appropriate vector and characterized by DNA sequence analysis. Furthermore, oligonucleotides corresponding to *AAP* sequences can be prepared by standard synthetic techniques, *e.g.*, an automated DNA synthesizer.

3. *oligonucleotide*

An oligonucleotide comprises a series of linked nucleotide residues, which oligonucleotide has a sufficient number of nucleotide bases to be used in a PCR reaction or other application. A short oligonucleotide sequence may be based on, or designed from, a genomic or cDNA sequence and is used to amplify, confirm, or reveal the presence of an identical, similar or complementary DNA or RNA in a particular cell or tissue. Oligonucleotides comprise portions of a nucleic acid sequence having about 10 nt, 50 nt, or 100 nt in length, preferably about 15 nt to 30 nt in length. In one embodiment of the invention, an oligonucleotide comprising a nucleic acid molecule less than 100 nt in length would further comprise at least 6 contiguous nucleotides of SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15, or a complement thereof. Oligonucleotides may be chemically synthesized and may also be used as probes.

4. *complementary nucleic acid sequences; binding*

In another embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule that is a complement of the nucleotide sequence shown in SEQ ID NOS: 1, 3, 5, 7, 9, 11, 13 or 15, or a portion of this nucleotide sequence (*e.g.*, a fragment that can be used as a probe or primer or a fragment encoding a biologically-active portion of an *AAP*). A nucleic acid molecule that is complementary to the nucleotide sequence shown in SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15, is one that is sufficiently complementary to the nucleotide sequence shown in SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15, that it can hydrogen bond with little or no mismatches to the nucleotide sequence shown in SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15, thereby forming a stable duplex.

“Complementary” refers to Watson-Crick or Hoogsteen base pairing between nucleotides units of a nucleic acid molecule, and the term “binding” means the physical

or chemical interaction between two polypeptides or compounds or associated polypeptides or compounds or combinations thereof. Binding includes ionic, non-ionic, van der Waals, hydrophobic interactions, and the like. A physical interaction can be either direct or indirect. Indirect interactions may be through or due to the effects of another polypeptide or compound. Direct binding refers to interactions that do not take place through, or due to, the effect of another polypeptide or compound, but instead are without other substantial chemical intermediates.

Nucleic acid fragments are at least 6 (contiguous) nucleic acids or at least 4 (contiguous) amino acids, a length sufficient to allow for specific hybridization in the case of nucleic acids or for specific recognition of an epitope in the case of amino acids, respectively, and are at most some portion less than a full-length sequence. Fragments may be derived from any contiguous portion of a nucleic acid or amino acid sequence of choice.

5. *derivatives, and analogs*

Derivatives are nucleic acid sequences or amino acid sequences formed from the native compounds either directly or by modification or partial substitution. Analogs are nucleic acid sequences or amino acid sequences that have a structure similar to, but not identical to, the native compound but differ from it in respect to certain components or side chains. Analogs may be synthetic or from a different evolutionary origin and may have a similar or opposite metabolic activity compared to wild type. Homologs are nucleic acid sequences or amino acid sequences of a particular gene that are derived from different species.

Derivatives and analogs may be full length or other than full length, if the derivative or analog contains a modified nucleic acid or amino acid, as described below. Derivatives or analogs of the nucleic acids or proteins of the invention include, but are not limited to, molecules comprising regions that are substantially homologous to the nucleic acids or proteins of the invention, in various embodiments, by at least about 70%, 80%, or 95% identity (with a preferred identity of 80-95%) over a nucleic acid or amino acid sequence of identical size or when compared to an aligned sequence in which the alignment is done by a computer homology program known in the art, or whose encoding nucleic acid is capable of hybridizing to the complement of a sequence encoding the

aforementioned proteins under stringent, moderately stringent, or low stringent conditions (Ausubel *et al.*, 1987).

6. *homology*

A “homologous nucleic acid sequence” or “homologous amino acid sequence,” or variations thereof, refer to sequences characterized by homology at the nucleotide level or amino acid level as discussed above. Homologous nucleotide sequences encode those sequences coding for isoforms of AAP. Isoforms can be expressed in different tissues of the same organism as a result of, for example, alternative splicing of RNA. Alternatively, different genes can encode isoforms. In the invention, homologous nucleotide sequences include nucleotide sequences encoding for an AAP of species other than humans, including, but not limited to: vertebrates, and thus can include, *e.g.*, frog, mouse, rat, rabbit, dog, cat, cow, horse, and other organisms. Homologous nucleotide sequences also include, but are not limited to, naturally occurring allelic variations and mutations of the nucleotide sequences set forth herein. A homologous nucleotide sequence does not, however, include the exact nucleotide sequence encoding human AAP. Homologous nucleic acid sequences include those nucleic acid sequences that encode conservative amino acid substitutions (see below) in SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16, as well as a polypeptide possessing AAP biological activity. Various biological activities of the AAP are described below.

7. *open reading frames*

The open reading frame (ORF) of an AAP gene encodes an AAP. An ORF is a nucleotide sequence that has a start codon (ATG) and terminates with one of the three “stop” codons (TAA, TAG, or TGA). In this invention, however, an ORF may be any part of a coding sequence that may or may not comprise a start codon and a stop codon. To achieve a unique sequence, preferable AAP ORFs encode at least 50 amino acids.

AAP polypeptides

1. *mature*

An AAP can encode a mature AAP. A “mature” form of a polypeptide or protein disclosed in the present invention is the product of a naturally occurring polypeptide or precursor form or proprotein. The naturally occurring polypeptide, precursor or proprotein includes, by way of nonlimiting example, the full-length gene product,

encoded by the corresponding gene. Alternatively, it may be defined as the polypeptide, precursor or proprotein encoded by an open reading frame described herein. The product “mature” form arises, again by way of nonlimiting example, as a result of one or more naturally occurring processing steps as they may take place within the cell, or host cell, in which the gene product arises. Examples of such processing steps leading to a “mature” form of a polypeptide or protein include the cleavage of the N-terminal methionine residue encoded by the initiation codon of an open reading frame, or the proteolytic cleavage of a signal peptide or leader sequence. Thus a mature form arising from a precursor polypeptide or protein that has residues 1 to N, where residue 1 is the N-terminal methionine, would have residues 2 through N remaining after removal of the N-terminal methionine. Alternatively, a mature form arising from a precursor polypeptide or protein having residues 1 to N, in which an N-terminal signal sequence from residue 1 to residue M is cleaved, would have the residues from residue M+1 to residue N remaining. Further as used herein, a “mature” form of a polypeptide or protein may arise from a step of post-translational modification other than a proteolytic cleavage event. Such additional processes include, by way of non-limiting example, glycosylation, myristoylation or phosphorylation. In general, a mature polypeptide or protein may result from the operation of only one of these processes, or a combination of any of them.

2. *active*

An active AAP polypeptide or AAP polypeptide fragment retains a biological and/or an immunological activity similar, but not necessarily identical, to an activity of a naturally-occurring (wild-type) AAP polypeptide of the invention, including mature forms. A particular biological assay, with or without dose dependency, can be used to determine AAP activity. A nucleic acid fragment encoding a biologically-active portion of AAP can be prepared by isolating a portion of SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15 that encodes a polypeptide having an AAP biological activity (the biological activities of the AAP are described below), expressing the encoded portion of AAP (*e.g.*, by recombinant expression *in vitro*) and assessing the activity of the encoded portion of AAP. Immunological activity refers to the ability to induce the production of an antibody against an antigenic epitope possessed by a native AAP; biological activity refers to a function, either inhibitory or stimulatory, caused by a native AAP that excludes immunological activity.

AAP nucleic acid variants and hybridization

1. *variant polynucleotides, genes and recombinant genes* The invention further encompasses nucleic acid molecules that differ from the nucleotide sequences shown in SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15 due to degeneracy of the genetic code and thus encode the same AAP as that encoded by the nucleotide sequences shown in SEQ ID NO NOS:1, 3, 5, 7, 9, 11, 13 or 15. An isolated nucleic acid molecule of the invention has a nucleotide sequence encoding a protein having an amino acid sequence shown in SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16.

In addition to the AAP sequences shown in SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15, DNA sequence polymorphisms that change the amino acid sequences of the AAP may exist within a population. For example, allelic variation among individuals will exhibit genetic polymorphism in an AAP. The terms "gene" and "recombinant gene" refer to nucleic acid molecules comprising an open reading frame (ORF) encoding an AAP, preferably a vertebrate AAP. Such natural allelic variations can typically result in 1-5% variance in an AAP. Any and all such nucleotide variations and resulting amino acid polymorphisms in an AAP, which are the result of natural allelic variation and that do not alter the functional activity of an AAP are within the scope of the invention.

Moreover, AAP from other species that have a nucleotide sequence that differs from the human sequence of SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15, are contemplated. Nucleic acid molecules corresponding to natural allelic variants and homologues of an AAP cDNAs of the invention can be isolated based on their homology to an AAP of SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15 using cDNA-derived probes to hybridize to homologous AAP sequences under stringent conditions.

"AAP variant polynucleotide" or "AAP variant nucleic acid sequence" means a nucleic acid molecule which encodes an active AAP that (1) has at least about 80% nucleic acid sequence identity with a nucleotide acid sequence encoding a full-length native AAP, (2) a full-length native AAP lacking the signal peptide, (3) an extracellular domain of an AAP, with or without the signal peptide, or (4) any other fragment of a full-length AAP. Ordinarily, an AAP variant polynucleotide will have at least about 80% nucleic acid sequence identity, more preferably at least about 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% nucleic

acid sequence identity and yet more preferably at least about 99% nucleic acid sequence identity with the nucleic acid sequence encoding a full-length native AAP. An AAP variant polynucleotide may encode a full-length native AAP lacking the signal peptide, an extracellular domain of an AAP, with or without the signal sequence, or any other fragment of a full-length AAP. Variants do not encompass the native nucleotide sequence.

Ordinarily, AAP variant polynucleotides are at least about 30 nucleotides in length, often at least about 60, 90, 120, 150, 180, 210, 240, 270, 300, 450, 600 nucleotides in length, more often at least about 900 nucleotides in length, or more.

“Percent (%) nucleic acid sequence identity” with respect to AAP-encoding nucleic acid sequences identified herein is defined as the percentage of nucleotides in a candidate sequence that are identical with the nucleotides in the AAP sequence of interest, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity. Alignment for purposes of determining % nucleic acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN or Megalign (DNASTAR) software. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full length of the sequences being compared.

When nucleotide sequences are aligned, the % nucleic acid sequence identity of a given nucleic acid sequence C to, with, or against a given nucleic acid sequence D (which can alternatively be phrased as a given nucleic acid sequence C that has or comprises a certain % nucleic acid sequence identity to, with, or against a given nucleic acid sequence D) can be calculated as follows:

$$\% \text{nucleic acid sequence identity} = W/Z \cdot 100$$

where

W is the number of nucleotides cored as identical matches by the sequence alignment program's or algorithm's alignment of C and D

and

Z is the total number of nucleotides in D.

When the length of nucleic acid sequence C is not equal to the length of nucleic acid sequence D, the % nucleic acid sequence identity of C to D will not equal the % nucleic acid sequence identity of D to C.

2. Stringency

Homologs (*i.e.*, nucleic acids encoding an AAP derived from species other than human) or other related sequences (*e.g.*, paralogs) can be obtained by low, moderate or high stringency hybridization with all or a portion of the particular human sequence as a probe using methods well known in the art for nucleic acid hybridization and cloning.

The specificity of single stranded DNA to hybridize complementary fragments is determined by the “stringency” of the reaction conditions. Hybridization stringency increases as the propensity to form DNA duplexes decreases. In nucleic acid hybridization reactions, the stringency can be chosen to either favor specific hybridizations (high stringency), which can be used to identify, for example, full-length clones from a library. Less-specific hybridizations (low stringency) can be used to identify related, but not exact, DNA molecules (homologous, but not identical) or segments.

DNA duplexes are stabilized by: (1) the number of complementary base pairs, (2) the type of base pairs, (3) salt concentration (ionic strength) of the reaction mixture, (4) the temperature of the reaction, and (5) the presence of certain organic solvents, such as formamide which decreases DNA duplex stability. In general, the longer the probe, the higher the temperature required for proper annealing. A common approach is to vary the temperature: higher relative temperatures result in more stringent reaction conditions. (Ausubel *et al.*, 1987) provide an excellent explanation of stringency of hybridization reactions.

To hybridize under “stringent conditions” describes hybridization protocols in which nucleotide sequences at least 60% homologous to each other remain hybridized. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength, pH and nucleic acid concentration) at which 50% of the probes complementary to the target sequence hybridize to the target sequence at equilibrium. Since the target sequences are generally present at excess, at T_m , 50% of the probes are occupied at equilibrium.

(a) *high stringency*

“Stringent hybridization conditions” conditions enable a probe, primer or oligonucleotide to hybridize only to its target sequence. Stringent conditions are sequence-dependent and will differ. Stringent conditions comprise: (1) low ionic strength and high temperature washes (*e.g.* 15 mM sodium chloride, 1.5 mM sodium citrate, 0.1 % sodium dodecyl sulfate at 50°C); (2) a denaturing agent during hybridization (*e.g.* 50% (v/v) formamide, 0.1% bovine serum albumin, 0.1% Ficoll, 0.1% polyvinylpyrrolidone, 50mM sodium phosphate buffer (pH 6.5; 750 mM sodium chloride, 75 mM sodium citrate at 42°C); or (3) 50% formamide. Washes typically also comprise 5X SSC (0.75 M NaCl, 75 mM sodium citrate), 50 mM sodium phosphate (pH 6.8), 0.1% sodium pyrophosphate, 5 x Denhardt’s solution, sonicated salmon sperm DNA (50 µg/ml), 0.1% SDS, and 10% dextran sulfate at 42°C, with washes at 42°C in 0.2 x SSC (sodium chloride/sodium citrate) and 50% formamide at 55°C, followed by a high-stringency wash consisting of 0.1 x SSC containing EDTA at 55°C. Preferably, the conditions are such that sequences at least about 65%, 70%, 75%, 85%, 90%, 95%, 98%, or 99% homologous to each other typically remain hybridized to each other. These conditions are presented as examples and are not meant to be limiting.

(b) *moderate stringency*

“Moderately stringent conditions” use washing solutions and hybridization conditions that are less stringent (Sambrook, 1989), such that a polynucleotide will hybridize to the entire, fragments, derivatives or analogs of SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15. One example comprises hybridization in 6X SSC, 5X Denhardt’s solution, 0.5% SDS and 100 mg/ml denatured salmon sperm DNA at 55°C, followed by one or more washes in 1X SSC, 0.1% SDS at 37°C. The temperature, ionic strength, *etc.*, can be adjusted to accommodate experimental factors such as probe length. Other moderate stringency conditions are described in (Ausubel *et al.*, 1987; Kriegler, 1990).

(c) *low stringency*

“Low stringent conditions” use washing solutions and hybridization conditions that are less stringent than those for moderate stringency (Sambrook, 1989), such that a polynucleotide will hybridize to the entire, fragments, derivatives or analogs of SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15. A non-limiting example of low stringency hybridization conditions are hybridization in 35% formamide, 5X SSC, 50 mM Tris-HCl (pH 7.5), 5

mM EDTA, 0.02% PVP, 0.02% Ficoll, 0.2% BSA, 100 mg/ml denatured salmon sperm DNA, 10% (wt/vol) dextran sulfate at 40°C, followed by one or more washes in 2X SSC, 25 mM Tris-HCl (pH 7.4), 5 mM EDTA, and 0.1% SDS at 50°C. Other conditions of low stringency, such as those for cross-species hybridizations are described in (Ausubel *et al.*, 1987; Kriegler, 1990; Shilo and Weinberg, 1981).

3. *Conservative mutations*

In addition to naturally-occurring allelic variants of AAP, changes can be introduced by mutation into SEQ ID NO NOS:1, 3, 5, 7, 9, 11, 13 or 15 sequences that incur alterations in the amino acid sequences of the encoded AAP that do not alter the AAP function. For example, nucleotide substitutions leading to amino acid substitutions at "non-essential" amino acid residues can be made in the sequence of SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequences of the AAP without altering their biological activity, whereas an "essential" amino acid residue is required for such biological activity. For example, amino acid residues that are conserved among the AAP of the invention are predicted to be particularly non-amenable to alteration. Amino acids for which conservative substitutions can be made are well-known in the art.

Useful conservative substitutions are shown in Table A, "Preferred substitutions." Conservative substitutions whereby an amino acid of one class is replaced with another amino acid of the same type fall within the scope of the subject invention so long as the substitution does not materially alter the biological activity of the compound. If such substitutions result in a change in biological activity, then more substantial changes, indicated in Table B as exemplary are introduced and the products screened for an AAP polypeptide's biological activity.

Table A Preferred substitutions

Original residue	Exemplary substitutions	Preferred substitutions
Ala (A)	Val, Leu, Ile	Val
Arg (R)	Lys, Gln, Asn	Lys
Asn (N)	Gln, His, Lys, Arg	Gln
Asp (D)	Glu	Glu
Cys (C)	Ser	Ser
Gln (Q)	Asn	Asn
Glu (E)	Asp	Asp

Gly (G)	Pro, Ala	Ala
His (H)	Asn, Gln, Lys, Arg	Arg
Ile (I)	Leu, Val, Met, Ala, Phe, Norleucine	Leu
Leu (L)	Norleucine, Ile, Val, Met, Ala, Phe	Ile
Lys (K)	Arg, Gln, Asn	Arg
Met (M)	Leu, Phe, Ile	Leu
Phe (F)	Leu, Val, Ile, Ala, Tyr	Leu
Pro (P)	Ala	Ala
Ser (S)	Thr	Thr
Thr (T)	Ser	Ser
Trp (W)	Tyr, Phe	Tyr
Tyr (Y)	Trp, Phe, Thr, Ser	Phe
Val (V)	Ile, Leu, Met, Phe, Ala, Norleucine	Leu

Non-conservative substitutions that effect (1) the structure of the polypeptide backbone, such as a β -sheet or α -helical conformation, (2) the charge or (3) hydrophobicity, or (4) the bulk of the side chain of the target site can modify an AAP polypeptide's function or immunological identity. Residues are divided into groups based on common side-chain properties as denoted in Table B. Non-conservative substitutions entail exchanging a member of one of these classes for another class. Substitutions may be introduced into conservative substitution sites or more preferably into non-conserved sites.

Table B Amino acid classes

Class	Amino acids
hydrophobic	Norleucine, Met, Ala, Val, Leu, Ile
neutral hydrophilic	Cys, Ser, Thr
acidic	Asp, Glu
basic	Asn, Gln, His, Lys, Arg
disrupt chain conformation	Gly, Pro
aromatic	Trp, Tyr, Phe

The variant polypeptides can be made using methods known in the art such as oligonucleotide-mediated (site-directed) mutagenesis, alanine scanning, and PCR mutagenesis. Site-directed mutagenesis (Carter, 1986; Zoller and Smith, 1987), cassette mutagenesis, restriction selection mutagenesis (Wells *et al.*, 1985) or other known

techniques can be performed on the cloned DNA to produce the AAP variant DNA (Ausubel *et al.*, 1987; Sambrook, 1989).

In one embodiment, the isolated nucleic acid molecule comprises a nucleotide sequence encoding a protein, wherein the protein comprises an amino acid sequence at least about 45%, preferably 60%, more preferably 70%, 80%, 90%, and most preferably about 95% homologous to SEQ ID NOS:1, 3, 5, 7, 9, 11, 13, or 15.

A mutant AAP can be assayed for blocking angiogenesis *in vitro*.

4. *Anti-sense nucleic acids*

Using antisense and sense AAP oligonucleotides can prevent AAP polypeptide expression. These oligonucleotides bind to target nucleic acid sequences, forming duplexes that block transcription or translation of the target sequence by enhancing degradation of the duplexes, terminating prematurely transcription or translation, or by other means.

Antisense or sense oligonucleotides are single-stranded nucleic acids, either RNA or DNA, which can bind a target AAP mRNA (sense) or an AAP DNA (antisense) sequences. Anti-sense nucleic acids can be designed according to Watson and Crick or Hoogsteen base pairing rules. The anti-sense nucleic acid molecule can be complementary to the entire coding region of an AAP mRNA, but more preferably, to only a portion of the coding or noncoding region of an AAP mRNA. For example, the anti-sense oligonucleotide can be complementary to the region surrounding the translation start site of an AAP mRNA. Antisense or sense oligonucleotides may comprise a fragment of the AAP DNA coding region of at least about 14 nucleotides, preferably from about 14 to 30 nucleotides. In general, antisense RNA or DNA molecules can comprise at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100 bases in length or more. Among others, (Stein and Cohen, 1988; van der Krol *et al.*, 1988a) describe methods to derive antisense or a sense oligonucleotides from a given cDNA sequence.

Examples of modified nucleotides that can be used to generate the anti-sense nucleic acid include: 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-

5 methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the anti-sense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been sub-cloned in an anti-sense orientation such that the transcribed RNA will be complementary to a target nucleic acid of interest.

To introduce antisense or sense oligonucleotides into target cells (cells containing the target nucleic acid sequence), any gene transfer method may be used. Examples of gene transfer methods include (1) biological, such as gene transfer vectors like Epstein-Barr virus or conjugating the exogenous DNA to a ligand-binding molecule, (2) physical, such as electroporation and injection, and (3) chemical, such as CaPO_4 precipitation and oligonucleotide-lipid complexes.

An antisense or sense oligonucleotide is inserted into a suitable gene transfer retroviral vector. A cell containing the target nucleic acid sequence is contacted with the recombinant retroviral vector, either *in vivo* or *ex vivo*. Examples of suitable retroviral vectors include those derived from the murine retrovirus M-MuLV, N2 (a retrovirus derived from M-MuLV), or the double copy vectors designated DCT5A, DCT5B and DCT5C (WO 90/13641, 1990). To achieve sufficient nucleic acid molecule transcription, vector constructs in which the transcription of the anti-sense nucleic acid molecule is controlled by a strong pol II or pol III promoter are preferred.

To specify target cells in a mixed population of cells cell surface receptors that are specific to the target cells can be exploited. Antisense and sense oligonucleotides are conjugated to a ligand-binding molecule, as described in (WO 91/04753, 1991). Ligands are chosen for receptors that are specific to the target cells. Examples of suitable ligand-binding molecules include cell surface receptors, growth factors, cytokines, or other ligands that bind to cell surface receptors or molecules. Preferably, conjugation of the ligand-binding molecule does not substantially interfere with the ability of the receptors

or molecule to bind the ligand-binding molecule conjugate, or block entry of the sense or antisense oligonucleotide or its conjugated version into the cell.

Liposomes efficiently transfer sense or an antisense oligonucleotide to cells (WO 90/10448, 1990). The sense or antisense oligonucleotide-lipid complex is preferably dissociated within the cell by an endogenous lipase.

The anti-sense nucleic acid molecule of the invention may be an α -anomeric nucleic acid molecule. An α -anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual α -units, the strands run parallel to each other (Gautier *et al.*, 1987). The anti-sense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide (Inoue *et al.*, 1987a) or a chimeric RNA-DNA analogue (Inoue *et al.*, 1987b).

In one embodiment, an anti-sense nucleic acid of the invention is a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity that are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they have a complementary region. Thus, ribozymes, such as hammerhead ribozymes (Haseloff and Gerlach, 1988) can be used to catalytically cleave AAP mRNA transcripts and thus inhibit translation. A ribozyme specific for an AAP-encoding nucleic acid can be designed based on the nucleotide sequence of an AAP cDNA (*i.e.*, SEQ ID NOS:1, 3, 5, 7, 9, 11, 13 or 15). For example, a derivative of a *Tetrahymena* L-19 IVS RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in an AAP-encoding mRNA (Cech *et al.*, U.S. Patent No. 5,116,742, 1992; Cech *et al.*, U.S. Patent No. 4,987,071, 1991). An AAP mRNA can also be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules (Bartel and Szostak, 1993).

Alternatively, AAP expression can be inhibited by targeting nucleotide sequences complementary to the regulatory region of an AAP (*e.g.*, an AAP promoter and/or enhancers) to form triple helical structures that prevent transcription of an AAP in target cells (Helene, 1991; Helene *et al.*, 1992; Maher, 1992).

Modifications of antisense and sense oligonucleotides can augment their effectiveness. Modified sugar-phosphodiester bonds or other sugar linkages (WO 91/06629, 1991), increase *in vivo* stability by conferring resistance to endogenous nucleases without disrupting binding specificity to target sequences. Other modifications

can increase the affinities of the oligonucleotides for their targets, such as covalently linked organic moieties (WO 90/10448, 1990) or poly-(L)-lysine. Other attachments modify binding specificities of the oligonucleotides for their targets, including metal complexes or intercalating (*e.g.* ellipticine) and alkylating agents.

For example, the deoxyribose phosphate backbone of the nucleic acids can be modified to generate peptide nucleic acids (Hyrup and Nielsen, 1996). "Peptide nucleic acids" or "PNAs" refer to nucleic acid mimics (*e.g.*, DNA mimics) in that the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs allows for specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols (Hyrup and Nielsen, 1996; Perry-O'Keefe *et al.*, 1996).

PNAs of an AAP can be used in therapeutic and diagnostic applications. For example, PNAs can be used as anti-sense or antigene agents for sequence-specific modulation of gene expression by inducing transcription or translation arrest or inhibiting replication. AAP PNAs may also be used in the analysis of single base pair mutations (*e.g.*, PNA directed PCR clamping; as artificial restriction enzymes when used in combination with other enzymes, *e.g.*, S₁ nucleases (Hyrup and Nielsen, 1996); or as probes or primers for DNA sequence and hybridization (Hyrup and Nielsen, 1996; Perry-O'Keefe *et al.*, 1996).

PNAs of an AAP can be modified to enhance their stability or cellular uptake. Lipophilic or other helper groups may be attached to PNAs, PNA-DNA dimmers formed, or the use of liposomes or other drug delivery techniques. For example, PNA-DNA chimeras can be generated that may combine the advantageous properties of PNA and DNA. Such chimeras allow DNA recognition enzymes (*e.g.*, RNase H and DNA polymerases) to interact with the DNA portion while the PNA portion provides high binding affinity and specificity. PNA-DNA chimeras can be linked using linkers of appropriate lengths selected in terms of base stacking, number of bonds between the nucleobases, and orientation (Hyrup and Nielsen, 1996). The synthesis of PNA-DNA chimeras can be performed (Finn *et al.*, 1996; Hyrup and Nielsen, 1996). For example, a DNA chain can be synthesized on a solid support using standard phosphoramidite coupling chemistry, and modified nucleoside analogs, *e.g.*, 5'-(4-methoxytrityl)amino-5'-

deoxy-thymidine phosphoramidite, can be used between the PNA and the 5' end of DNA (Finn *et al.*, 1996; Hyrup and Nielsen, 1996). PNA monomers are then coupled in a stepwise manner to produce a chimeric molecule with a 5' PNA segment and a 3' DNA segment (Finn *et al.*, 1996). Alternatively, chimeric molecules can be synthesized with a 5' DNA segment and a 3' PNA segment (Petersen *et al.*, 1976).

The oligonucleotide may include other appended groups such as peptides (*e.g.*, for targeting host cell receptors *in vivo*), or agents facilitating transport across the cell membrane (Lemaitre *et al.*, 1987; Letsinger *et al.*, 1989) or PCT Publication No. WO88/09810) or the blood-brain barrier (*e.g.*, PCT Publication No. WO 89/10134). In addition, oligonucleotides can be modified with hybridization-triggered cleavage agents (van der Krol *et al.*, 1988b) or intercalating agents (Zon, 1988). The oligonucleotide may be conjugated to another molecule, *e.g.*, a peptide, a hybridization triggered cross-linking agent, a transport agent, a hybridization-triggered cleavage agent, and the like.

AAP polypeptides

One aspect of the invention pertains to isolated AAP, and biologically-active portions derivatives, fragments, analogs or homologs thereof. Also provided are polypeptide fragments suitable for use as immunogens to raise anti-AAP Abs. In one embodiment, a native AAP can be isolated from cells or tissue sources by an appropriate purification scheme using standard protein purification techniques. In another embodiment, AAP are produced by recombinant DNA techniques. Alternative to recombinant expression, an AAP can be synthesized chemically using standard peptide synthesis techniques.

1. Polypeptides

An AAP polypeptide includes the amino acid sequence of an AAP whose sequences are provided in SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16. The invention also includes a mutant or variant protein any of whose residues may be changed from the corresponding residues shown in SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16, while still encoding a protein that maintains its AAP activities and physiological functions, or a functional fragment thereof.

2. Variant AAP polypeptides

In general, an AAP variant that preserves an AAP-like function and includes any variant in which residues at a particular position in the sequence have been substituted by other amino acids, and further includes the possibility of inserting an additional residue or residues between two residues of the parent protein as well as the possibility of deleting one or more residues from the parent sequence. Any amino acid substitution, insertion, or deletion is encompassed by the invention. In favorable circumstances, the substitution is a conservative substitution as defined above.

“AAP polypeptide variant” means an active AAP polypeptide having at least: (1) about 80% amino acid sequence identity with a full-length native sequence AAP polypeptide sequence, (2) an AAP polypeptide sequence lacking the signal peptide, (3) an extracellular domain of an AAP polypeptide, with or without the signal peptide, or (4) any other fragment of a full-length AAP polypeptide sequence. For example, AAP polypeptide variants include AAP polypeptides wherein one or more amino acid residues are added or deleted at the N- or C- terminus of the full-length native amino acid sequence. An AAP polypeptide variant will have at least about 80% amino acid sequence identity, preferably at least about 81% amino acid sequence identity, more preferably at least about 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% amino acid sequence identity and most preferably at least about 99% amino acid sequence identity with a full-length native sequence AAP polypeptide sequence. An AAP polypeptide variant may have a sequence lacking the signal peptide, an extracellular domain of an AAP polypeptide, with or without the signal peptide, or any other fragment of a full-length AAP polypeptide sequence. Ordinarily, AAP variant polypeptides are at least about 10 amino acids in length, often at least about 20 amino acids in length, more often at least about 30, 40, 50, 60, 70, 80, 90, 100, 150, 200, or 300 amino acids in length, or more.

“Percent (%) amino acid sequence identity” is defined as the percentage of amino acid residues that are identical with amino acid residues in a disclosed AAP polypeptide sequence in a candidate sequence when the two sequences are aligned. To determine % amino acid identity, sequences are aligned and if necessary, gaps are introduced to achieve the maximum % sequence identity; conservative substitutions are not considered as part of the sequence identity. Amino acid sequence alignment procedures to determine percent identity are well known to those of skill in the art. Often publicly available

computer software such as BLAST, BLAST2, ALIGN2 or Megalign (DNASTAR) software is used to align peptide sequences. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full length of the sequences being compared.

When amino acid sequences are aligned, the % amino acid sequence identity of a given amino acid sequence A to, with, or against a given amino acid sequence B (which can alternatively be phrased as a given amino acid sequence A that has or comprises a certain % amino acid sequence identity to, with, or against a given amino acid sequence B) can be calculated as:

$$\% \text{amino acid sequence identity} = X/Y \cdot 100$$

where

X is the number of amino acid residues scored as identical matches by the sequence alignment program's or algorithm's alignment of A and B

and

Y is the total number of amino acid residues in B.

If the length of amino acid sequence A is not equal to the length of amino acid sequence B, the % amino acid sequence identity of A to B will not equal the % amino acid sequence identity of B to A.

3. *Isolated/purified polypeptides*

An "isolated" or "purified" polypeptide, protein or biologically active fragment is separated and/or recovered from a component of its natural environment. Contaminant components include materials that would typically interfere with diagnostic or therapeutic uses for the polypeptide, and may include enzymes, hormones, and other proteinaceous or non-proteinaceous materials. Preferably, the polypeptide is purified to a sufficient degree to obtain at least 15 residues of N-terminal or internal amino acid sequence. To be substantially isolated, preparations having less than 30% by dry weight of non-AAP contaminating material (contaminants), more preferably less than 20%, 10% and most preferably less than 5% contaminants. An isolated, recombinantly-produced AAP or biologically active portion is preferably substantially free of culture medium, *i.e.*, culture medium represents less than 20%, more preferably less than about 10%, and most

preferably less than about 5% of the volume of the AAP preparation. Examples of contaminants include cell debris, culture media, and substances used and produced during *in vitro* synthesis of an AAP.

4. *Biologically active*

Biologically active portions of an AAP include peptides comprising amino acid sequences sufficiently homologous to or derived from the amino acid sequences of an AAP (SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16) that include fewer amino acids than a full-length AAP, and exhibit at least one activity of an AAP. Biologically active portions comprise a domain or motif with at least one activity of a native AAP. A biologically active portion of an AAP can be a polypeptide that is, for example, 10, 25, 50, 100 or more amino acid residues in length. Other biologically active portions, in which other regions of the protein are deleted, can be prepared by recombinant techniques and evaluated for one or more of the functional activities of a native AAP.

Biologically active portions of an AAP may have an amino acid sequence shown in SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16, or substantially homologous to SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16, and retains the functional activity of the protein of SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16, yet differs in amino acid sequence due to natural allelic variation or mutagenesis. Other biologically active AAP may comprise an amino acid sequence at least 45% homologous to the amino acid sequence of SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16, and retains the functional activity of native AAP.

5. *Determining homology between two or more sequences*

“AAP variant” means an active AAP having at least: (1) about 80% amino acid sequence identity with a full-length native sequence AAP sequence, (2) an AAP sequence lacking the signal peptide, (3) an extracellular domain of an AAP, with or without the signal peptide, or (4) any other fragment of a full-length AAP sequence. For example, AAP variants include an AAP wherein one or more amino acid residues are added or deleted at the N- or C- terminus of the full-length native amino acid sequence. An AAP variant will have at least about 80% amino acid sequence identity, preferably at least about 81% amino acid sequence identity, more preferably at least about 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% amino acid sequence identity and most preferably at least about 99% amino acid sequence identity with a full-length native sequence AAP sequence. An AAP variant may have a

sequence lacking the signal peptide, an extracellular domain of an AAP, with or without the signal peptide, or any other fragment of a full-length AAP sequence. Ordinarily, AAP variant polypeptides are at least about 10 amino acids in length, often at least about 20 amino acids in length, more often at least about 30, 40, 50, 60, 70, 80, 90, 100, 150, 200, or 300 amino acids in length, or more.

“Percent (%) amino acid sequence identity” is defined as the percentage of amino acid residues that are identical with amino acid residues in a disclosed AAP sequence in a candidate sequence when the two sequences are aligned. To determine % amino acid identity, sequences are aligned and if necessary, gaps are introduced to achieve the maximum % sequence identity; conservative substitutions are not considered as part of the sequence identity. Amino acid sequence alignment procedures to determine percent identity are well known to those of skill in the art. Often publicly available computer software such as BLAST, BLAST2, ALIGN2 or Megalign (DNASTAR) software is used to align peptide sequences. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full length of the sequences being compared.

When amino acid sequences are aligned, the % amino acid sequence identity of a given amino acid sequence A to, with, or against a given amino acid sequence B (which can alternatively be phrased as a given amino acid sequence A that has or comprises a certain % amino acid sequence identity to, with, or against a given amino acid sequence B) can be calculated as:

$$\% \text{amino acid sequence identity} = X/Y \cdot 100$$

where

X is the number of amino acid residues scored as identical matches by the sequence alignment program's or algorithm's alignment of A and B

and

Y is the total number of amino acid residues in B.

If the length of amino acid sequence A is not equal to the length of amino acid sequence B, the % amino acid sequence identity of A to B will not equal the % amino acid sequence identity of B to A.

6. *Chimeric and fusion proteins*

Fusion polypeptides are useful in expression studies, cell-localization, bioassays, and AAP purification. An AAP "chimeric protein" or "fusion protein" comprises an AAP fused to a non-AAP polypeptide. A non-AAP polypeptide is not substantially homologous to an AAP (SEQ ID NOS:2, 4, 6, 8, 10, 12, 14 or 16). An AAP fusion protein may include any portion to an entire AAP, including any number of the biologically active portions. An AAP may be fused to the C-terminus of the GST (glutathione S-transferase) sequences. Such fusion proteins facilitate the purification of a recombinant AAP. In certain host cells, (*e.g.* mammalian), heterologous signal sequences fusions may ameliorate AAP expression and/or secretion. Additional exemplary fusions are presented in Table C.

Other fusion partners can adapt an AAP therapeutically. Fusions with members of the immunoglobulin (Ig) protein family are useful in therapies that inhibit an AAP ligand or substrate interactions, consequently suppressing an AAP-mediated signal transduction *in vivo*. Such fusions, incorporated into pharmaceutical compositions, may be used to treat proliferative and differentiation disorders, as well as modulating cell survival. An AAP-Ig fusion polypeptides can also be used as immunogens to produce an anti-AAP Abs in a subject, to purify AAP ligands, and to screen for molecules that inhibit interactions of an AAP with other molecules.

Fusion proteins can be easily created using recombinant methods. A nucleic acid encoding an AAP can be fused in-frame with a non-AAP encoding nucleic acid, to an AAP NH₂- or COO- -terminus, or internally. Fusion genes may also be synthesized by conventional techniques, including automated DNA synthesizers. PCR amplification using anchor primers that give rise to complementary overhangs between two consecutive gene fragments that can subsequently be annealed and reamplified to generate a chimeric gene sequence (Ausubel *et al.*, 1987) is also useful. Many vectors are commercially available that facilitate sub-cloning an AAP in-frame to a fusion moiety.

Table C Useful non-AAP fusion polypeptides

Reporter	<i>in vitro</i>	<i>in vivo</i>	Notes	Reference
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Human growth hormone (hGH)	Radioimmunoassay	none	Expensive, insensitive, narrow linear range.	(Selden <i>et al.</i> , 1986)
β -glucuronidase (GUS)	Colorimetric, fluorescent, or chemiluminescent	colorimetric (histo-chemical staining with X-gluc)	sensitive, broad linear range, non-isotopic.	(Gallagher, 1992)
Green fluorescent protein (GFP) and related molecules (RFP, BFP, AAP, <i>etc.</i>)	Fluorescent	fluorescent	can be used in live cells; resists photobleaching	(Chalfie <i>et al.</i> , 1994)
Luciferase (firefly)	bioluminescent	Bio-luminescent	protein is unstable, difficult to reproduce, signal is brief	(de Wet <i>et al.</i> , 1987)
Chloramphenicol acetyltransferase (CAT)	Chromatography, differential extraction, fluorescent, or immunoassay	none	Expensive radioactive substrates, time-consuming, insensitive, narrow linear range	(Gorman <i>et al.</i> , 1982)
β -galactosidase	colorimetric, fluorescence, chemiluminescence	colorimetric (histochemical staining with X-gal), bioluminescent in live cells	sensitive, broad linear range; some cells have high endogenous activity	(Alam and Cook, 1990)
Secreted alkaline phosphatase (SEAP)	colorimetric, bioluminescent, chemiluminescent	none	Chemiluminescence assay is sensitive and broad linear range; some cells have endogenous alkaline phosphatase activity	(Berger <i>et al.</i> , 1988)

1. *Agonists and antagonists*

“Antagonist” includes any molecule that partially or fully blocks, inhibits, or neutralizes a biological activity of an endogenous AAP. Similarly, “agonist” includes any molecule that mimics a biological activity of an endogenous AAP. Molecules that can act as agonists or antagonists include Abs or antibody fragments, fragments or variants of an endogenous AAP, peptides, antisense oligonucleotides, small organic molecules, *etc.*

2. *Identifying antagonists and agonists*

To assay for antagonists, an AAP is added to, or expressed in, a cell along with the compound to be screened for a particular activity. If the compound inhibits the activity of interest in the presence of an AAP, that compound is an antagonist to the AAP; if an AAP activity is enhanced, the compound is an agonist.

(a) *Specific examples of potential antagonists and agonist*

Any molecule that alters AAP cellular effects is a candidate antagonist or agonist. Screening techniques well known to those skilled in the art can identify these molecules. Examples of antagonists and agonists include: (1) small organic and inorganic compounds, (2) small peptides, (3) Abs and derivatives, (4) polypeptides closely related to an AAP, (5) antisense DNA and RNA, (6) ribozymes, (7) triple DNA helices and (8) nucleic acid aptamers.

Small molecules that bind to an AAP active site or other relevant part of the polypeptide and inhibit the biological activity of the AAP are antagonists. Examples of small molecule antagonists include small peptides, peptide-like molecules, preferably soluble, and synthetic non-peptidyl organic or inorganic compounds. These same molecules, if they enhance an AAP activity, are examples of agonists.

Almost any antibody that affects an AAP’s function is a candidate antagonist, and occasionally, agonist. Examples of antibody antagonists include polyclonal, monoclonal, single-chain, anti-idiotypic, chimeric Abs, or humanized versions of such Abs or fragments. Abs may be from any species in which an immune response can be raised. Humanized Abs are also contemplated.

Alternatively, a potential antagonist or agonist may be a closely related protein, for example, a mutated form of an AAP that recognizes an AAP-interacting protein but imparts no effect, thereby competitively inhibiting AAP action. Alternatively, a mutated AAP may be constitutively activated and may act as an agonist.

Antisense RNA or DNA constructs can be effective antagonists. Antisense RNA or DNA molecules block function by inhibiting translation by hybridizing to targeted mRNA. Antisense technology can be used to control gene expression through triple-helix formation or antisense DNA or RNA, both of which depend on polynucleotide binding to DNA or RNA. For example, the 5' coding portion of an *AAP* sequence is used to design an antisense RNA oligonucleotide of from about 10 to 40 base pairs in length. A DNA oligonucleotide is designed to be complementary to a region of the gene involved in transcription (triple helix) (Beal and Dervan, 1991; Cooney *et al.*, 1988; Lee *et al.*, 1979), thereby preventing transcription and the production of the AAP. The antisense RNA oligonucleotide hybridizes to the mRNA *in vivo* and blocks translation of the mRNA molecule into the AAP (antisense) (Cohen, 1989; Okano *et al.*, 1991). These oligonucleotides can also be delivered to cells such that the antisense RNA or DNA may be expressed *in vivo* to inhibit production of the AAP. When antisense DNA is used, oligodeoxyribonucleotides derived from the translation-initiation site, *e.g.*, between about -10 and +10 positions of the target gene nucleotide sequence, are preferred.

Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. Ribozymes act by sequence-specific hybridization to the complementary target RNA, followed by endonucleolytic cleavage. Specific ribozyme cleavage sites within a potential RNA target can be identified by known techniques (WO 97/33551, 1997; Rossi, 1994).

To inhibit transcription, triple-helix nucleic acids that are single-stranded and comprise deoxynucleotides are useful antagonists. These oligonucleotides are designed such that triple-helix formation via Hoogsteen base-pairing rules is promoted, generally requiring stretches of purines or pyrimidines (WO 97/33551, 1997).

Aptamers are short oligonucleotide sequences that can be used to recognize and specifically bind almost any molecule. The systematic evolution of ligands by exponential enrichment (SELEX) process (Ausubel *et al.*, 1987; Ellington and Szostak, 1990; Tuerk and Gold, 1990) is powerful and can be used to find such aptamers. Aptamers have many diagnostic and clinical uses; almost any use in which an antibody has been used clinically or diagnostically, aptamers too may be used. In addition, they are cheaper to make once they have been identified, and can be easily applied in a variety

of formats, including administration in pharmaceutical compositions, in bioassays, and diagnostic tests (Jayasena, 1999).

Anti-AAP Abs

The invention encompasses Abs and antibody fragments, such as F_{ab} or $(F_{ab})_2$, that bind immunospecifically to any AAP epitopes.

“Antibody” (Ab) comprises single Abs directed against an AAP (anti-AAP Ab; including agonist, antagonist, and neutralizing Abs), anti-AAP Ab compositions with poly-epitope specificity, single chain anti-AAP Abs, and fragments of anti-AAP Abs. A “monoclonal antibody” is obtained from a population of substantially homogeneous Abs, *i.e.*, the individual Abs comprising the population are identical except for possible naturally-occurring mutations that may be present in minor amounts. Exemplary Abs include polyclonal (pAb), monoclonal (mAb), humanized, bi-specific (bsAb), and heteroconjugate Abs.

1. *Polyclonal Abs (pAbs)*

Polyclonal Abs can be raised in a mammalian host, for example, by one or more injections of an immunogen and, if desired, an adjuvant. Typically, the immunogen and/or adjuvant are injected in the mammal by multiple subcutaneous or intraperitoneal injections. The immunogen may include an AAP or a fusion protein. Examples of adjuvants include Freund’s complete and monophosphoryl Lipid A synthetic-trehalose dicorynomycolate (MPL-TDM). To improve the immune response, an immunogen may be conjugated to a protein that is immunogenic in the host, such as keyhole limpet hemocyanin (KLH), serum albumin, bovine thyroglobulin, and soybean trypsin inhibitor. Protocols for antibody production are described by (Ausubel *et al.*, 1987; Harlow and Lane, 1988). Alternatively, pAbs may be made in chickens, producing IgY molecules (Schade *et al.*, 1996).

2. *Monoclonal Abs (mAbs)*

Anti-AAP mAbs may be prepared using hybridoma methods (Milstein and Cuello, 1983). Hybridoma methods comprise at least four steps: (1) immunizing a host, or lymphocytes from a host; (2) harvesting the mAb secreting (or potentially secreting) lymphocytes, (3) fusing the lymphocytes to immortalized cells, and (4) selecting those cells that secrete the desired (anti-AAP) mAb.

A mouse, rat, guinea pig, hamster, or other appropriate host is immunized to elicit lymphocytes that produce or are capable of producing Abs that will specifically bind to the immunogen. Alternatively, the lymphocytes may be immunized *in vitro*. If human cells are desired, peripheral blood lymphocytes (PBLs) are generally used; however, spleen cells or lymphocytes from other mammalian sources are preferred. The immunogen typically includes an AAP or a fusion protein.

The lymphocytes are then fused with an immortalized cell line to form hybridoma cells, facilitated by a fusing agent such as polyethylene glycol (Goding, 1996). Rodent, bovine, or human myeloma cells immortalized by transformation may be used, or rat or mouse myeloma cell lines. Because pure populations of hybridoma cells and not unfused immortalized cells are preferred, the cells after fusion are grown in a suitable medium that contains one or more substances that inhibit the growth or survival of unfused, immortalized cells. A common technique uses parental cells that lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT). In this case, hypoxanthine, aminopterin and thymidine are added to the medium (HAT medium) to prevent the growth of HGPRT-deficient cells while permitting hybridomas to grow.

Preferred immortalized cells fuse efficiently, can be isolated from mixed populations by selecting in a medium such as HAT, and support stable and high-level expression of antibody after fusion. Preferred immortalized cell lines are murine myeloma lines, available from the American Type Culture Collection (Manassas, VA). Human myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human mAbs (Kozbor *et al.*, 1984; Schook, 1987).

Because hybridoma cells secrete antibody extracellularly, the culture media can be assayed for the presence of mAbs directed against an AAP (anti-AAP mAbs). Immunoprecipitation or *in vitro* binding assays, such as radio immunoassay (RIA) or enzyme-linked immunoabsorbent assay (ELISA), measure the binding specificity of mAbs (Harlow and Lane, 1988; Harlow and Lane, 1999), including Scatchard analysis (Munson and Rodbard, 1980).

Anti-AAP mAb secreting hybridoma cells may be isolated as single clones by limiting dilution procedures and sub-cultured (Goding, 1996). Suitable culture media include Dulbecco's Modified Eagle's Medium, RPMI-1640, or if desired, a protein-free

or -reduced or serum-free medium (*e.g.*, Ultra DOMA PF or HL-1; Biowhittaker; Walkersville, MD). The hybridoma cells may also be grown *in vivo* as ascites.

The mAbs may be isolated or purified from the culture medium or ascites fluid by conventional Ig purification procedures such as protein A-Sepharose, hydroxylapatite chromatography, gel electrophoresis, dialysis, ammonium sulfate precipitation or affinity chromatography (Harlow and Lane, 1988; Harlow and Lane, 1999).

The mAbs may also be made by recombinant methods (U.S. Patent No. 4166452, 1979). DNA encoding anti-AAP mAbs can be readily isolated and sequenced using conventional procedures, *e.g.*, using oligonucleotide probes that specifically bind to murine heavy and light antibody chain genes, to probe preferably DNA isolated from anti-AAP-secreting mAb hybridoma cell lines. Once isolated, the isolated DNA fragments are sub-cloned into expression vectors that are then transfected into host cells such as simian COS-7 cells, Chinese hamster ovary (CHO) cells, or myeloma cells that do not otherwise produce Ig protein, to express mAbs. The isolated DNA fragments can be modified, for example, by substituting the coding sequence for human heavy and light chain constant domains in place of the homologous murine sequences (U.S. Patent No. 4816567, 1989; Morrison *et al.*, 1987), or by fusing the Ig coding sequence to all or part of the coding sequence for a non-Ig polypeptide. Such a non-Ig polypeptide can be substituted for the constant domains of an antibody, or can be substituted for the variable domains of one antigen-combining site to create a chimeric bivalent antibody.

3. *Monovalent Abs*

The Abs may be monovalent Abs that consequently do not cross-link with each other. For example, one method involves recombinant expression of Ig light chain and modified heavy chain. Heavy chain truncations generally at any point in the F_c region will prevent heavy chain cross-linking. Alternatively, the relevant cysteine residues are substituted with another amino acid residue or are deleted, preventing crosslinking. *In vitro* methods are also suitable for preparing monovalent Abs. Abs can be digested to produce fragments, such as F_{ab} fragments (Harlow and Lane, 1988; Harlow and Lane, 1999).

4. *Humanized and human Abs*

Anti-AAP Abs may further comprise humanized or human Abs. Humanized forms of non-human Abs are chimeric Igs, Ig chains or fragments (such as F_v , F_{ab} , F_{ab}' ,

$F_{(ab')_2}$ or other antigen-binding subsequences of Abs) that contain minimal sequence derived from non-human Ig.

Generally, a humanized antibody has one or more amino acid residues introduced from a non-human source. These non-human amino acid residues are often referred to as “import” residues, which are typically taken from an “import” variable domain. Humanization is accomplished by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody (Jones *et al.*, 1986; Riechmann *et al.*, 1988; Verhoeyen *et al.*, 1988). Such “humanized” Abs are chimeric Abs (U.S. Patent No. 4816567, 1989), wherein substantially less than an intact human variable domain has been substituted by the corresponding sequence from a non-human species. In practice, humanized Abs are typically human Abs in which some CDR residues and possibly some FR residues are substituted by residues from analogous sites in rodent Abs. Humanized Abs include human Igs (recipient antibody) in which residues from a complementary determining region (CDR) of the recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit, having the desired specificity, affinity and capacity. In some instances, corresponding non-human residues replace F_v framework residues of the human Ig. Humanized Abs may comprise residues that are found neither in the recipient antibody nor in the imported CDR or framework sequences. In general, the humanized antibody comprises substantially all of at least one, and typically two, variable domains, in which most if not all of the CDR regions correspond to those of a non-human Ig and most if not all of the FR regions are those of a human Ig consensus sequence. The humanized antibody optimally also comprises at least a portion of an Ig constant region (F_c), typically that of a human Ig (Jones *et al.*, 1986; Presta, 1992; Riechmann *et al.*, 1988).

Human Abs can also be produced using various techniques, including phage display libraries (Hoogenboom *et al.*, 1991; Marks *et al.*, 1991) and the preparation of human mAbs (Boerner *et al.*, 1991; Reisfeld and Sell, 1985). Similarly, introducing human Ig genes into transgenic animals in which the endogenous Ig genes have been partially or completely inactivated can be exploited to synthesize human Abs. Upon challenge, human antibody production is observed, which closely resembles that seen in humans in all respects, including gene rearrangement, assembly, and antibody repertoire (U.S. Patent No. 5545807, 1996; U.S. Patent No. 5545806, 1996; U.S. Patent No.

5569825, 1996; U.S. Patent No. 5633425, 1997; U.S. Patent No. 5661016, 1997; U.S. Patent No. 5625126, 1997; Fishwild *et al.*, 1996; Lonberg and Huszar, 1995; Lonberg *et al.*, 1994; Marks *et al.*, 1992).

5. *Bi-specific mAbs*

Bi-specific Abs are monoclonal, preferably human or humanized, that have binding specificities for at least two different antigens. For example, a binding specificity is an AAP; the other is for any antigen of choice, preferably a cell-surface protein or receptor or receptor subunit.

Traditionally, the recombinant production of bi-specific Abs is based on the co-expression of two Ig heavy-chain/light-chain pairs, where the two heavy chains have different specificities (Milstein and Cuello, 1983). Because of the random assortment of Ig heavy and light chains, the resulting hybridomas (quadromas) produce a potential mixture of ten different antibody molecules, of which only one has the desired bi-specific structure. The desired antibody can be purified using affinity chromatography or other techniques (WO 93/08829, 1993; Traunecker *et al.*, 1991).

To manufacture a bi-specific antibody (Suresh *et al.*, 1986), variable domains with the desired antibody-antigen combining sites are fused to Ig constant domain sequences. The fusion is preferably with an Ig heavy-chain constant domain, comprising at least part of the hinge, CH2, and CH3 regions. Preferably, the first heavy-chain constant region (CH1) containing the site necessary for light-chain binding is in at least one of the fusions. DNAs encoding the Ig heavy-chain fusions and, if desired, the Ig light chain, are inserted into separate expression vectors and are co-transfected into a suitable host organism.

The interface between a pair of antibody molecules can be engineered to maximize the percentage of heterodimers that are recovered from recombinant cell culture (WO 96/27011, 1996). The preferred interface comprises at least part of the CH3 region of an antibody constant domain. In this method, one or more small amino acid side chains from the interface of the first antibody molecule are replaced with larger side chains (*e.g.* tyrosine or tryptophan). Compensatory "cavities" of identical or similar size to the large side chain(s) are created on the interface of the second antibody molecule by replacing large amino acid side chains with smaller ones (*e.g.* alanine or threonine). This

mechanism increases the yield of the heterodimer over unwanted end products such as homodimers.

Bi-specific Abs can be prepared as full length Abs or antibody fragments (*e.g.* $F_{(ab')_2}$ bi-specific Abs). One technique to generate bi-specific Abs exploits chemical linkage. Intact Abs can be proteolytically cleaved to generate $F_{(ab')_2}$ fragments (Brennan *et al.*, 1985). Fragments are reduced with a dithiol complexing agent, such as sodium arsenite, to stabilize vicinal dithiols and prevent intermolecular disulfide formation. The generated $F_{ab'}$ fragments are then converted to thionitrobenzoate (TNB) derivatives. One of the $F_{ab'}$ -TNB derivatives is then reconverted to the $F_{ab'}$ -thiol by reduction with mercaptoethylamine and is mixed with an equimolar amount of the other $F_{ab'}$ -TNB derivative to form the bi-specific antibody. The produced bi-specific Abs can be used as agents for the selective immobilization of enzymes.

$F_{ab'}$ fragments may be directly recovered from *E. coli* and chemically coupled to form bi-specific Abs. For example, fully humanized bi-specific $F_{(ab')_2}$ Abs can be produced (Shalaby *et al.*, 1992). Each $F_{ab'}$ fragment is separately secreted from *E. coli* and directly coupled chemically *in vitro*, forming the bi-specific antibody.

Various techniques for making and isolating bi-specific antibody fragments directly from recombinant cell culture have also been described. For example, leucine zipper motifs can be exploited (Kostelny *et al.*, 1992). Peptides from the *Fos* and *Jun* proteins are linked to the $F_{ab'}$ portions of two different Abs by gene fusion. The antibody homodimers are reduced at the hinge region to form monomers and then re-oxidized to form antibody heterodimers. This method can also produce antibody homodimers. The "diabody" technology (Holliger *et al.*, 1993) provides an alternative method to generate bi-specific antibody fragments. The fragments comprise a heavy-chain variable domain (V_H) connected to a light-chain variable domain (V_L) by a linker that is too short to allow pairing between the two domains on the same chain. The V_H and V_L domains of one fragment are forced to pair with the complementary V_L and V_H domains of another fragment, forming two antigen-binding sites. Another strategy for making bi-specific antibody fragments is the use of single-chain F_v (sF_v) dimers (Gruber *et al.*, 1994). Abs with more than two valencies are also contemplated, such as tri-specific Abs (Tutt *et al.*, 1991).

Exemplary bi-specific Abs may bind to two different epitopes on a given AAP. Alternatively, cellular defense mechanisms can be restricted to a particular cell expressing the particular AAP: an anti-AAP arm may be combined with an arm that binds to a leukocyte triggering molecule, such as a T-cell receptor molecule (*e.g.* CD2, CD3, CD28, or B7), or to F_c receptors for IgG (F_cγR), such as F_cγRI (CD64), F_cγRII (CD32) and F_cγRIII (CD16). Bi-specific Abs may also be used to target cytotoxic agents to cells that express a particular AAP. These Abs possess an AAP-binding arm and an arm that binds a cytotoxic agent or a radionuclide chelator.

6. *Heteroconjugate Abs*

Heteroconjugate Abs, consisting of two covalently joined Abs, have been proposed to target immune system cells to unwanted cells (4,676,980, 1987) and for treatment of human immunodeficiency virus (HIV) infection (WO 91/00360, 1991; WO 92/20373, 1992). Abs prepared *in vitro* using synthetic protein chemistry methods, including those involving cross-linking agents, are contemplated. For example, immunotoxins may be constructed using a disulfide exchange reaction or by forming a thioether bond. Examples of suitable reagents include iminothiolate and methyl-4-mercaptobutyrimidate (4,676,980, 1987).

7. *Immunoconjugates*

Immunoconjugates may comprise an antibody conjugated to a cytotoxic agent such as a chemotherapeutic agent, toxin (*e.g.*, an enzymatically active toxin or fragment of bacterial, fungal, plant, or animal origin), or a radioactive isotope (*i.e.*, a radioconjugate).

Useful enzymatically-active toxins and fragments include Diphtheria A chain, non-binding active fragments of Diphtheria toxin, exotoxin A chain from *Pseudomonas aeruginosa*, ricin A chain, abrin A chain, modeccin A chain, α-sarcin, *Aleurites fordii* proteins, Dianthin proteins, *Phytolaca americana* proteins, *Momordica charantia* inhibitor, curcin, crotin, *Sapaonaria officinalis* inhibitor, gelonin, mitogellin, restrictocin, phenomycin, enomycin, and the tricothecenes. A variety of radionuclides are available for the production of radioconjugated Abs, such as ²¹²Bi, ¹³¹I, ¹³¹In, ⁹⁰Y, and ¹⁸⁶Re.

Conjugates of the antibody and cytotoxic agent are made using a variety of bi-functional protein-coupling agents, such as N-succinimidyl-3-(2-pyridyldithiol) propionate (SPDP), iminothiolane (IT), bi-functional derivatives of imidoesters (such as

dimethyl adipimidate HCl), active esters (such as disuccinimidyl suberate), aldehydes (such as glutaraldehyde), *bis*-azido compounds (such as bis (p-azidobenzoyl) hexanediamine), *bis*-diazonium derivatives (such as bis-(p-diazoniumbenzoyl)-ethylenediamine), diisocyanates (such as tolyene 2,6- diisocyanate), and *bis*-active fluorine compounds (such as 1,5-difluoro-2,4-dinitrobenzene). For example, a ricin immunotoxin can be prepared (Vitetta *et al.*, 1987). ¹⁴C-labeled 1-isothiocyanatobenzyl-3-methyldiethylene triaminepentaacetic acid (MX-DTPA) is an exemplary chelating agent for conjugating radionuclide to antibody (WO 94/11026, 1994).

In another embodiment, the antibody may be conjugated to a “receptor” (such as streptavidin) for utilization in tumor pre-targeting wherein the antibody-receptor conjugate is administered to the patient, followed by removal of unbound conjugate from the circulation using a clearing agent and then administration of a streptavidin “ligand” (*e.g.*, biotin) that is conjugated to a cytotoxic agent (*e.g.*, a radionuclide).

8. *Effector function engineering*

The antibody can be modified to enhance its effectiveness in treating a disease, such as cancer. For example, cysteine residue(s) may be introduced into the F_c region, thereby allowing interchain disulfide bond formation in this region. Such homodimeric Abs may have improved internalization capability and/or increased complement-mediated cell killing and antibody-dependent cellular cytotoxicity (ADCC) (Caron *et al.*, 1992; Shopes, 1992). Homodimeric Abs with enhanced anti-tumor activity can be prepared using hetero-bifunctional cross-linkers (Wolff *et al.*, 1993). Alternatively, an antibody engineered with dual F_c regions may have enhanced complement lysis (Stevenson *et al.*, 1989).

9. *Immunoliposomes*

Liposomes containing the antibody may also be formulated (U.S. Patent No. 4485045, 1984; U.S. Patent No. 4544545, 1985; U.S. Patent No. 5013556, 1991; Eppstein *et al.*, 1985; Hwang *et al.*, 1980). Useful liposomes can be generated by a reverse-phase evaporation method with a lipid composition comprising phosphatidylcholine, cholesterol, and PEG-derivatized phosphatidylethanolamine (PEG- PE). Such preparations are extruded through filters of defined pore size to yield liposomes with a desired diameter. F_{ab} fragments of the antibody can be conjugated to the liposomes (Martin and Papahadjopoulos, 1982) via a disulfide-interchange reaction. A

chemotherapeutic agent, such as Doxorubicin, may also be contained in the liposome (Gabizon *et al.*, 1989). Other useful liposomes with different compositions are contemplated.

10. *Diagnostic applications of Abs directed against an AAP*

Anti-AAP Abs can be used to localize and/or quantitate an AAP (*e.g.*, for use in measuring levels of an AAP within tissue samples or for use in diagnostic methods, *etc.*). Anti-AAP epitope Abs can be utilized as pharmacologically-active compounds.

Anti-AAP Abs can be used to isolate an AAP by standard techniques, such as immunoaffinity chromatography or immunoprecipitation. These approaches facilitate purifying an endogenous AAP antigen-containing polypeptides from cells and tissues. These approaches, as well as others, can be used to detect an AAP in a sample to evaluate the abundance and pattern of expression of the antigenic protein. Anti-AAP Abs can be used to monitor protein levels in tissues as part of a clinical testing procedure; for example, to determine the efficacy of a given treatment regimen. Coupling the antibody to a detectable substance (label) allows detection of Ab-antigen complexes. Classes of labels include fluorescent, luminescent, bioluminescent, and radioactive materials, enzymes and prosthetic groups. Useful labels include horseradish peroxidase, alkaline phosphatase, β -galactosidase, acetylcholinesterase, streptavidin/biotin, avidin/biotin, umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride, phycoerythrin, luminol, luciferase, luciferin, aequorin, and ^{125}I , ^{131}I , ^{35}S or ^3H .

11. *Antibody therapeutics*

Abs of the invention, including polyclonal, monoclonal, humanized and fully human Abs, can be used therapeutically. Such agents will generally be employed to treat or prevent a disease or pathology in a subject. An antibody preparation, preferably one having high antigen specificity and affinity generally mediates an effect by binding the target epitope(s). Generally, administration of such Abs may mediate one of two effects: (1) the antibody may prevent ligand binding, eliminating endogenous ligand binding and subsequent signal transduction, or (2) the antibody elicits a physiological result by binding an effector site on the target molecule, initiating signal transduction.

A therapeutically effective amount of an antibody relates generally to the amount needed to achieve a therapeutic objective, epitope binding affinity, administration rate,

and depletion rate of the antibody from a subject. Common ranges for therapeutically effective doses may be, as a nonlimiting example, from about 0.1 mg/kg body weight to about 50 mg/kg body weight. Dosing frequencies may range, for example, from twice daily to once a week.

12. *Pharmaceutical compositions of Abs*

Anti-AAP Abs, as well as other AAP interacting molecules (such as aptamers) identified in other assays, can be administered in pharmaceutical compositions to treat various disorders. Principles and considerations involved in preparing such compositions, as well as guidance in the choice of components can be found in (de Boer, 1994; Gennaro, 2000; Lee, 1990).

Since some AAP are intracellular, Abs that are internalized are preferred used when whole Abs are used as inhibitors. Liposomes may also be used as a delivery vehicle for intracellular introduction. Where antibody fragments are used, the smallest inhibitory fragment that specifically binds to the epitope is preferred. For example, peptide molecules can be designed that bind a preferred epitope based on the variable-region sequences of a useful antibody. Such peptides can be synthesized chemically and/or produced by recombinant DNA technology (Marasco *et al.*, 1993). Formulations may also contain more than one active compound for a particular treatment, preferably those with activities that do not adversely affect each other. The composition may comprise an agent that enhances function, such as a cytotoxic agent, cytokine, chemotherapeutic agent, or growth-inhibitory agent.

The active ingredients can also be entrapped in microcapsules prepared by coacervation techniques or by interfacial polymerization; for example, hydroxymethylcellulose or gelatin-microcapsules and poly-(methylmethacrylate) microcapsules, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles, and nanocapsules) or in macroemulsions.

The formulations to be used for *in vivo* administration are highly preferred to be sterile. This is readily accomplished by filtration through sterile filtration membranes or any of a number of techniques.

Sustained-release preparations may also be prepared, such as semi-permeable matrices of solid hydrophobic polymers containing the antibody, which matrices are in

the form of shaped articles, *e.g.*, films, or microcapsules. Examples of sustained-release matrices include polyesters, hydrogels (for example, poly(2-hydroxyethyl-methacrylate), or poly(vinylalcohol)), polylactides (Boswell and Scribner, U.S. Patent No. 3,773,919, 1973), copolymers of L-glutamic acid and γ ethyl-L-glutamate, non-degradable ethylene-vinyl acetate, degradable lactic acid-glycolic acid copolymers such as injectable microspheres composed of lactic acid-glycolic acid copolymer, and poly-D-(-)-3-hydroxybutyric acid. While polymers such as ethylene-vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods and may be preferred.

AAP recombinant expression vectors and host cells

Vectors are tools used to shuttle DNA between host cells or as a means to express a nucleotide sequence. Some vectors function only in prokaryotes, while others function in both prokaryotes and eukaryotes, enabling large-scale DNA preparation from prokaryotes for expression in eukaryotes. Inserting the DNA of interest, such as an AAP nucleotide sequence or a fragment, is accomplished by ligation techniques and/or mating protocols well-known to the skilled artisan. Such DNA is inserted such that its integration does not disrupt any necessary components of the vector. In the case of vectors that are used to express the inserted DNA protein, the introduced DNA is operably-linked to the vector elements that govern its transcription and translation.

Vectors can be divided into two general classes: Cloning vectors are replicating plasmid or phage with regions that are non-essential for propagation in an appropriate host cell, and into which foreign DNA can be inserted; the foreign DNA is replicated and propagated as if it were a component of the vector. An expression vector (such as a plasmid, yeast, or animal virus genome) is used to introduce foreign genetic material into a host cell or tissue in order to transcribe and translate the foreign DNA. In expression vectors, the introduced DNA is operably-linked to elements, such as promoters, that signal to the host cell to transcribe the inserted DNA. Some promoters are exceptionally useful, such as inducible promoters that control gene transcription in response to specific factors. Operably-linking an AAP or anti-sense construct to an inducible promoter can control the expression of an AAP or fragments, or anti-sense constructs. Examples of classic inducible promoters include those that are responsive to α -interferon, heat-shock,

heavy metal ions, and steroids such as glucocorticoids (Kaufman, 1990) and tetracycline. Other desirable inducible promoters include those that are not endogenous to the cells in which the construct is being introduced, but, however, is responsive in those cells when the induction agent is exogenously supplied.

Vectors have many difference manifestations. A "plasmid" is a circular double stranded DNA molecule into which additional DNA segments can be introduced. Viral vectors can accept additional DNA segments into the viral genome. Certain vectors are capable of autonomous replication in a host cell (*e.g.*, bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (*e.g.*, non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. In general, useful expression vectors are often plasmids. However, other forms of expression vectors, such as viral vectors (*e.g.*, replication defective retroviruses, adenoviruses and adeno-associated viruses) are contemplated.

Recombinant expression vectors that comprise an *AAP* (or fragments) regulate an *AAP* transcription by exploiting one or more host cell-responsive (or that can be manipulated *in vitro*) regulatory sequences that is operably-linked to an *AAP*. "Operably-linked" indicates that a nucleotide sequence of interest is linked to regulatory sequences such that expression of the nucleotide sequence is achieved.

Vectors can be introduced in a variety of organisms and/or cells (Table D). Alternatively, the vectors can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

Table D Examples of hosts for cloning or expression

Organisms	Examples	Sources and References*
Prokaryotes		
Enterobacteriaceae	<i>E. coli</i>	
	K 12 strain MM294	ATCC 31,446
	X1776	ATCC 31,537
	W3110	ATCC 27,325
	K5 772	ATCC 53,635
	<i>Enterobacter</i>	
	<i>Erwinia</i>	
	<i>Klebsiella</i>	
	<i>Proteus</i>	

Table D Examples of hosts for cloning or expression

Organisms	Examples	Sources and References*
	<i>Salmonella</i> (<i>S. typhimurium</i>)	
	<i>Serratia</i> (<i>S. marcescens</i>)	
	<i>Shigella</i>	
	<i>Bacilli</i> (<i>B. subtilis</i> and <i>B. licheniformis</i>)	
	<i>Pseudomonas</i> (<i>P. aeruginosa</i>)	
	<i>Streptomyces</i>	
Eukaryotes		
Yeasts	<i>Saccharomyces cerevisiae</i>	
	<i>Schizosaccharomyces pombe</i>	
	<i>Kluyveromyces</i>	(Fleer <i>et al.</i> , 1991)
	<i>K. lactis</i> MW98-8C, CBS683, CBS4574	(de Louvencourt <i>et al.</i> , 1983)
	<i>K. fragilis</i>	ATCC 12,424
	<i>K. bulgaricus</i>	ATCC 16,045
	<i>K. wickerhamii</i>	ATCC 24,178
	<i>K. waltii</i>	ATCC 56,500
	<i>K. drosophilae</i>	ATCC 36,906
	<i>K. thermotolerans</i>	
	<i>K. marxianus</i> ; <i>yarrowia</i>	(EPO 402226, 1990)
	<i>Pichia pastoris</i>	(Sreekrishna <i>et al.</i> , 1988)
	<i>Candida</i>	
	<i>Trichoderma reesei</i>	
Filamentous Fungi	<i>Neurospora crassa</i>	(Case <i>et al.</i> , 1979)
	<i>Torulopsis</i>	
	<i>Rhodotorula</i>	
	<i>Schwanniomyces</i> (<i>S. occidentalis</i>)	
Filamentous Fungi	<i>Neurospora</i>	
	<i>Penicillium</i>	
	<i>Tolypocladium</i>	(WO 91/00357, 1991)
	<i>Aspergillus</i> (<i>A. nidulans</i> and <i>A. niger</i>)	(Kelly and Hynes, 1985; Tilburn <i>et al.</i> , 1983; Yelton <i>et al.</i> , 1984)
Invertebrate cells	<i>Drosophila</i> S2	
	<i>Spodoptera</i> Sf9	
Vertebrate cells	Chinese Hamster Ovary (CHO)	
	simian COS	
	COS-7	ATCC CRL 1651
	HEK 293	

*Unreferenced cells are generally available from American Type Culture Collection (Manassas, VA).

Vector choice is dictated by the organism or cells being used and the desired fate of the vector. Vectors may replicate once in the target cells, or may be “suicide” vectors. In general, vectors comprise signal sequences, origins of replication, marker genes, enhancer elements, promoters, and transcription termination sequences. The choice of these elements depends on the organisms in which the vector will be used and are easily determined. Some of these elements may be conditional, such as an inducible or conditional promoter that is turned “on” when conditions are appropriate. Examples of inducible promoters include those that are tissue-specific, which relegate expression to certain cell types, steroid-responsive, or heat-shock reactive. Some bacterial repression systems, such as the *lac* operon, have been exploited in mammalian cells and transgenic animals (Fieck *et al.*, 1992; Wyborski *et al.*, 1996; Wyborski and Short, 1991). Vectors often use a selectable marker to facilitate identifying those cells that have incorporated the vector. Many selectable markers are well known in the art for the use with prokaryotes, usually antibiotic-resistance genes or the use of autotrophy and auxotrophy mutants.

Using antisense and sense AAP oligonucleotides can prevent an AAP polypeptide expression. These oligonucleotides bind to target nucleic acid sequences, forming duplexes that block transcription or translation of the target sequence by enhancing degradation of the duplexes, terminating prematurely transcription or translation, or by other means.

Antisense or sense oligonucleotides are single-stranded nucleic acids, either RNA or DNA, which can bind a target AAP mRNA (sense) or an AAP DNA (antisense) sequences. According to the present invention, antisense or sense oligonucleotides comprise a fragment of an AAP DNA coding region of at least about 14 nucleotides, preferably from about 14 to 30 nucleotides. In general, antisense RNA or DNA molecules can comprise at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100 bases in length or more. Among others, (Stein and Cohen, 1988; van der Krol *et al.*, 1988a) describe methods to derive antisense or a sense oligonucleotides from a given cDNA sequence.

Modifications of antisense and sense oligonucleotides can augment their effectiveness. Modified sugar-phosphodiester bonds or other sugar linkages (WO

91/06629, 1991), increase *in vivo* stability by conferring resistance to endogenous nucleases without disrupting binding specificity to target sequences. Other modifications can increase the affinities of the oligonucleotides for their targets, such as covalently linked organic moieties (WO 90/10448, 1990) or poly-(L)-lysine. Other attachments modify binding specificities of the oligonucleotides for their targets, including metal complexes or intercalating (*e.g.* ellipticine) and alkylating agents.

To introduce antisense or sense oligonucleotides into target cells (cells containing the target nucleic acid sequence), any gene transfer method may be used and are well known to those of skill in the art. Examples of gene transfer methods include 1) biological, such as gene transfer vectors like Epstein-Barr virus or conjugating the exogenous DNA to a ligand-binding molecule (WO 91/04753, 1991), 2) physical, such as electroporation, and 3) chemical, such as CaPO₄ precipitation and oligonucleotide-lipid complexes (WO 90/10448, 1990).

The terms "host cell" and "recombinant host cell" are used interchangeably. Such terms refer not only to a particular subject cell but also to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term.

Methods of eukaryotic cell transfection and prokaryotic cell transformation are well known in the art. The choice of host cell will dictate the preferred technique for introducing the nucleic acid of interest. Table E, which is not meant to be limiting, summarizes many of the known techniques in the art. Introduction of nucleic acids into an organism may also be done with *ex vivo* techniques that use an *in vitro* method of transfection, as well as established genetic techniques, if any, for that particular organism.

Table E Methods to introduce nucleic acid into cells

Cells	Methods	References	Notes
Prokaryotes (bacteria)	Calcium chloride	(Cohen <i>et al.</i> , 1972; Hanahan, 1983; Mandel and Higa, 1970)	
	Electroporation	(Shigekawa and Dower, 1988)	
Eukaryotes			

Table E Methods to introduce nucleic acid into cells

Cells	Methods	References	Notes
Mammalian cells	Calcium phosphate transfection	<i>N</i> -(2-Hydroxyethyl)piperazine- <i>N'</i> -(2-ethanesulfonic acid (HEPES) buffered saline solution (Chen and Okayama, 1988; Graham and van der Eb, 1973; Wigler <i>et al.</i> , 1978) BES (<i>N,N</i> -bis(2-hydroxyethyl)-2-aminoethanesulfonic acid) buffered solution (Ishiura <i>et al.</i> , 1982)	Cells may be "shocked" with glycerol or dimethylsulfoxide (DMSO) to increase transfection efficiency (Ausubel <i>et al.</i> , 1987).
	Diethylaminoethyl (DEAE)-Dextran transfection	(Fujita <i>et al.</i> , 1986; Lopata <i>et al.</i> , 1984; Selden <i>et al.</i> , 1986)	Most useful for transient, but not stable, transfections. Chloroquine can be used to increase efficiency.
	Electroporation	(Neumann <i>et al.</i> , 1982; Potter, 1988; Potter <i>et al.</i> , 1984; Wong and Neumann, 1982)	Especially useful for hard-to-transfect lymphocytes.
	Cationic lipid reagent transfection	(Elroy-Stein and Moss, 1990; Felgner <i>et al.</i> , 1987; Rose <i>et al.</i> , 1991; Whitt <i>et al.</i> , 1990)	Applicable to both <i>in vivo</i> and <i>in vitro</i> transfection.
	Retroviral	Production exemplified by (Cepko <i>et al.</i> , 1984; Miller and Buttimore, 1986; Pear <i>et al.</i> , 1993) Infection <i>in vitro</i> and <i>in vivo</i> : (Austin and Cepko, 1990; Bodine <i>et al.</i> , 1991; Fekete and Cepko, 1993; Lemischka <i>et al.</i> , 1986; Turner <i>et al.</i> , 1990; Williams <i>et al.</i> , 1984)	Lengthy process, many packaging lines available at ATCC. Applicable to both <i>in vivo</i> and <i>in vitro</i> transfection.
	Polybrene	(Chaney <i>et al.</i> , 1986; Kawai and Nishizawa, 1984)	
	Microinjection	(Capecchi, 1980)	Can be used to establish cell lines carrying integrated copies of AAP DNA sequences.

Table E Methods to introduce nucleic acid into cells

Cells	Methods	References	Notes
	Protoplast fusion	(Rassoulzadegan <i>et al.</i> , 1982; Sandri-Goldin <i>et al.</i> , 1981; Schaffner, 1980)	
Insect cells (<i>in vitro</i>)	Baculovirus systems	(Luckow, 1991; Miller, 1988; O'Reilly <i>et al.</i> , 1992)	Useful for <i>in vitro</i> production of proteins with eukaryotic modifications.
Yeast	Electroporation	(Becker and Guarente, 1991)	
	Lithium acetate	(Gietz <i>et al.</i> , 1998; Ito <i>et al.</i> , 1983)	
	Spheroplast fusion	(Beggs, 1978; Hinnen <i>et al.</i> , 1978)	Laborious, can produce aneuploids.
Plant cells (general reference: (Hansen and Wright, 1999))	Agrobacterium transformation	(Bechtold and Pelletier, 1998; Escudero and Hohn, 1997; Hansen and Chilton, 1999; Touraev and al., 1997)	
	Biolistics (microprojectiles)	(Finer <i>et al.</i> , 1999; Hansen and Chilton, 1999; Shillito, 1999)	
	Electroporation (protoplasts)	(Fromm <i>et al.</i> , 1985; Ou-Lee <i>et al.</i> , 1986; Rhodes <i>et al.</i> , 1988; Saunders <i>et al.</i> , 1989) May be combined with liposomes (Trick and al., 1997)	
	Polyethylene glycol (PEG) treatment	(Shillito, 1999)	
	Liposomes	May be combined with electroporation (Trick and al., 1997)	
	<i>in planta</i> microinjection	(Leduc and al., 1996; Zhou and al., 1983)	
	Seed imbibition	(Trick and al., 1997)	
	Laser beam	(Hoffman, 1996)	
	Silicon carbide whiskers	(Thompson and al., 1995)	

Vectors often use a selectable marker to facilitate identifying those cells that have incorporated the vector. Many selectable markers are well known in the art for the use with prokaryotes, usually antibiotic-resistance genes or the use of autotrophy and

auxotrophy mutants. Table F lists often-used selectable markers for mammalian cell transfection.

Table F Useful selectable markers for eukaryote cell transfection

Selectable Marker	Selection	Action	Reference
Adenosine deaminase (ADA)	Media includes 9- β -D-xylofuranosyl adenine (Xyl-A)	Conversion of Xyl-A to Xyl-ATP, which incorporates into nucleic acids, killing cells. ADA detoxifies	(Kaufman <i>et al.</i> , 1986)
Dihydrofolate reductase (DHFR)	Methotrexate (MTX) and dialyzed serum (purine-free media)	MTX competitive inhibitor of DHFR. In absence of exogenous purines, cells require DHFR, a necessary enzyme in purine biosynthesis.	(Simonsen and Levinson, 1983)
Aminoglycoside phosphotransferase ("APH", "neo", "G418")	G418	G418, an aminoglycoside detoxified by APH, interferes with ribosomal function and consequently, translation.	(Southern and Berg, 1982)
Hygromycin-B-phosphotransferase (HPH)	hygromycin-B	Hygromycin-B, an aminocyclitol detoxified by HPH, disrupts protein translocation and promotes mistranslation.	(Palmer <i>et al.</i> , 1987)
Thymidine kinase (TK)	Forward selection (TK+): Media (HAT) incorporates aminopterin. Reverse selection (TK-): Media incorporates 5-bromodeoxyuridine (BrdU).	Forward: Aminopterin forces cells to synthesize dTTP from thymidine, a pathway requiring TK. Reverse: TK phosphorylates BrdU, which incorporates into nucleic acids, killing cells.	(Littlefield, 1964)

A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce an AAP. Accordingly, the invention provides methods for producing an AAP using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of the invention (into which a recombinant expression vector encoding an AAP has been introduced) in a suitable medium, such that an AAP is produced. In another embodiment, the method further comprises isolating an AAP from the medium or the host cell.

Transgenic AAP animals

Transgenic animals are useful for studying the function and/or activity of an AAP and for identifying and/or evaluating modulators of AAP activity. "Transgenic animals" are non-human animals, preferably mammals, more preferably a rodents such as rats or mice, in which one or more of the cells include a transgene. Other transgenic animals include primates, sheep, dogs, cows, goats, chickens, amphibians, *etc.* A "transgene" is exogenous DNA that is integrated into the genome of a cell from which a transgenic animal develops, and that remains in the genome of the mature animal. Transgenes preferably direct the expression of an encoded gene product in one or more cell types or tissues of the transgenic animal with the purpose of preventing expression of a naturally encoded gene product in one or more cell types or tissues (a "knockout" transgenic animal), or serving as a marker or indicator of an integration, chromosomal location, or region of recombination (*e.g.* *cre/loxP* mice). A "homologous recombinant animal" is a non-human animal, such as a rodent, in which an endogenous AAP has been altered by an exogenous DNA molecule that recombines homologously with an endogenous AAP in a (*e.g.* embryonic) cell prior to development the animal. Host cells with an exogenous AAP can be used to produce non-human transgenic animals, such as fertilized oocytes or embryonic stem cells into which an AAP-coding sequences have been introduced. Such host cells can then be used to create non-human transgenic animals or homologous recombinant animals.

1. Approaches to transgenic animal production

A transgenic animal can be created by introducing an AAP into the male pronuclei of a fertilized oocyte (*e.g.*, by microinjection, retroviral infection) and allowing the oocyte to develop in a pseudopregnant female foster animal (pffa). An AAP cDNA sequences (SEQ ID NO:1, 3, 5, 7, 9, 11, 13 or 15) can be introduced as a transgene into the genome

of a non-human animal. Alternatively, a homologue of an *AAP*, such as the naturally-occurring variant of an *AAP*, can be used as a transgene. Intronic sequences and polyadenylation signals can also be included in the transgene to increase transgene expression. Tissue-specific regulatory sequences can be operably-linked to the *AAP* transgene to direct expression of the *AAP* to particular cells. Methods for generating transgenic animals via embryo manipulation and microinjection, particularly animals such as mice, have become conventional in the art, *e.g.* (Evans *et al.*, U.S. Patent No. 4,870,009, 1989; Hogan, 0879693843, 1994; Leder and Stewart, U.S. Patent No. 4,736,866, 1988; Wagner and Hoppe, US Patent No. 4,873,191, 1989). Other non-mice transgenic animals may be made by similar methods. A transgenic founder animal, which can be used to breed additional transgenic animals, can be identified based upon the presence of the transgene in its genome and/or expression of the transgene mRNA in tissues or cells of the animals. Transgenic animals can be bred to other transgenic animals carrying other transgenes.

2. *Vectors for transgenic animal production*

To create a homologous recombinant animal, a vector containing at least a portion of an *AAP* into which a deletion, addition or substitution has been introduced to thereby alter, *e.g.*, disrupt or alter the expression of, an *AAP*. An *AAP* can be a murine gene, or other *AAP* homologue, such as a naturally occurring variant. In one approach, a knockout vector functionally disrupts an endogenous *AAP* gene upon homologous recombination, and thus a non-functional *AAP* protein, if any, is expressed.

Alternatively, the vector can be designed such that, upon homologous recombination, an endogenous *AAP* is mutated or otherwise altered but still encodes functional protein (*e.g.*, the upstream regulatory region can be altered to thereby alter the expression of an endogenous *AAP*). In this type of homologous recombination vector, the altered portion of the *AAP* is flanked at its 5'- and 3'-termini by additional nucleic acid of the *AAP* to allow for homologous recombination to occur between the exogenous *AAP* carried by the vector and an endogenous *AAP* in an embryonic stem cell. The additional flanking *AAP* nucleic acid is sufficient to engender homologous recombination with the endogenous *AAP*. Typically, several kilobases of flanking DNA (both at the 5'- and 3'-termini) are included in the vector (Thomas and Capecchi, 1987). The vector is then introduced into an embryonic stem cell line (*e.g.*, by electroporation), and cells in which

the introduced AAP has homologously-recombined with an endogenous AAP are selected (Li *et al.*, 1992).

3. *Introduction of an AAP transgene cells during development*

Selected cells are then injected into a blastocyst of an animal (*e.g.*, a mouse) to form aggregation chimeras (Bradley, 1987). A chimeric embryo can then be implanted into a suitable pffa and the embryo brought to term. Progeny harboring the homologously-recombined DNA in their germ cells can be used to breed animals in which all cells of the animal contain the homologously-recombined DNA by germline transmission of the transgene. Methods for constructing homologous recombination vectors and homologous recombinant animals are described (Berns *et al.*, WO 93/04169, 1993; Bradley, 1991; Kucherlapati *et al.*, WO 91/01140, 1991; Le Mouellic and Brullet, WO 90/11354, 1990).

Alternatively, transgenic animals that contain selected systems that allow for regulated expression of the transgene can be produced. An example of such a system is the *cre/loxP* recombinase system of bacteriophage P1 (Lakso *et al.*, 1992). Another recombinase system is the FLP recombinase system of *Saccharomyces cerevisiae* (O'Gorman *et al.*, 1991). If a *cre/loxP* recombinase system is used to regulate expression of the transgene, animals containing transgenes encoding both the *Cre* recombinase and a selected protein are required. Such animals can be produced as "double" transgenic animals, by mating an animal containing a transgene encoding a selected protein to another containing a transgene encoding a recombinase.

Clones of transgenic animals can also be produced (Wilmot *et al.*, 1997). In brief, a cell from a transgenic animal can be isolated and induced to exit the growth cycle and enter G₀ phase. The quiescent cell can then be fused to an enucleated oocyte from an animal of the same species from which the quiescent cell is isolated. The reconstructed oocyte is then cultured to develop to a morula or blastocyte and then transferred to a pffa. The offspring borne of this female foster animal will be a clone of the "parent" transgenic animal.

Pharmaceutical compositions

The AAP nucleic acid molecules, AAP polypeptides, and anti-AAP Abs (active compounds) of the invention, and derivatives, fragments, analogs and homologs thereof, can be incorporated into pharmaceutical compositions. Such compositions typically

comprise the nucleic acid molecule, protein, or antibody and a pharmaceutically acceptable carrier. A "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration (Gennaro, 2000). Preferred examples of such carriers or diluents include, but are not limited to, water, saline, finger's solutions, dextrose solution, and 5% human serum albumin. Liposomes and non-aqueous vehicles such as fixed oils may also be used. Except when a conventional media or agent is incompatible with an active compound, use of these compositions is contemplated. Supplementary active compounds can also be incorporated into the compositions.

1. *General considerations*

A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration, including intravenous, intradermal, subcutaneous, oral (*e.g.*, inhalation), transdermal (*i.e.*, topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid (EDTA); buffers such as acetates, citrates or phosphates, and agents for the adjustment of tonicity such as sodium chloride or dextrose. The pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

2. *Injectable formulations*

Pharmaceutical compositions suitable for injection include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, CREMOPHOR EL[™] (BASF, Parsippany, N.J.) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid so as to be administered using a syringe. Such compositions should be stable during manufacture and storage and must be

preserved against contamination from microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (such as glycerol, propylene glycol, and liquid polyethylene glycol), and suitable mixtures. Proper fluidity can be maintained, for example, by using a coating such as lecithin, by maintaining the required particle size in the case of dispersion and by using surfactants. Various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, and thimerosal, can contain microorganism contamination. Isotonic agents, for example, sugars, polyalcohols such as manitol, sorbitol, and sodium chloride can be included in the composition. Compositions that can delay absorption include agents such as aluminum monostearate and gelatin.

Sterile injectable solutions can be prepared by incorporating the active compound (*e.g.*, an AAP or anti-AAP antibody) in the required amount in an appropriate solvent with one or a combination of ingredients as required, followed by sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle that contains a basic dispersion medium, and the other required ingredients as discussed. Sterile powders for the preparation of sterile injectable solutions, methods of preparation include vacuum drying and freeze-drying that yield a powder containing the active ingredient and any desired ingredient from a sterile solutions.

3. *Oral compositions*

Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally. Pharmaceutically compatible binding agents, and/or adjuvant materials can be included. Tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, PRIMOGEL, or corn starch; a lubricant such as magnesium stearate or STEROTES; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

4. *Compositions for inhalation*

For administration by inhalation, the compounds are delivered as an aerosol spray from a nebulizer or a pressurized container that contains a suitable propellant, *e.g.*, a gas such as carbon dioxide.

5. *Systemic administration*

Systemic administration can also be transmucosal or transdermal. For transmucosal or transdermal administration, penetrants that can permeate the target barrier(s) are selected. Transmucosal penetrants include, detergents, bile salts, and fusidic acid derivatives. Nasal sprays or suppositories can be used for transmucosal administration. For transdermal administration, the active compounds are formulated into ointments, salves, gels, or creams.

The compounds can also be prepared in the form of suppositories (*e.g.*, with bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

6. *Carriers*

In one embodiment, the active compounds are prepared with carriers that protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Such materials can be obtained commercially from ALZA Corporation (Mountain View, CA) and NOVA Pharmaceuticals, Inc. (Lake Elsinore, CA), or prepared by one of skill in the art. Liposomal suspensions can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, such as in (Eppstein *et al.*, US Patent No. 4,522,811, 1985).

7. *Unit dosage*

Oral formulations or parenteral compositions in unit dosage form can be created to facilitate administration and dosage uniformity. Unit dosage form refers to physically discrete units suited as single dosages for the subject to be treated, containing a therapeutically effective quantity of active compound in association with the required pharmaceutical carrier. The specification for the unit dosage forms of the invention are dictated by, and directly dependent on, the unique characteristics of the active compound

and the particular desired therapeutic effect, and the inherent limitations of compounding the active compound.

8. *Gene therapy compositions*

The nucleic acid molecules of the invention can be inserted into vectors and used as gene therapy vectors. Gene therapy vectors can be delivered to a subject by, for example, intravenous injection, local administration (Nabel and Nabel, US Patent No. 5,328,470, 1994), or by stereotactic injection (Chen *et al.*, 1994). The pharmaceutical preparation of a gene therapy vector can include an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery vector can be produced intact from recombinant cells, *e.g.*, retroviral vectors, the pharmaceutical preparation can include one or more cells that produce the gene delivery system.

9. *Dosage*

The pharmaceutical composition and method of the present invention may further comprise other therapeutically active compounds as noted herein that are usually applied in the treatment of the above mentioned pathological conditions.

In the treatment or prevention of conditions which require AAP modulation an appropriate dosage level will generally be about 0.01 to 500 mg per kg patient body weight per day which can be administered in single or multiple doses. Preferably, the dosage level will be about 0.1 to about 250 mg/kg per day; more preferably about 0.5 to about 100 mg/kg per day. A suitable dosage level may be about 0.01 to 250 mg/kg per day, about 0.05 to 100 mg/kg per day, or about 0.1 to 50 mg/kg per day. Within this range the dosage may be 0.05 to 0.5, 0.5 to 5 or 5 to 50 mg/kg per day. For oral administration, the compositions are preferably provided in the form of tablets containing 1.0 to 1000 milligrams of the active ingredient, particularly 1.0, 5.0, 10.0, 15.0, 20.0, 25.0, 50.0, 75.0, 100.0, 150.0, 200.0, 250.0, 300.0, 400.0, 500.0, 600.0, 750.0, 800.0, 900.0, and 1000.0 milligrams of the active ingredient for the symptomatic adjustment of the dosage to the patient to be treated. The compounds may be administered on a regimen of 1 to 4 times per day, preferably once or twice per day.

It will be understood, however, that the specific dose level and frequency of dosage for any particular patient may be varied and will depend upon a variety of factors including the activity of the specific compound employed, the metabolic stability and

length of action of that compound, the age, body weight, general health, sex, diet, mode and time of administration, rate of excretion, drug combination, the severity of the particular condition, and the host undergoing therapy.

10. *Kits for pharmaceutical compositions*

The pharmaceutical compositions can be included in a kit, container, pack, or dispenser together with instructions for administration. When the invention is supplied as a kit, the different components of the composition may be packaged in separate containers and admixed immediately before use. Such packaging of the components separately may permit long-term storage without losing the active components' functions.

Kits may also include reagents in separate containers that facilitate the execution of a specific test, such as diagnostic tests or tissue typing. For example, AAP DNA templates and suitable primers may be supplied for internal controls.

(a) *Containers or vessels*

The reagents included in the kits can be supplied in containers of any sort such that the life of the different components are preserved, and are not adsorbed or altered by the materials of the container. For example, sealed glass ampules may contain lyophilized luciferase or buffer that have been packaged under a neutral, non-reacting gas, such as nitrogen. Ampoules may consist of any suitable material, such as glass, organic polymers, such as polycarbonate, polystyrene, etc., ceramic, metal or any other material typically employed to hold reagents. Other examples of suitable containers include simple bottles that may be fabricated from similar substances as ampules, and envelopes, that may consist of foil-lined interiors, such as aluminum or an alloy. Other containers include test tubes, vials, flasks, bottles, syringes, or the like. Containers may have a sterile access port, such as a bottle having a stopper that can be pierced by a hypodermic injection needle. Other containers may have two compartments that are separated by a readily removable membrane that upon removal permits the components to mix. Removable membranes may be glass, plastic, rubber, etc.

(b) *Instructional materials*

Kits may also be supplied with instructional materials. Instructions may be printed on paper or other substrate, and/or may be supplied as an electronic-readable medium, such as a floppy disc, CD-ROM, DVD-ROM, Zip disc, videotape, audiotape, etc. Detailed instructions may not be physically associated with the kit; instead, a user

may be directed to an internet web site specified by the manufacturer or distributor of the kit, or supplied as electronic mail.

Screening and detection methods

The isolated nucleic acid molecules of the invention can be used to express an AAP (*e.g.*, via a recombinant expression vector in a host cell in gene therapy applications), to detect an AAP mRNA (*e.g.*, in a biological sample) or a genetic lesion in an AAP, and to modulate AAP activity, as described below. In addition, AAP polypeptides can be used to screen drugs or compounds that modulate the AAP activity or expression as well as to treat disorders characterized by insufficient or excessive production of an AAP or production of AAP forms that have decreased or aberrant activity compared to an AAP wild-type protein, or modulate biological function that involve an AAP. In addition, the anti-AAP Abs of the invention can be used to detect and isolate an AAP and modulate AAP activity.

1. Screening assays

The invention provides a method (screening assay) for identifying modalities, *i.e.*, candidate or test compounds or agents (*e.g.*, peptides, peptidomimetics, small molecules or other drugs), foods, combinations thereof, *etc.*, that effect an AAP, a stimulatory or inhibitory effect, including translation, transcription, activity or copies of the gene in cells. The invention also includes compounds identified in screening assays.

Testing for compounds that increase or decrease AAP activity are desirable. A compound may modulate an AAP activity by affecting: (1) the number of copies of the gene in the cell (amplifiers and deamplifiers); (2) increasing or decreasing transcription of an AAP (transcription up-regulators and down-regulators); (3) by increasing or decreasing the translation of an AAP mRNA into protein (translation up-regulators and down-regulators); or (4) by increasing or decreasing the activity of an AAP itself (agonists and antagonists).

(a) effects of compounds

To identify compounds that affect an AAP at the DNA, RNA and protein levels, cells or organisms are contacted with a candidate compound and the corresponding change in an AAP DNA, RNA or protein is assessed (Ausubel *et al.*, 1987). For DNA amplifiers and deamplifiers, the amount of an AAP DNA is measured, for those

compounds that are transcription up-regulators and down-regulators the amount of an AAP mRNA is determined; for translational up- and down-regulators, the amount of an AAP polypeptide is measured. Compounds that are agonists or antagonists may be identified by contacting cells or organisms with the compound, and then examining, for example, the model of angiogenesis *in vitro*.

In one embodiment, many assays for screening candidate or test compounds that bind to or modulate the activity of an AAP or polypeptide or biologically-active portion are available. Test compounds can be obtained using any of the numerous approaches in combinatorial library methods, including: biological libraries; spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring deconvolution; the "one-bead one-compound" library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to peptides, while the other four approaches encompass peptide, non-peptide oligomer or small molecule libraries of compounds (Lam, 1997).

(b) *small molecules*

A "small molecule" refers to a composition that has a molecular weight of less than about 5 kD and more preferably less than about 4 kD, most preferably less than 0.6 kD. Small molecules can be, nucleic acids, peptides, polypeptides, peptidomimetics, carbohydrates, lipids or other organic or inorganic molecules. Libraries of chemical and/or biological mixtures, such as fungal, bacterial, or algal extracts, are known in the art and can be screened with any of the assays of the invention. Examples of methods for the synthesis of molecular libraries can be found in: (Carell *et al.*, 1994a; Carell *et al.*, 1994b; Cho *et al.*, 1993; DeWitt *et al.*, 1993; Gallop *et al.*, 1994; Zuckermann *et al.*, 1994).

Libraries of compounds may be presented in solution (Houghten *et al.*, 1992) or on beads (Lam *et al.*, 1991), on chips (Fodor *et al.*, 1993), bacteria, spores (Ladner *et al.*, US Patent No. 5,223,409, 1993), plasmids (Cull *et al.*, 1992) or on phage (Cwirla *et al.*, 1990; Devlin *et al.*, 1990; Felici *et al.*, 1991; Ladner *et al.*, US Patent No. 5,223,409, 1993; Scott and Smith, 1990). A cell-free assay comprises contacting an AAP or biologically-active fragment with a known compound that binds the AAP to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with the AAP, where determining the ability of the

test compound to interact with the AAP comprises determining the ability of the AAP to preferentially bind to or modulate the activity of an AAP target molecule.

(c) *cell-free assays*

The cell-free assays of the invention may be used with both soluble or a membrane-bound forms of an AAP. In the case of cell-free assays comprising the membrane-bound form, a solubilizing agent to maintain the AAP in solution. Examples of such solubilizing agents include non-ionic detergents such as n-octylglucoside, n-dodecylglucoside, n-dodecylmaltoside, octanoyl-N-methylglucamide, decanoyl-N-methylglucamide, TRITON[®] X-100 and others from the TRITON[®] series, THESIT[®], Isotridecypoly(ethylene glycol ether)_n, N-dodecyl--N,N-dimethyl-3-ammonio-1-propane sulfonate, 3-(3-cholamidopropyl) dimethylamminiol-1-propane sulfonate (CHAPS), or 3-(3-cholamidopropyl)dimethylamminiol-2-hydroxy-1-propane sulfonate (CHAPSO).

(d) *immobilization of target molecules to facilitate screening*

In more than one embodiment of the assay methods, immobilizing either an AAP or its partner molecules can facilitate separation of complexed from uncomplexed forms of one or both of the proteins, as well as to accommodate high throughput assays. Binding of a test compound to an AAP, or interaction of an AAP with a target molecule in the presence and absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants, such as microtiter plates, test tubes, and micro-centrifuge tubes. A fusion protein can be provided that adds a domain that allows one or both of the proteins to be bound to a matrix. For example, GST-AAP fusion proteins or GST-target fusion proteins can be adsorbed onto glutathione sepharose beads (SIGMA Chemical, St. Louis, MO) or glutathione derivatized microtiter plates that are then combined with the test compound or the test compound and either the non-adsorbed target protein or an AAP, and the mixture is incubated under conditions conducive to complex formation (e.g., at physiological conditions for salt and pH). Following incubation, the beads or microtiter plate wells are washed to remove any unbound components, the matrix immobilized in the case of beads, complex determined either directly or indirectly, for example, as described. Alternatively, the complexes can be dissociated from the matrix, and the level of AAP binding or activity determined using standard techniques.

Other techniques for immobilizing proteins on matrices can also be used in screening assays. Either an AAP or its target molecule can be immobilized using biotin-avidin or biotin-streptavidin systems. Biotinylation can be accomplished using many reagents, such as biotin-NHS (N-hydroxy-succinimide; PIERCE Chemicals, Rockford, IL), and immobilized in wells of streptavidin-coated 96 well plates (PIERCE Chemical). Alternatively, Abs reactive with an AAP or target molecules, but which do not interfere with binding of the AAP to its target molecule, can be derivatized to the wells of the plate, and unbound target or an AAP trapped in the wells by antibody conjugation. Methods for detecting such complexes, in addition to those described for the GST-immobilized complexes, include immunodetection of complexes using Abs reactive with an AAP or its target, as well as enzyme-linked assays that rely on detecting an enzymatic activity associated with the AAP or target molecule.

(e) *screens to identify modulators*

Modulators of AAP expression can be identified in a method where a cell is contacted with a candidate compound and the expression of an AAP mRNA or protein in the cell is determined. The expression level of the AAP mRNA or protein in the presence of the candidate compound is compared to the AAP mRNA or protein levels in the absence of the candidate compound. The candidate compound can then be identified as a modulator of the AAP mRNA or protein expression based upon this comparison. For example, when expression of an AAP mRNA or protein is greater (*i.e.*, statistically significant) in the presence of the candidate compound than in its absence, the candidate compound is identified as a stimulator of the AAP mRNA or protein expression. Alternatively, when expression of the AAP mRNA or protein is less (statistically significant) in the presence of the candidate compound than in its absence, the candidate compound is identified as an inhibitor of the AAP mRNA or protein expression. The level of an AAP mRNA or protein expression in the cells can be determined by methods described for detecting an AAP mRNA or protein.

(i) *hybrid assays*

In yet another aspect of the invention, an AAP can be used as "bait" in two-hybrid or three hybrid assays (Bartel *et al.*, 1993; Brent *et al.*, WO94/10300, 1994; Iwabuchi *et al.*, 1993; Madura *et al.*, 1993; Saifer *et al.*, US Patent No. 5,283,317, 1994; Zervos *et al.*, 1993) to identify other proteins that bind or interact with the AAP and modulate AAP

activity. Such AAP-bps are also likely to be involved in the propagation of signals by the AAP as, for example, upstream or downstream elements of an AAP pathway.

The two-hybrid system is based on the modular nature of most transcription factors, which consist of separable DNA-binding and activation domains. Briefly, the assay utilizes two different DNA constructs. In one construct, the gene that codes for an AAP is fused to a gene encoding the DNA binding domain of a known transcription factor (*e.g.*, GAL4). The other construct, a DNA sequence from a library of DNA sequences that encodes an unidentified protein ("prey" or "sample") is fused to a gene that codes for the activation domain of the known transcription factor. If the "bait" and the "prey" proteins are able to interact *in vivo*, forming an AAP-dependent complex, the DNA-binding and activation domains of the transcription factor are brought into close proximity. This proximity allows transcription of a reporter gene (*e.g.*, LacZ) that is operably-linked to a transcriptional regulatory site responsive to the transcription factor. Expression of the reporter gene can be detected, and cell colonies containing the functional transcription factor can be isolated and used to obtain the cloned gene that encodes the AAP-interacting protein.

The invention further pertains to novel agents identified by the aforementioned screening assays and uses thereof for treatments as described herein.

2. *Detection assays*

Portions or fragments of an AAP cDNA sequences identified herein (and the complete AAP gene sequences) are useful in themselves. By way of non-limiting example, these sequences can be used to: (1) identify an individual from a minute biological sample (tissue typing); and (2) aid in forensic identification of a biological sample.

(a) *Tissue typing*

The AAP sequences of the invention can be used to identify individuals from minute biological samples. In this technique, an individual's genomic DNA is digested with one or more restriction enzymes and probed on a Southern blot to yield unique bands. The sequences of the invention are useful as additional DNA markers for "restriction fragment length polymorphisms" (RFLP; (Smulson *et al.*, US Patent No. 5,272,057, 1993)).

Furthermore, the AAP sequences can be used to determine the actual base-by-base DNA sequence of targeted portions of an individual's genome. AAP sequences can be used to prepare two PCR primers from the 5'- and 3'-termini of the sequences that can then be used to amplify an the corresponding sequences from an individual's genome and then sequence the amplified fragment.

Panels of corresponding DNA sequences from individuals can provide unique individual identifications, as each individual will have a unique set of such DNA sequences due to allelic differences. The sequences of the invention can be used to obtain such identification sequences from individuals and from tissue. The AAP sequences of the invention uniquely represent portions of an individual's genome. Allelic variation occurs to some degree in the coding regions of these sequences, and to a greater degree in the noncoding regions. The allelic variation between individual humans occurs with a frequency of about once ever 500 bases. Much of the allelic variation is due to single nucleotide polymorphisms (SNPs), which include RFLPs.

Each of the sequences described herein can, to some degree, be used as a standard against which DNA from an individual can be compared for identification purposes. Because greater numbers of polymorphisms occur in noncoding regions, fewer sequences are necessary to differentiate individuals. Noncoding sequences can positively identify individuals with a panel of 10 to 1,000 primers that each yield a noncoding amplified sequence of 100 bases. If predicted coding sequences, such as those in SEQ ID NOS:1, 3, 5, 7, 9, 11, 13, and 15 are used, a more appropriate number of primers for positive individual identification would be 500-2,000.

Predictive medicine

The invention also pertains to the field of predictive medicine in which diagnostic assays, prognostic assays, pharmacogenomics, and monitoring clinical trials are used for prognostic (predictive) purposes to treat an individual prophylactically. Accordingly, one aspect of the invention relates to diagnostic assays for determining an AAP and/or nucleic acid expression as well as AAP activity, in the context of a biological sample (*e.g.*, blood, serum, cells, tissue) to determine whether an individual is afflicted with a disease or disorder, or is at risk of developing a disorder, associated with aberrant AAP expression or activity, including cancer. The invention also provides for prognostic (or predictive)

assays for determining whether an individual is at risk of developing a disorder associated with an AAP, nucleic acid expression or activity. For example, mutations in an AAP can be assayed in a biological sample. Such assays can be used for prognostic or predictive purpose to prophylactically treat an individual prior to the onset of a disorder characterized by or associated with the AAP, nucleic acid expression, or biological activity.

Another aspect of the invention provides methods for determining AAP activity, or nucleic acid expression, in an individual to select appropriate therapeutic or prophylactic agents for that individual (referred to herein as "pharmacogenomics"). Pharmacogenomics allows for the selection of modalities (*e.g.*, drugs, foods) for therapeutic or prophylactic treatment of an individual based on the individual's genotype (*e.g.*, the individual's genotype to determine the individual's ability to respond to a particular agent). Another aspect of the invention pertains to monitoring the influence of modalities (*e.g.*, drugs, foods) on the expression or activity of an AAP in clinical trials.

1. *Diagnostic assays*

An exemplary method for detecting the presence or absence of an AAP in a biological sample involves obtaining a biological sample from a subject and contacting the biological sample with a compound or an agent capable of detecting the AAP or the AAP nucleic acid (*e.g.*, mRNA, genomic DNA) such that the presence of the AAP is confirmed in the sample. An agent for detecting the AAP mRNA or genomic DNA is a labeled nucleic acid probe that can hybridize to the AAP mRNA or genomic DNA. The nucleic acid probe can be, for example, a full-length AAP nucleic acid, such as the nucleic acid of SEQ ID NOS: 1, 3, 5, 7, 9, 11, 13 or 15, or a portion thereof, such as an oligonucleotide of at least 15, 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to an AAP mRNA or genomic DNA.

An agent for detecting an AAP polypeptide is an antibody capable of binding to the AAP, preferably an antibody with a detectable label. Abs can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment (*e.g.*, F_{ab} or F(ab')₂) can be used. A labeled probe or antibody is coupled (*i.e.*, physically linking) to a detectable substance, as well as indirect detection of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently-labeled secondary antibody and end-labeling of a

DNA probe with biotin such that it can be detected with fluorescently-labeled streptavidin. The term "biological sample" includes tissues, cells and biological fluids isolated from a subject, as well as tissues, cells and fluids present within a subject. The detection method of the invention can be used to detect an *AAP* mRNA, protein, or genomic DNA in a biological sample *in vitro* as well as *in vivo*. For example, *in vitro* techniques for detection of an *AAP* mRNA include Northern hybridizations and *in situ* hybridizations. *In vitro* techniques for detection of an *AAP* polypeptide include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations, and immunofluorescence. *In vitro* techniques for detection of an *AAP* genomic DNA include Southern hybridizations and fluorescence in situ hybridization (FISH). Furthermore, *in vivo* techniques for detecting an *AAP* include introducing into a subject a labeled anti-*AAP* antibody. For example, the antibody can be labeled with a radioactive marker whose presence and location in a subject can be detected by standard imaging techniques.

In one embodiment, the biological sample from the subject contains protein molecules, and/or mRNA molecules, and/or genomic DNA molecules. A preferred biological sample is blood.

In another embodiment, the methods further involve obtaining a biological sample from a subject to provide a control, contacting the sample with a compound or agent to detect an *AAP*, mRNA, or genomic DNA, and comparing the presence of the *AAP*, mRNA or genomic DNA in the control sample with the presence of the *AAP*, mRNA or genomic DNA in the test sample.

The invention also encompasses kits for detecting an *AAP* in a biological sample. For example, the kit can comprise: a labeled compound or agent capable of detecting an *AAP* or an *AAP* mRNA in a sample; reagent and/or equipment for determining the amount of an *AAP* in the sample; and reagent and/or equipment for comparing the amount of an *AAP* in the sample with a standard. The compound or agent can be packaged in a suitable container. The kit can further comprise instructions for using the kit to detect the *AAP* or nucleic acid.

2. Prognostic assays

The diagnostic methods described herein can furthermore be utilized to identify subjects having or at risk of developing a disease or disorder associated with an aberrant *AAP* expression or activity. For example, the assays described herein, can be used to

identify a subject having or at risk of developing a disorder associated with AAP, nucleic acid expression or activity. Alternatively, the prognostic assays can be used to identify a subject having or at risk for developing a disease or disorder. The invention provides a method for identifying a disease or disorder associated with an aberrant AAP expression or activity in which a test sample is obtained from a subject and the AAP or nucleic acid (*e.g.*, mRNA, genomic DNA) is detected. A test sample is a biological sample obtained from a subject. For example, a test sample can be a biological fluid (*e.g.*, serum), cell sample, or tissue.

Prognostic assays can be used to determine whether a subject can be administered a modality (*e.g.*, an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, food, *etc.*) to treat a disease or disorder associated with an aberrant AAP expression or activity. Such methods can be used to determine whether a subject can be effectively treated with an agent for a disorder. The invention provides methods for determining whether a subject can be effectively treated with an agent for a disorder associated with an aberrant AAP expression or activity in which a test sample is obtained and the AAP or nucleic acid is detected (*e.g.*, where the presence of the AAP or nucleic acid is diagnostic for a subject that can be administered the agent to treat a disorder associated with the aberrant AAP expression or activity).

The methods of the invention can also be used to detect genetic lesions in an AAP to determine if a subject with the genetic lesion is at risk for a disorder characterized by aberrant angiogenesis. Methods include detecting, in a sample from the subject, the presence or absence of a genetic lesion characterized by at an alteration affecting the integrity of a gene encoding an AAP polypeptide, or the mis-expression of an AAP. Such genetic lesions can be detected by ascertaining: (1) a deletion of one or more nucleotides from an AAP; (2) an addition of one or more nucleotides to an AAP; (3) a substitution of one or more nucleotides in an AAP, (4) a chromosomal rearrangement of an AAP gene; (5) an alteration in the level of an AAP mRNA transcripts, (6) aberrant modification of an AAP, such as a change genomic DNA methylation, (7) the presence of a non-wild-type splicing pattern of an AAP mRNA transcript, (8) a non-wild-type level of an AAP, (9) allelic loss of an AAP, and/or (10) inappropriate post-translational modification of an AAP polypeptide. There are a large number of known assay techniques that can be used

to detect lesions in an *AAP*. Any biological sample containing nucleated cells may be used.

In certain embodiments, lesion detection may use a probe/primer in a polymerase chain reaction (PCR) (*e.g.*, (Mullis, US Patent No. 4,683,202, 1987; Mullis *et al.*, US Patent No. 4,683,195, 1987), such as anchor PCR or rapid amplification of cDNA ends (RACE) PCR, or, alternatively, in a ligation chain reaction (LCR) (*e.g.*, (Landegren *et al.*, 1988; Nakazawa *et al.*, 1994), the latter is particularly useful for detecting point mutations in *AAP*-genes (Abravaya *et al.*, 1995). This method may include collecting a sample from a patient, isolating nucleic acids from the sample, contacting the nucleic acids with one or more primers that specifically hybridize to an *AAP* under conditions such that hybridization and amplification of the *AAP* (if present) occurs, and detecting the presence or absence of an amplification product, or detecting the size of the amplification product and comparing the length to a control sample. It is anticipated that PCR and/or LCR may be desirable to use as a preliminary amplification step in conjunction with any of the techniques used for detecting mutations described herein.

Alternative amplification methods include: self sustained sequence replication (Guatelli *et al.*, 1990), transcriptional amplification system (Kwoh *et al.*, 1989); Q β Replicase (Lizardi *et al.*, 1988), or any other nucleic acid amplification method, followed by the detection of the amplified molecules using techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules present in low abundance.

Mutations in an *AAP* from a sample can be identified by alterations in restriction enzyme cleavage patterns. For example, sample and control DNA is isolated, amplified (optionally), digested with one or more restriction endonucleases, and fragment length sizes are determined by gel electrophoresis and compared. Differences in fragment length sizes between sample and control DNA indicates mutations in the sample DNA. Moreover, the use of sequence specific ribozymes can be used to score for the presence of specific mutations by development or loss of a ribozyme cleavage site.

Hybridizing a sample and control nucleic acids, *e.g.*, DNA or RNA, to high-density arrays containing hundreds or thousands of oligonucleotides probes, can identify genetic mutations in an *AAP* (Cronin *et al.*, 1996; Kozal *et al.*, 1996). For example, genetic mutations in an *AAP* can be identified in two-dimensional arrays containing

light-generated DNA probes as described in Cronin, *et al.*, supra. Briefly, a first hybridization array of probes can be used to scan through long stretches of DNA in a sample and control to identify base changes between the sequences by making linear arrays of sequential overlapping probes. This step allows the identification of point mutations. This is followed by a second hybridization array that allows the characterization of specific mutations by using smaller, specialized probe arrays complementary to all variants or mutations detected. Each mutation array is composed of parallel probe sets, one complementary to the wild-type gene and the other complementary to the mutant gene.

In yet another embodiment, any of a variety of sequencing reactions known in the art can be used to directly sequence an AAP and detect mutations by comparing the sequence of the sample AAP-with the corresponding wild-type (control) sequence. Examples of sequencing reactions include those based on classic techniques (Maxam and Gilbert, 1977; Sanger *et al.*, 1977). Any of a variety of automated sequencing procedures can be used when performing diagnostic assays (Naeve *et al.*, 1995) including sequencing by mass spectrometry (Cohen *et al.*, 1996; Griffin and Griffin, 1993; Koster, WO94/16101, 1994).

Other methods for detecting mutations in an AAP include those in which protection from cleavage agents is used to detect mismatched bases in RNA/RNA or RNA/DNA heteroduplexes (Myers *et al.*, 1985). In general, the technique of "mismatch cleavage" starts by providing heteroduplexes formed by hybridizing (labeled) RNA or DNA containing a wild-type AAP sequence with potentially mutant RNA or DNA obtained from a sample. The double-stranded duplexes are treated with an agent that cleaves single-stranded regions of the duplex such as those that arise from base pair mismatches between the control and sample strands. For instance, RNA/DNA duplexes can be treated with RNase and DNA/DNA hybrids treated with S₁ nuclease to enzymatically digest the mismatched regions. In other embodiments, either DNA/DNA or RNA/DNA duplexes can be treated with hydroxylamine or osmium tetroxide and with piperidine in order to digest mismatched regions. The digested material is then separated by size on denaturing polyacrylamide gels to determine the mutation site (Grompe *et al.*, 1989; Saleeba and Cotton, 1993). The control DNA or RNA can be labeled for detection.

Mismatch cleavage reactions may employ one or more proteins that recognize mismatched base pairs in double-stranded DNA (DNA mismatch repair) in defined systems for detecting and mapping point mutations in an *AAP* cDNAs obtained from samples of cells. For example, the *mutY* enzyme of *E. coli* cleaves A at G/A mismatches and the thymidine DNA glycosylase from HeLa cells cleaves T at G/T mismatches (Hsu *et al.*, 1994). According to an exemplary embodiment, a probe based on a wild-type *AAP* sequence is hybridized to a cDNA or other DNA product from a test cell(s). The duplex is treated with a DNA mismatch repair enzyme, and the cleavage products, if any, can be detected from electrophoresis protocols or the like (Modrich *et al.*, US Patent No. 5,459,039, 1995).

Electrophoretic mobility alterations can be used to identify mutations in an *AAP*. For example, single strand conformation polymorphism (SSCP) may be used to detect differences in electrophoretic mobility between mutant and wild type nucleic acids (Cotton, 1993; Hayashi, 1992; Orita *et al.*, 1989). Single-stranded DNA fragments of sample and control *AAP* nucleic acids are denatured and then renatured. The secondary structure of single-stranded nucleic acids varies according to sequence; the resulting alteration in electrophoretic mobility allows detection of even a single base change. The DNA fragments may be labeled or detected with labeled probes. The sensitivity of the assay may be enhanced by using RNA (rather than DNA), in which the secondary structure is more sensitive to a sequence changes. The subject method may use heteroduplex analysis to separate double stranded heteroduplex molecules on the basis of changes in electrophoretic mobility (Keen *et al.*, 1991).

The migration of mutant or wild-type fragments can be assayed using denaturing gradient gel electrophoresis (DGGE; (Myers *et al.*, 1985). In DGGE, DNA is modified to prevent complete denaturation, for example by adding a GC clamp of approximately 40 bp of high-melting GC-rich DNA by PCR. A temperature gradient may also be used in place of a denaturing gradient to identify differences in the mobility of control and sample DNA (Rossiter and Caskey, 1990).

Examples of other techniques for detecting point mutations include, but are not limited to, selective oligonucleotide hybridization, selective amplification, or selective primer extension. For example, oligonucleotide primers may be prepared in which the known mutation is placed centrally and then hybridized to target DNA under conditions

that permit hybridization only if a perfect match is found (Saiki *et al.*, 1986; Saiki *et al.*, 1989). Such allele-specific oligonucleotides are hybridized to PCR-amplified target DNA or a number of different mutations when the oligonucleotides are attached to the hybridizing membrane and hybridized with labeled target DNA.

Alternatively, allele specific amplification technology that depends on selective PCR amplification may be used. Oligonucleotide primers for specific amplifications may carry the mutation of interest in the center of the molecule (so that amplification depends on differential hybridization (Gibbs *et al.*, 1989)) or at the extreme 3'-terminus of one primer where, under appropriate conditions, mismatch can prevent, or reduce polymerase extension (Prosser, 1993). Novel restriction site in the region of the mutation may be introduced to create cleavage-based detection (Gasparini *et al.*, 1992). Certain amplification may also be performed using *Taq* ligase for amplification (Barany, 1991). In such cases, ligation occurs only if there is a perfect match at the 3'-terminus of the 5' sequence, allowing detection of a known mutation by scoring for amplification.

The described methods may be performed, for example, by using pre-packaged kits comprising at least one probe (nucleic acid or antibody) that may be conveniently used, for example, in clinical settings to diagnose patients exhibiting symptoms or family history of a disease or illness involving an AAP.

Furthermore, any cell type or tissue in which an AAP is expressed may be utilized in the prognostic assays described herein.

3. Pharmacogenomics

Agents, or modulators that have a stimulatory or inhibitory effect on AAP activity or expression, as identified by a screening assay can be administered to individuals to treat, prophylactically or therapeutically, disorders, including those associated with angiogenesis. In conjunction with such treatment, the pharmacogenomics (*i.e.*, the study of the relationship between a subject's genotype and the subject's response to a foreign modality, such as a food, compound or drug) may be considered. Metabolic differences of therapeutics can lead to severe toxicity or therapeutic failure by altering the relation between dose and blood concentration of the pharmacologically active drug. Thus, the pharmacogenomics of the individual permits the selection of effective agents (*e.g.*, drugs) for prophylactic or therapeutic treatments based on a consideration of the individual's genotype. Pharmacogenomics can further be used to determine appropriate dosages and

therapeutic regimens. Accordingly, the activity of an AAP, expression of an AAP nucleic acid, or an AAP mutation(s) in an individual can be determined to guide the selection of appropriate agent(s) for therapeutic or prophylactic treatment.

Pharmacogenomics deals with clinically significant hereditary variations in the response to modalities due to altered modality disposition and abnormal action in affected persons (Eichelbaum and Evert, 1996; Linder *et al.*, 1997). In general, two pharmacogenetic conditions can be differentiated: (1) genetic conditions transmitted as a single factor altering the interaction of a modality with the body (altered drug action) or (2) genetic conditions transmitted as single factors altering the way the body acts on a modality (altered drug metabolism). These pharmacogenetic conditions can occur either as rare defects or as nucleic acid polymorphisms. For example, glucose-6-phosphate dehydrogenase (G6PD) deficiency is a common inherited enzymopathy in which the main clinical complication is hemolysis after ingestion of oxidant drugs (anti-malarials, sulfonamides, analgesics, nitrofurans) and consumption of fava beans.

As an illustrative embodiment, the activity of drug metabolizing enzymes is a major determinant of both the intensity and duration of drug action. The discovery of genetic polymorphisms of drug metabolizing enzymes (*e.g.*, N-acetyltransferase 2 (NAT 2) and cytochrome P450 enzymes CYP2D6 and CYP2C19) explains the phenomena of some patients who show exaggerated drug response and/or serious toxicity after taking the standard and safe dose of a drug. These polymorphisms are expressed in two phenotypes in the population, the extensive metabolizer (EM) and poor metabolizer (PM). The prevalence of PM is different among different populations. For example, the *CYP2D6* gene is highly polymorphic and several mutations have been identified in PM, which all lead to the absence of functional CYP2D6. Poor metabolizers due to mutant *CYP2D6* and *CYP2C19* frequently experience exaggerated drug responses and side effects when they receive standard doses. If a metabolite is the active therapeutic moiety, PM shows no therapeutic response, as demonstrated for the analgesic effect of codeine mediated by its CYP2D6-formed metabolite morphine. At the other extreme are the so-called ultra-rapid metabolizers who are unresponsive to standard doses. Recently, the molecular basis of ultra-rapid metabolism has been identified to be due to CYP2D6 gene amplification.

The activity of an AAP, expression of an AAP nucleic acid, or mutation content of an AAP in an individual can be determined to select appropriate agent(s) for therapeutic or prophylactic treatment of the individual. In addition, pharmacogenetic studies can be used to apply genotyping of polymorphic alleles encoding drug-metabolizing enzymes to the identification of an individual's drug responsiveness phenotype. This knowledge, when applied to dosing or drug selection, can avoid adverse reactions or therapeutic failure and thus enhance therapeutic or prophylactic efficiency when treating a subject with an AAP modulator, such as a modulator identified by one of the described exemplary screening assays.

4. *Monitoring effects during clinical trials*

Monitoring the influence of agents (*e.g.*, drugs, compounds) on the expression or activity of an AAP (*e.g.*, the ability to modulate angiogenesis) can be applied not only in basic drug screening, but also in clinical trials. For example, the effectiveness of an agent determined by a screening assay to increase an AAP expression, protein levels, or up-regulate an AAP's activity can be monitored in clinical trials of subjects exhibiting decreased AAP expression, protein levels, or down-regulated AAP activity. Alternatively, the effectiveness of an agent determined to decrease an AAP expression, protein levels, or down-regulate an AAP's activity, can be monitored in clinical trials of subjects exhibiting increased the AAP expression, protein levels, or up-regulated AAP activity. In such clinical trials, the expression or activity of the AAP and, preferably, other genes that have been implicated in, for example, angiogenesis can be used as a "read out" or markers for a particular cell's responsiveness.

For example, genes, including an AAP, that are modulated in cells by treatment with a modality (*e.g.*, food, compound, drug or small molecule) can be identified. To study the effect of agents on angiogenesis, for example, in a clinical trial, cells can be isolated and RNA prepared and analyzed for the levels of expression of an AAP and other genes implicated in the disorder. The gene expression pattern can be quantified by Northern blot analysis, nuclear run-on or RT-PCR experiments, or by measuring the amount of protein, or by measuring the activity level of the AAP or other gene products. In this manner, the gene expression pattern itself can serve as a marker, indicative of the cellular physiological response to the agent. Accordingly, this response state may be

determined before, and at various points during, treatment of the individual with the agent.

The invention provides a method for monitoring the effectiveness of treatment of a subject with an agent (*e.g.*, an agonist, antagonist, protein, peptide, peptidomimetic, nucleic acid, small molecule, food or other drug candidate identified by the screening assays described herein) comprising the steps of (1) obtaining a pre-administration sample from a subject; (2) detecting the level of expression of an AAP, mRNA, or genomic DNA in the preadministration sample; (3) obtaining one or more post-administration samples from the subject; (4) detecting the level of expression or activity of the AAP, mRNA, or genomic DNA in the post-administration samples; (5) comparing the level of expression or activity of the AAP, mRNA, or genomic DNA in the pre-administration sample with the AAP, mRNA, or genomic DNA in the post administration sample or samples; and (6) altering the administration of the agent to the subject accordingly. For example, increased administration of the agent may be desirable to increase the expression or activity of the AAP to higher levels than detected, *i.e.*, to increase the effectiveness of the agent. Alternatively, decreased administration of the agent may be desirable to decrease expression or activity of the AAP to lower levels than detected, *i.e.*, to decrease the effectiveness of the agent.

5. *Methods of treatment*

The invention provides for both prophylactic and therapeutic methods of treating a subject at risk of (or susceptible to) a disorder or having a disorder associated with aberrant AAP expression or activity. Furthermore, these same methods of treatment may be used to induce or inhibit angiogenesis, by changing the level of expression or activity of an AAP.

6. *Disease and disorders*

Diseases and disorders that are characterized by increased AAP levels or biological activity may be treated with therapeutics that antagonize (*i.e.*, reduce or inhibit) activity. Antagonists may be administered in a therapeutic or prophylactic manner. Therapeutics that may be used include: (1) AAP peptides, or analogs, derivatives, fragments or homologs thereof; (2) Abs to an AAP peptide; (3) AAP nucleic acids; (4) administration of antisense nucleic acid and nucleic acids that are “dysfunctional” (*i.e.*, due to a heterologous insertion within the coding sequences) that are used to eliminate

endogenous function of by homologous recombination (Capecchi, 1989); or (5) modulators (*i.e.*, inhibitors, agonists and antagonists, including additional peptide mimetic of the invention or Abs specific to an AAP) that alter the interaction between an AAP and its binding partner.

5 Diseases and disorders that are characterized by decreased AAP levels or biological activity may be treated with therapeutics that increase (*i.e.*, are agonists to) activity. Therapeutics that upregulate activity may be administered therapeutically or prophylactically. Therapeutics that may be used include peptides, or analogs, derivatives, fragments or homologs thereof; or an agonist that increases bioavailability.

0 Increased or decreased levels can be readily detected by quantifying peptide and/or RNA, by obtaining a patient tissue sample (*e.g.*, from biopsy tissue) and assaying *in vitro* for RNA or peptide levels, structure and/or activity of the expressed peptides (or AAP mRNAs). Methods include, but are not limited to, immunoassays (*e.g.*, by Western blot analysis, immunoprecipitation followed by sodium dodecyl sulfate (SDS) polyacrylamide gel electrophoresis, immunocytochemistry, *etc.*) and/or hybridization assays to detect expression of mRNAs (*e.g.*, Northern assays, dot blots, *in situ* hybridization, and the like).

5 7. Prophylactic methods

0 The invention provides a method for preventing, in a subject, a disease or condition associated with an aberrant AAP expression or activity, by administering an agent that modulates an AAP expression or at least one AAP activity. Subjects at risk for a disease that is caused or contributed to by an aberrant AAP expression or activity can be identified by, for example, any or a combination of diagnostic or prognostic assays. Administration of a prophylactic agent can occur prior to the manifestation of symptoms characteristic of the AAP aberrancy, such that a disease or disorder is prevented or, alternatively, delayed in its progression. Depending on the type of AAP aberrancy, for example, an AAP agonist or AAP antagonist can be used to treat the subject. The appropriate agent can be determined based on screening assays.

5 8. Therapeutic methods

Another aspect of the invention pertains to methods of modulating an AAP expression or activity for therapeutic purposes. The modulatory method of the invention involves contacting a cell with an agent that modulates one or more of the activities of

AAP activity associated with the cell. An agent that modulates AAP activity can be a nucleic acid or a protein, a naturally occurring cognate ligand of an AAP, a peptide, an AAP peptidomimetic, or other small molecule. The agent may stimulate AAP activity. Examples of such stimulatory agents include an active AAP and an AAP nucleic acid molecule that has been introduced into the cell. In another embodiment, the agent inhibits AAP activity. Examples of inhibitory agents include antisense AAP nucleic acids and anti-AAP Abs. Modulatory methods can be performed *in vitro* (e.g., by culturing the cell with the agent) or, alternatively, *in vivo* (e.g., by administering the agent to a subject). As such, the invention provides methods of treating an individual afflicted with a disease or disorder characterized by aberrant expression or activity of an AAP or nucleic acid molecule. In one embodiment, the method involves administering an agent (e.g., an agent identified by a screening assay), or combination of agents that modulates (e.g., up-regulates or down-regulates) AAP expression or activity. In another embodiment, the method involves administering an AAP or nucleic acid molecule as therapy to compensate for reduced or aberrant AAP expression or activity.

Stimulation of AAP activity is desirable in situations in which AAP is abnormally down-regulated and/or in which increased AAP activity is likely to have a beneficial effect. One example of such a situation is where a subject has a disorder characterized by aberrant angiogenesis (e.g., cancer).

9. *Determination of the biological effect of the therapeutic*

Suitable *in vitro* or *in vivo* assays can be performed to determine the effect of a specific therapeutic and whether its administration is indicated for treatment of the affected tissue.

In various specific embodiments, *in vitro* assays may be performed with representative cells of the type(s) involved in the patient's disorder, to determine if a given therapeutic exerts the desired effect upon the cell type(s). Modalities for use in therapy may be tested in suitable animal model systems including, but not limited to rats, mice, chicken, cows, monkeys, rabbits, and the like, prior to testing in human subjects. Similarly, for *in vivo* testing, any of the animal model system known in the art may be used prior to administration to human subjects.

10. *Prophylactic and therapeutic uses of the compositions of the invention*

AAP nucleic acids and proteins are useful in potential prophylactic and therapeutic applications implicated in a variety of disorders including, but not limited to those related to angiogenesis.

As an example, a cDNA encoding an AAP may be useful in gene therapy, and the protein may be useful when administered to a subject in need thereof.

AAP nucleic acids, or fragments thereof, may also be useful in diagnostic applications, wherein the presence or amount of the nucleic acid or the protein is to be assessed. A further use could be as an anti-bacterial molecule (*i.e.*, some peptides have been found to possess anti-bacterial properties). These materials are further useful in the generation of Abs that immunospecifically bind to the novel substances of the invention for use in therapeutic or diagnostic methods.

The following example is meant to not be limiting.

EXAMPLE

Identification of genes differentially-regulated

A comprehensive mRNA profiling technique (GeneCalling) was used to determine differential gene expression profiles of human endothelial cells undergoing differentiation into tube-like structures (Kahn *et al.*, 2000). To confirm the expression data from GeneCalling, independent experiments were undertaken that used gene-specific PCR oligonucleotide primer pairs and an oligonucleotide probe labeled with a fluorescent dye at the 5' end and quencher fluorescent dye at the 3' end.. Total RNA (50 ng) was added to a 50 µl RT-PCR mixture and run.

The following data were collected:

hEF G	collagen gel 24 hr versus 4h	4.5 fold upregulated
hTRG	collagen gel 24 hr versus 4h	3.5 fold upregulated
KLP	collagen gel 24 hr versus 4h	3.5 fold upregulated
myosin X	collagen gel 24 hr versus 4h	3.5 fold upregulated
NHR	collagen gel 24 hr versus 4h	7.3 fold downregulated
HBAZF	collagen gel 24 hr versus 4h	2.1 fold upregulated

EQUIVALENTS

Although particular embodiments have been disclosed herein in detail, this has been done by way of example for purposes of illustration only, and is not intended to be limiting with respect to the scope of the appended claims that follow. In particular, it is contemplated by the inventors that various substitutions, alterations, and modifications may be made to the invention without departing from the spirit and scope of the invention as defined by the claims. The choice of nucleic acid starting material, clone of interest, or library type is believed to be a matter of routine for a person of ordinary skill in the art with knowledge of the embodiments described herein. Other aspects, advantages, and modifications considered within the scope of the following claims.

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All publications and patents mentioned in the above specification are herein incorporated by reference.